



TOXICOLOGICAL REVIEW

OF

**Cerium Oxide and Cerium
Compounds**

(CAS No. 1306-38-3)

**In Support of Summary Information on the
Integrated Risk Information System (IRIS)**

May 2008

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AND CERIUM COMPOUNDS (CAS NO. 1306-38-3)**

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LIST OF ABBREVIATIONS AND ACRONYMS

1		
2		
3		
4	ALT	alanine aminotransferase
5	AST	aspartate aminotransferase
6	BMC	benchmark concentration
7	BMD	benchmark dose
8	CASRN	Chemical Abstract Service Registry Number
9	CFU	colony-forming unit
10	CI	confidence interval
11	COH	coumarin 7-hydroxylase
12	CT	computer tomography
13	CYP	cytochrome P450 (in connection with isozyme abbreviations)
14	CYP450	cytochrome P450
15	DAF	dosimetric adjustment factor
16	DCF	dichlorodihydrofluorescein diacetate
17	DPO	dendriform pulmonary ossification
18	EDTA	ethylenediamine tetraacetic acid
19	EGTA	ethylene glycol bis(2-aminoethylether)tetraacetic acid
20	EPA	Environmental Protection Agency
21	GD	gestation day
22	GI	gastrointestinal
23	GSD	geometric standard deviation
24	GSH	glutathione
25	HEC	human equivalent concentration
26	HEI	Health Effects Institute
27	i.v.	intravenous
28	IRIS	Integrated Risk Information System
29	LC₅₀	median lethal concentration
30	LD_x	dose that kills X% of animals
31	LDH	lactate dehydrogenase
32	LOAEL	lowest-observed-adverse-effect level
33	MCP	monocyte chemoattractant protein
34	MDA	malodialdehyde
35	MMAD	mass median aerodynamic diameter
36	MT	metallothionein
37	NOAEL	no-observed-adverse-effect level
38	NTP	National Toxicology Program
39	OCT	ornithine-carbamyl transferase
40	PAM	pulmonary alveolar macrophage
41	PBTK	physiologically based toxicokinetic
42	PCNA	proliferating cell nuclear antigen
43	PND	postnatal day
44	RDDR	regional deposited dose ratio
45	RfC	reference concentration
46	RfD	reference dose

- 1 **SDH** sorbitol dehydrogenase
- 2 **SOD** superoxide dismutase
- 3 **UF** uncertainty factor
- 4

FOREWORD

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4 The purpose of this Toxicological Review is to provide scientific support and rationale
5 for the hazard and dose-response assessment in IRIS pertaining to chronic exposure to cerium
6 oxide and cerium compounds. It is not intended to be a comprehensive treatise on the chemical
7 or toxicological nature of cerium oxide and cerium compounds.

8 The intent of Section 6, *Major Conclusions in the Characterization of Hazard and Dose*
9 *Response*, is to present the major conclusions reached in the derivation of the reference dose,
10 reference concentration and cancer assessment, where applicable, and to characterize the overall
11 confidence in the quantitative and qualitative aspects of hazard and dose response by addressing
12 the quality of the data and related uncertainties. The discussion is intended to convey the
13 limitations of the assessment and to aid and guide the risk assessor in the ensuing steps of the
14 risk assessment process.

15 For other general information about this assessment or other questions relating to IRIS,
16 the reader is referred to EPA's IRIS Hotline at (202) 566-1676 (phone), (202) 566-1749 (fax), or
17 hotline.iris@epa.gov (email address).

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1. INTRODUCTION

This document presents background information and justification for the Integrated Risk Information System (IRIS) Summary of the hazard and dose-response assessment of cerium oxide and cerium compounds. IRIS Summaries may include oral reference dose (RfD) and inhalation reference concentration (RfC) values for chronic and less-than-lifetime exposure durations, and a carcinogenicity assessment.

The RfD and RfC provide quantitative information for use in risk assessments for health effects known or assumed to be produced through a nonlinear (presumed threshold) mode of action. The RfD (expressed in units of mg/kg-day) is defined as an estimate (with uncertainty spanning perhaps an order of magnitude) of a daily exposure to the human population (including sensitive subgroups) that is likely to be without an appreciable risk of deleterious effects during a lifetime. The inhalation RfC (expressed in units of mg/m³) is analogous to the oral RfD, but provides a continuous inhalation exposure estimate. The inhalation RfC considers toxic effects for both the respiratory system (portal of entry) and for effects peripheral to the respiratory system (extrarespiratory or systemic effects). Reference values may also be derived for acute (≤ 24 hours), short-term (> 24 hours up to 30 days), and subchronic (30 days up to 10% of average lifetime) exposure durations, all of which are derived based on an assumption of continuous exposure throughout the duration specified.

The carcinogenicity assessment provides information on the carcinogenic hazard potential of the substance in question and quantitative estimates of risk from oral and inhalation exposure. The information includes a weight-of-evidence judgment of the likelihood that the agent is a human carcinogen and the conditions under which the carcinogenic effects may be expressed. Quantitative risk estimates are derived from the application of a low-dose extrapolation procedure. The oral slope factor is an upper bound on the estimate of risk per mg/kg-day of oral exposure. Similarly, an inhalation unit risk is an upper bound on the estimate of risk per $\mu\text{g}/\text{m}^3$ air breathed.

Development of these hazard identification and dose-response assessments for cerium oxide and cerium compounds has followed the general guidelines for risk assessment as set forth by the National Research Council (1983). EPA guidelines and Risk Assessment Forum Technical Panel Reports that were used in the development of this assessment include the following: *Guidelines for Mutagenicity Risk Assessment* (U.S. EPA, 1986b), *Guidelines for Developmental Toxicity Risk Assessment* (U.S. EPA, 1991), *Guidelines for Reproductive Toxicity Risk Assessment* (U.S. EPA, 1996b), *Guidelines for Neurotoxicity Risk Assessment* (U.S. EPA, 1998a), *Guidelines for Carcinogen Risk Assessment* (U.S. EPA, 2005a), *Supplemental Guidance for Assessing Susceptibility from Early-Life Exposure to Carcinogens* (U.S. EPA, 2005b),

1 *Recommendations for and Documentation of Biological Values for Use in Risk Assessment* (U.S.
2 EPA, 1988), (proposed) *Interim Policy for Particle Size and Limit Concentration Issues in*
3 *Inhalation Toxicity* (U.S. EPA, 1994a), *Methods for Derivation of Inhalation Reference*
4 *Concentrations and Application of Inhalation Dosimetry* (U.S. EPA, 1994b), *Use of the*
5 *Benchmark Dose Approach in Health Risk Assessment* (U.S. EPA, 1995), *Science Policy Council*
6 *Handbook: Peer Review* (U.S. EPA, 1998b, 2000a, 2005c), *Science Policy Council Handbook:*
7 *Risk Characterization* (U.S. EPA, 2000b), *Benchmark Dose Technical Guidance Document*
8 (U.S. EPA, 2000c), and *A Review of the Reference Dose and Reference Concentration Processes*
9 (U.S. EPA, 2002).

10 The literature search strategy employed for this compound was based on the Chemical
11 Abstract Service Registry Number (CASRN) and at least one common name. Any pertinent
12 scientific information submitted by the public to the IRIS Submission Desk was also considered
13 in the development of this document. The relevant literature was reviewed through June 2007.

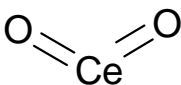
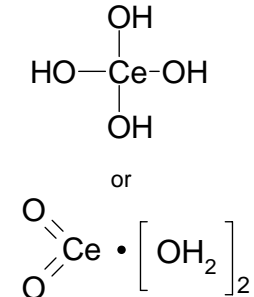
2. CHEMICAL AND PHYSICAL INFORMATION

Cerium is a member of the lanthanide series of metals and is the most abundant of the rare-earth elements in the earth's crust (average concentration of 50 ppm) (Hedrick, 2004). Elemental cerium is an iron-gray, ductile, malleable metal (O'Neil, 2001). Cerium metal is very reactive and is a strong oxidizing agent that is stabilized when associated with an oxygen ligand (Kilbourn, 2003). When present in compounds, cerium exists in both the trivalent state (Ce^{3+} , cerous) and the tetravalent state (Ce^{4+} , ceric) (Kilbourn, 2003; Reinhardt and Winkler, 2002). Chemical structures and selected chemical and physical properties of cerium and cerium compounds are listed in Table 2-1.

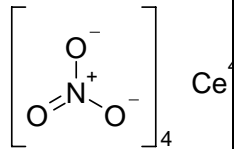
Cerium is found in nature along with other lanthanide elements in the minerals alanite, bastanite, monazite, cerite, and samarskite; however, only bastanite and monazite are important sources commercially (Lide, 2005; Kilbourn, 2003). Because of its unique stability in the tetravalent state (other lanthanides are stable in only the trivalent state), cerium can be separated out from the other rare-earth elements through oxidation (forming CeO_2) followed by variable solubility filtration (Reinhardt and Winkler, 2002). Cerium salts can be prepared by liquid-liquid extraction from rare-earth cerium-containing solutions. Cerium metal is prepared by reacting CeF_3 with an excess of calcium at approximately 900°C (Kilbourn, 2003). It can also be obtained by the fused-salt electrolysis of a mixture of cerium chlorides and fluorides (Reinhardt and Winkler, 2002).

Cerium is most heavily used in the form of mischmetal for metallurgical purposes (Kilbourn, 2003; Reinhardt and Winkler, 1996). Cerium is the major component of mischmetal (50–75% by weight for the most common grades), a commercial mixture of metallic light lanthanides prepared by the electrolysis of mixed lanthanide chlorides and fluorides obtained from bastanite or monazite (Kilbourn, 2003; Reinhardt and Winkler, 2002). Mischmetal reacts with the impurities found in metals to form solid compounds, thereby reducing the effect of these impurities on the properties of the metal (Reinhardt and Winkler, 2002). Mischmetal has been used in the manufacture of steel to improve shape control, reduce hot shortness, and increase heat and oxidation resistance. It can be added to cast iron to improve ductility, toughness, and microstructure. Mischmetal is also used in the manufacture of cerium-iron alloy lighter flints (Kilbourn, 2003; Reinhardt and Winkler, 2002).

Table 2-1. Physical properties of cerium and selected cerium compounds

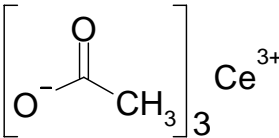
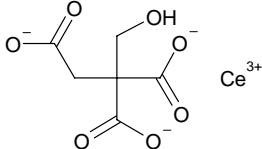
Name	Cerium	Cerium oxide	Hydrated cerium oxide
CASRN	7440-45-1	1306-38-3	12014-56-1 (hydroxide) 23322-64-7 (hydrate)
Synonyms		Cerium dioxide; ceria; cerium(IV) oxide	Hydrated ceric oxide; cerium hydrate; ceric hydroxide; cerium(IV) hydroxide; cerium perhydroxide; cerium tetrahydroxide
Structure			
Molecular weight	140.116	172.11	208.148
Molecular formula	Ce	CeO ₂	CeO ₂ ·2H ₂ O
Form	Iron-gray, ductile, malleable metal	Pale-yellow, heavy powder (white when pure); commercial product is brown	The hydroxide precipitate is amorphous and on drying, converts to hydrated ceric oxide; whitish powder when pure.
Melting point	798°C; boiling point = 3443°C	2400°C	Not available
Density	6.770 g/cm ³	7.65 g/cm ³	Not available
Water solubility	Decomposes slowly with cold water and rapidly with hot water	Insoluble in water	Insoluble in water
Other solubility	Soluble in dilute mineral acids	Insoluble in dilute acid	Soluble in concentrated mineral acid

1
2**Table 2-1 [continued]. Physical properties of cerium and selected cerium compounds**

Name	Cerium nitrate	Cerous chloride	Cerous fluoride
CASRN	13093-17-9	7790-86-5	7758-88-5
Synonyms	Cerium tetranitrate; nitric acid, cerium(4+) salt	Cerium(III) chloride; cerium trichloride	Cerium(III) fluoride; cerium trifluoride
Structure			
Molecular weight	388.136 9	246.48	197.11
Molecular formula	Ce(NO ₃) ₄	CeCl ₃	CeF ₃
Form	Not available	White crystals; fine powder	Hexagonal crystals or powder
Melting point	Not available	817°C	1430°C
Density	Not available	3.97 g/cm ³	6.157 g/cm ³
Water solubility	Not available	Soluble in water	Insoluble in water
Other solubility	Not available	Soluble in ethanol	Soluble in acids (monohydrate)

3

1
2**Table 2-1 [continued]. Physical properties of cerium and selected cerium compounds**

Name	Cerous acetate	Cerous citrate
CASRN	537-00-8	512-24-3
Synonyms	Cerium(III) acetate; cerium triacetate; acetic acid; cerium(3+) salt	Cerium citrate; cerium(III) citrate; cerium(3+) 2-hydroxypropane-1,2,3-tricarboxylate
Structure		
Molecular weight	317.251	329.219
Molecular formula	Ce(C ₂ H ₃ O ₂) ₃	C ₆ H ₈ O ₇ .Ce
Form	Not available	Not available
Melting point	Not available	Not available
Density	Not available	Not available
Water solubility	Not available	Not available
Other solubility	Not available	Not available

3
4

Sources: ChemIDplus (2006); Lide (2005); Lewis (2001); O'Neil (2001).

1 Exposure to commercially used cerium compounds is most likely through exposure to
 2 cerium (ceric) oxide (CeO₂). It is used either in the pure form or in a concentrate as a polishing
 3 agent for glass mirrors, plate glass, television tubes, ophthalmic lenses, and precision optics
 4 (Kilbourn, 2003; Reinhardt and Winkler, 2002). Cerium oxide is used as a glass constituent to
 5 prevent solarization and discoloration (especially in the faceplates of television screens)
 6 (Reinhardt and Winkler, 2002). Cerium oxide is also used in emission control systems in
 7 gasoline engines and as a diesel fuel-born catalyst to reduce particulate matter emissions (Health
 8 Effects Institute [HEI], 2001; Reinhardt and Winkler, 1996). Cerium nitrate has been used as a
 9 topical treatment for burn wounds (Monafo et al., 1976). Major uses for selected cerium
 10 compounds are listed in Table 2-2.

11
 12

Table 2-2. Major uses of selected cerium compounds

Name	CASRN	Use
Cerium oxide	1306-38-3	Polishing and decolorizing glass; opacifier in vitreous enamels and photochromic glasses; heat-resistant alloy coatings; as a cracking catalyst; as a catalyst for automobile emission control; in ceramic coatings; in phosphors; in cathodes; in capacitors; in semiconductors; in refractory oxides; gemstone polishing
Hydrated cerium oxide	23322-64-7	Production of cerium salts and cerium oxide
Cerous chloride	7790-86-5	In the manufacturing of cerium metal and cerium salts; catalyst for the polymerization of olefins
Cerous fluoride	7758-88-5	In the preparation of cerium metal; in arc carbons to increase brilliance

13
 14
 15

Sources: Kilbourn (2003); HEI (2001); Lewis (2001); O'Neil (2001); Wells and Wells (2001).

16 Cerium is not expected to exist in elemental form in the environment since it is a very
 17 reactive metal (Lewis, 2001). Cerium compounds are not expected to volatilize and will exist in
 18 the particulate form if released into the air. For cerium compounds that are soluble in water,
 19 Ce³⁺ would likely have a pKa close to La³⁺ (8.5) (Wulfsberg, 2000), which indicates that the
 20 hydrated Ce³⁺ ion ([Ce(H₂O)_n]³⁺) will remain in solution at environmental pHs (4–9). The
 21 hydrated Ce⁴⁺ ion ([Ce(H₂O)_n]⁴⁺) is expected to hydrolyze and polymerize at environmental pH
 22 (Cotton et al., 1999) and may precipitate out of solution. In general, metal cations in solution are
 23 attracted to the surfaces of soil particles, and the extent of adsorption to soils will depend on the
 24 soil characteristics (e.g., pH, mineral content, organic content) (Evans, 1989).

3. TOXICOKINETICS

Many of the studies of the toxicokinetics of cerium were conducted using radioactive cerium. Stable and radioactive cerium are expected to behave in a similar toxicokinetic manner and possess the same chemical properties. Radioactive cerium is a beta-emitter. As such, data from studies using either stable or radioactive cerium are presented below. Data characterizing the toxicokinetics of cerium compounds, such as cerium oxide, are discussed in order to inform the overall database.

3.1. ABSORPTION

3.1.1. Oral Exposure

Studies evaluating the absorption of cerium compounds following oral exposure in humans are not available.

In adult animals, cerium compounds are very poorly absorbed following oral exposure, while suckling animals exhibit higher absorption and retention of cerium in the gastrointestinal (GI) tissues. Observed absorption of radioactive cerium salts from the GI tract of adult rats ranged from 0.05% to less than 0.1% of the administered dose (Kostial et al., 1989b; Inaba and Lengemann, 1972; Shiraishi and Ichikawa, 1972). Suckling rats, however, absorbed 40–98% of administered dose, with the youngest rats retaining the largest percentage of the dose (Kostial et al., 1989a, b; Inaba and Lengemann, 1972).

Four litters of Sprague-Dawley rats (n = 7–9), age 0, 7, 14, or 26 days, were given a single dose of [¹⁴¹Ce]-ceric nitrate of unreported concentration by intragastric dosing (Inaba and Lengemann, 1972). Subsequent, periodic whole-body radioactivity measurements were taken immediately after dosing and periodically thereafter. In another experiment in this study, two litters of 1-day-old newborn rats were given single doses of [¹⁴¹Ce]-ceric nitrate, and one rat from each litter was sacrificed at 1, 3, 5, 7, and 10 days after dosing, after which radioactivity in the GI tract and whole body was measured and macro-autoradiographs of the GI tract were produced.

It was observed that on or about day 16 of life, rats began consuming a solid, grain-based diet and were completely weaned by day 26 (Inaba and Lengemann, 1972). In weanling rats (rats dosed at 26 days of age), only 0.04% of the administered radioactivity remained in the body by 3 days after dosing. However, radioactivity in newborns diminished more slowly, dropping from 98% of administered dose on day 1 to 29% on day 16. The GI tract accounted for nearly 100% of whole-body retention on day 1 and 93% on day 16. After the onset of weaning in the suckling rats dosed as newborns, whole-body radioactivity fell to 3% of administered dose by day 24, of which only 17% was measured in the GI tract. Autoradiographs of the GI tract from the two litters dosed as 1-day-old newborns suggest rapid transit of radioactivity to the lower

1 small intestine 1 day after exposure. Autoradiography of rats 5 days after dosing (6 days old)
2 suggests that the intestinal radioactivity is restricted to the upper two-thirds of the epithelial villi,
3 which the study authors associated with cerium concentration in the vacuoles.

4 Kostial et al. (1989b) administered single oral doses of an unreported concentration of
5 [^{141}Ce]-cerous chloride by intragastric dosing to 6-day-old and 6–8-week-old rats (strain
6 unreported) to investigate whether distribution of cerium (and other metals) in the GI tract
7 differed in suckling versus mature rats. Six-day-old rats were measured for whole-body and gut
8 radioactivity 2, 4, 6, and 12 days after dosing, while adult rats were measured 2 days after
9 dosing. Orally administered cerium was more readily retained in the whole body, gut, and
10 carcass of suckling rats than in older rats. The ileum was the main site for cerium accumulation
11 in the suckling rats following oral administration, while the stomach and large intestine were the
12 main sites for cerium accumulation in the 6–8-week-old rats. Whole-body radioactivity 6 days
13 postexposure in 2-week-old suckling rats orally dosed with [^{141}Ce] was 40% of the administered
14 dose, of which 95% was found in the gut. However, whole-body radioactivity 6 days
15 postexposure in adult (6–8-week-old) rats was only 0.05%, of which 66% was found in the gut,
16 mostly in the stomach and cecum.

17 Shiraishi and Ichikawa (1972) administered single oral doses of an unreported
18 concentration of [^{144}Ce]-cerous chloride by intragastric dosing to 0-, 7-, 14-, and 21-day-old
19 juvenile and 100-day-old adult Wistar rats. Cortisone acetate, which alters the morphology of
20 the absorptive epithelium of the small intestine, was injected to a group of 7-day-old rats 4 days
21 prior to [^{144}Ce]-cerous chloride exposure to observe its effect on whole-body retention.
22 Periodically (up to 70 days after dosing), small groups of rats from all age groups were sacrificed
23 and measured for radioactivity in the whole-body, gut, and various excised tissues. The decrease
24 in retention of whole-body radioactivity among suckling rats dosed at 0, 7, and 14 days after
25 birth was approximately 11, 6.5, and 1.5%, respectively, of the administered dose. Weanling and
26 adult rats exhibited a rapid decrease in whole-body radioactivity through 10 days following
27 dosing. At 2 weeks after dosing, the 21- and 100-day-old adults had whole-body radioactivity of
28 0.08 and 0.018%, respectively, of the administered dose. The intestinal content accounted for
29 most of the retained radioactivity in neonates until the age of weaning. Cortisone acetate
30 treatment resulted in a more rapid loss of the oral dose from young rats, although whether this is
31 due to lower uptake by intestinal cells or a more rapid release to feces is not presently clear.
32 This investigation demonstrated that the whole-body retention of cerium by suckling rats was
33 greater than the retention by weanling and adult rats, and this increased retention by suckling rats
34 may be due to increased pinocytotic activity in the absorptive epithelium of sucklings (Shiraishi
35 and Ichikawa, 1972).

36 Yorkshire piglets treated with [^{144}Ce]-cerous chloride by gavage on the first or fourth day
37 after birth and sacrificed 4 or 18 or 3 or 21 days, respectively, after dosing absorbed 2.5–8% of

1 the administered dose (Mraz and Eisele, 1977). Absorption was threefold greater in piglets
2 treated at 1 day of age versus those treated at 4 days of age. Body content of cerium did not
3 differ significantly between piglets sacrificed at the earlier dates and those sacrificed later,
4 indicating that absorption was almost complete within 3–4 days.

5 Eisele et al. (1980) gave single gavage doses of either [¹⁴⁴Ce]-cerous chloride or
6 [¹⁴⁴Ce]-cerous citrate (concentration unreported) to 0–6- or 6–24-hour-old C₃H mice and
7 Sprague-Dawley rats and to 6–24-hour-old Yorkshire piglets. Radioactivity levels were
8 measured in the GI tract and other tissues, including the remaining carcass, at days 1, 5, 7, 9, 12,
9 15, 17, 19, and 21. In the 0–6-hour-old mice, a high of 31% of the administered dose of cerium
10 chloride was retained in the body 9 days following exposure. At 21 days post administration, the
11 amount of pooled cerium citrate and chloride retained was approximately 25%. The mice dosed
12 0–6 hours after birth retained more cerium in the GI tract throughout the 21-day observation
13 period than the mice dosed 6–24 hours after birth. The 0–6- and 6–24-hour-old rats exhibited
14 absorption of 9–10% of administered cerium 21 days after exposure. In the Yorkshire piglets,
15 the absorbed dose did not differ significantly over the 21-day observation period.

16 Two studies in adult rats also reported low absorption of cerium following oral
17 exposures. Durbin et al. (1956) administered single [¹⁴⁴Ce]-ceric nitrate (unreported
18 concentrations) intramuscular and intragastric doses to adult female Sprague-Dawley rats. The
19 rats dosed intramuscularly were sacrificed at post-administration days 1, 4, 64, and 256, while
20 the intragastric-dosed rats were sacrificed 4 days after administration. Less than 0.1% of the
21 intragastric-administered dose was absorbed from the GI tract.

22 Stineman et al. (1978) administered single intragastric [¹⁴¹Ce]-cerous chloride doses of
23 1000 mg/kg (lethal to 5% of the animals [LD₅]) and 1163 (LD₂₅) mg/kg to 6–8-week-old male
24 Swiss ICR mice and sacrificed them at 4 hours or at 1, 3, or 7 days later. No measurements were
25 made of whole-body radioactivity; however, 97–99% of radioactivity in the 12 sampled tissues
26 was found in the gut (stomach + duodenum).

27 In young (suckling) animals, cerium appears to be retained in intestinal cells, particularly
28 in the ileum, possibly resulting in a much greater absorption than in adult animals (Kostial et al.,
29 1989a, b; Inaba and Lengemann, 1972). However, cerium retained in intestinal cells has been
30 demonstrated to be unavailable systemically (Inaba and Lengemann, 1972). The high gut
31 retention of cerium in young animals may be associated with high pinocytotic activity of the
32 newborn intestinal cells (Kostial et al., 1989b). Glucocorticoids, such as methylprednisolone,
33 stimulate production of endogenous corticosteroids, which cause precocious gut closure
34 (decreased pinocytosis) and maturation of the metal absorptive process (Kargacin and Landeka,
35 1990). Administration of methylprednisolone in conjunction with artificial feeding of [¹⁴¹Ce]-
36 cerous chloride in cows' milk to 4-day-old suckling rats (strain not reported) resulted in a nearly
37 40-fold reduction in the amount of cerium detected in gut tissue (Kargacin and Landeka, 1990).

1 This finding suggests that the pinocytotic activity of the intestinal cells contributed to the
2 differences. Similarly, injection with cortisone acetate resulted in a more rapid loss of an oral
3 dose to feces of young rats (Shiraishi and Ichikawa, 1972), although whether this is due to lower
4 uptake by intestinal cells or a more rapid release is not presently clear.

6 **3.1.2. Inhalation Exposure**

7 Studies evaluating the deposition or absorption of cerium compounds following
8 inhalation exposure in humans are not available. However, cerium has been detected in the lung
9 tissue and alveolar macrophages of subjects believed to have been exposed to cerium
10 occupationally.

11 Case reports and a retrospective occupational investigation provide support for the
12 limited absorption of cerium deposited in the lung following inhalation exposure.
13 Transbronchial biopsies in a 60-year-old movie projectionist showed cerium concentrations of
14 11 µg/g wet weight after 12 years of exposure (Porru et al., 2001). McDonald et al. (1995)
15 demonstrated particulate material (diameter range from <1 µm to 5–10 µm) localized within lung
16 biopsy cells by using a scanning electron microscope. Analysis of bronchioalveolar lavage fluid
17 from a 58-year-old patient exposed to rare earth dusts and asbestos revealed cerium and
18 phosphorus in the alveolar macrophages (Pairon et al., 1995). The cerium particles accounted
19 for 70% of the particles observed in the lung tissue and were also identified in the interstitial
20 macrophages.

21 Microscopic examination of the tracheobronchial lymph nodes of a movie projectionist
22 of 25 years revealed grey granules in large macrophages, which were characterized as calcium
23 and rare earth elements (cerium, lanthanum, and neodymium) by energy-dispersive analysis of
24 X-rays (Waring and Watling, 1990). Energy-dispersive X-ray also characterized dark particles
25 (diameter of 1 to 6 µm) as cerium, unidentifiable by optical microscopy, in the bronchioalveolar
26 lavage of a 13-year photoengraver.

27 Lung tissues from a photoengraver exposed to smoke from cored carbon arc lamps for
28 46 years (Pietra et al., 1985) were found to have cerium concentrations 2,800–207,000 times
29 higher than those of urine, blood, or nails, suggesting that cerium particles in the lung are poorly
30 mobilized. The concentration of cerium in the lung and lymph nodes of a subject exposed to
31 cerium for 46 years as a photoengraver was 167 and 5 µg/g wet tissue weight, respectively, and
32 2,400- and 53-times higher, respectively, than the concentration in unexposed control subjects
33 (Vocaturro et al., 1983; Sabbioni et al., 1982).

34 Pairon et al. (1994) performed a retrospective evaluation of retention of cerium-
35 containing particles in the lungs of workers previously exposed to mineral dusts.
36 Bronchioalveolar lavage and lung tissue samples from mineral dust exposed workers and
37 controls were examined for cerium content. In the seven cases that were judged to have high

1 cerium particle retention (as defined by having lavage fluid or tissue cerium concentrations at
2 least 5 times higher than those of controls), time since last exposure ranged from present to 29
3 years, with one patient's exposure time not available.

4 Limited animal data are available regarding total deposition of cerium aerosols within the
5 respiratory tract. Thomas et al. (1972) exposed 4-month-old, mixed sex Holtzman rats to two
6 concentrations (unreported) of aerosolized 1.4 μm median aerodynamic diameter (GSD of 2.0)
7 [^{144}Ce]-ceric hydroxide for 10 minutes. Whole-body radioactivity measurements were used to
8 identify a 28% deposition rate of inhaled cerium aerosol. Boecker and Cuddihy (1974) reported
9 an average deposition of 71% in beagles exposed to [^{144}Ce]-cerous chloride for 4–10 minutes. In
10 both studies, cerium was deposited in the lungs with evidence of absorption from the lung
11 provided by the subsequent detection of cerium in the skeleton, liver, and kidneys.

12 13 **3.2. DISTRIBUTION**

14 **3.2.1. Oral Exposure**

15 Although cerium appears to be poorly absorbed from the GI tract, the bone and liver were
16 the organs with the highest cerium levels in rats following oral gavage of cerium chloride
17 (Shiraishi and Ichikawa, 1972). The concentration of cerium in the kidney, liver, lung, and
18 spleen of male ICR mice was significantly elevated relative to controls following 6 and 12 weeks
19 of oral exposure to 20 or 200 ppm cerium chloride (Kawagoe et al., 2005). The lung and spleen
20 contained the highest cerium concentrations in male ICR mice.

21 Manoubi et al. (1998) gave a single intragastric dose of stable cerium nitrate (20 mg/mL)
22 to Wistar rats. Three hours after dosing, cerium was found in the lysosomes of the duodenal
23 villosity but not in the liver or spleen. In 1-day-old Sprague-Dawley rats given a single
24 intragastric dose of [^{141}Ce]-ceric nitrate of unreported concentration, Inaba and Lengermann
25 (1972) found cerium to be localized centrally, likely in the vacuoles, within epithelial cells of the
26 small intestine.

27 Cerium is capable of crossing the placenta and entering the fetal circulation in mice, but
28 the amounts found in the uterus and placenta were generally less than 5% of the maternal body
29 burden and decreased rapidly with increased time after exposure (Naharin et al., 1969). Fetal
30 body burdens in rodents were generally less than 1% of the initial maternal body burden after
31 either injected or oral administration (Levack et al., 2002; Inaba et al., 1992; Naharin et al.,
32 1969). Small amounts of injected cerium were also found in the maternal milk of mice (Naharin
33 et al., 1974), although at a very small proportion (<0.01%) of the maternal body burden.

34 35 **3.2.2. Inhalation Exposure**

36 As poorly soluble particles, cerium particles behave like other airborne particles,
37 depositing within the respiratory tract based on aerodynamic character (Schulz et al., 2000).

1 Hamsters inhaling [¹⁴⁴Ce]-cerium oxide aerosols with particle activity median diameters of 0.11
2 and 0.06 μm exhibited lung burdens of 3.6% at 5 hours and 50% at 3 hours after exposure,
3 respectively, of initial body burden (Kanapilly and Luna, 1975).

4 Once deposited in the lung, insoluble cerium compounds may dissolve slowly, as
5 evidenced by the low percentage of cerium found in other tissues. In an investigation of the
6 toxicity of insoluble cerium, Hahn et al. (2001) exposed beagles to aerosolized [¹⁴⁴Ce]-fused
7 aluminosilicate particles for 2–48 minutes and collected a variety of tissues (tissues studied not
8 reported) at death (up to 6,205 days following exposure). Cerium was dissolved from the lung
9 into the systemic circulation and observed in the liver, skeleton, and tracheobronchial lymph
10 nodes. Hahn et al. (2001) found between 1.0 and 10% of the initial lung burden of [¹⁴⁴Ce]-fused
11 aluminosilicate particle aerosol in the liver and skeleton of beagles observed for 800 days
12 following 2–48-minute inhalation exposures. Hahn et al. (1999) also observed [¹⁴⁴Ce]-fused
13 aluminosilicate particle translocation to the tracheobronchial lymph nodes following acute
14 inhalation of the particles. Lundgren et al. (1992) exposed adult F344/Crl rats to [¹⁴⁴Ce]-cerium
15 oxide for 5–50 minutes or to bimonthly exposures of 25 minutes for 1 year; rats were sacrificed
16 at 1 hour and 3, 7, 14, 28, 56, 112, 224, 448, 560, and 672 days after exposure. The lungs, heart,
17 liver, spleen, kidney, and skeleton (remaining carcass) were measured for cerium. Cerium was
18 detected in the liver and skeleton in increasing percentages of body burden with respect to time,
19 while cerium was not detected in the spleen and kidneys.

20 More soluble forms of cerium (e.g., cerium citrate) may be systemically absorbed more
21 easily from the lung due to the increased solubility of the compound. Morgan et al. (1970)
22 exposed Swiss mice to aerosols of [¹⁴⁴Ce] in the form of chloride, citrate, or fused clay, with
23 activity median diameters from 1.3 to 2.75 μm, in unreported concentrations or durations. While
24 the initial body burden of all forms decreased rapidly during the first 2 weeks, likely due to
25 mucociliary elimination to the GI tract, the remaining lung burden for the relatively insoluble
26 fused clay remained higher than the chloride or citrate for the duration of the study (130 days).
27 Conversely, the liver burdens of the citrate and chloride forms remained higher than the fused
28 clay by about an order of magnitude. Further, as lung burdens of the chloride and citrate forms
29 decreased, the bone burdens of these forms increased. The bone burden for the fused clay form,
30 like the liver, was about an order of magnitude lower than the citrate or chloride forms.

31 Sturbaum et al. (1970) exposed 40 Chinese hamsters via the nose to [¹⁴⁴Ce]-cerous
32 chloride aerosol, activity median diameter of 0.83 μm and a GSD of 1.7, for 20 minutes and
33 sacrificed small groups (n = 4) at 2, 8, 16, 28, 64, 128, and 256 days after exposure. Whole-body
34 and tissue (types unreported) measurements of cerium radioactivity were made. The liver and
35 skeleton exhibited between 1 and 10% of the initial body burden throughout the post-
36 administration measurements, while the lung portion of the initial body burden diminished from
37 approximately 20 at two hours to <1% by study's end.

1 Cerium has been observed to be localized in the cell, particularly in the lysosomes, where
2 it is concentrated and precipitated in an insoluble form in association with phosphorus. Wistar
3 rats were exposed to stable cerium chloride aerosol, mean diameter of 0.1 μm , 5 hours/day for
4 either 5 days or 4 days/week for 4 weeks (Galle et al., 1992; Berry et al., 1989, 1988). Several
5 hours after exposure, cerium was deposited in the lysosomes of alveolar macrophages. The
6 cerium deposits appeared to be in the form of aggregates of fine granules or fine needles that
7 varied in length from 30 to 60 nm, with the longer needles resulting from the 4-week exposure.
8 Cerium was found in the lysosomal fraction of liver centrifugate collected from rats (strain
9 unreported) given an intravenous (i.v.) injection of 1.3 mg/kg [^{141}Ce]-cerous chloride (Wiener-
10 Schmuck et al., 1990). Cerium was also found in the lysosomes of the duodenal villosity, but not
11 in the liver or spleen of Wistar rats following intragastric dosing. Cerium was also localized
12 centrally, likely in the vacuoles, within epithelial cells of the small intestine of Sprague-Dawley
13 rats.

14

15 **3.3. METABOLISM**

16 As an element, cerium is neither created nor destroyed within the body. The particular
17 cerium compound (e.g., cerium chloride, cerium oxide) may be altered as a result of various
18 chemical reactions within the body, particularly dissolution, but data have not demonstrated a
19 change in the oxidation state of the cerium cation. Exposure to cerium has been shown to
20 change hepatic levels of some cytochrome (CYP) P450 isozymes in a species- and strain-
21 sensitive manner for mice. Salonpää et al. (1992) gave i.v. cerous chloride injections of 2 mg/kg
22 to adult DBA/2 and C57BL/6 mice and observed increases in expression of CYP2A4 and
23 CYP2A5 in the livers (2 and 3 days after dosing) and in the kidneys (6 hours and 1 day after
24 dosing) of D2 mice but not in B6 mice. Arvela et al. (1991) gave i.v. cerous chloride injections
25 of 0.5, 1, and 2 mg/kg to adult male DBA/2 and C57BL/6 mice and found a greater sensitivity to
26 increased CYP450 expression (isoform not reported) in DBA/2 and C57BL/6 mice 24 hours and
27 3 days after exposure, respectively. Conversely, Arvela and Karki (1971) observed a 50%
28 reduction, compared to controls, in CYP450 activity in adult Sprague-Dawley rats 3 days after a
29 single i.v. injection of 2 mg/kg cerous chloride. The effect of changes in CYP450 levels on the
30 toxicokinetics or toxicity of cerium, if any, is not known. In addition, the relatively high
31 intravenous bolus doses used in the available studies may not be relevant to oral or inhaled
32 exposure to cerium oxide.

33

34 **3.4. ELIMINATION**

35 Following inhalation exposure, the initial rapid elimination of cerium from the body is
36 due primarily to transport up the respiratory tract by the mucociliary escalator and eventual
37 swallowing of the material, as with other poorly soluble particles (Boecker and Cuddihy, 1974).

1 Initial short-term clearance rates range from 35 to 95% of initial cerium body burden, depending
2 on the species tested and length of clearance time investigated. Lundgren et al. (1992) exposed
3 adult F344/Crl rats to [¹⁴⁴Ce]-cerium oxide aerosol for 5–50 minutes, with clearance of
4 approximately 90% of the initial body burden by 7 days. Kanapilly and Luna (1975) exposed
5 hamsters to [¹⁴⁴Ce]-cerium oxide aerosols with particle activity median aerodynamic diameters
6 of 0.11 and 0.06 μm and observed decreases in initial body burden of 95 and 60%, respectively,
7 4 days after exposure. Differences in clearance rates may have been dependent on particle size
8 differences, with the smaller particles taking more time for elimination; however, the authors
9 also stated that the difference may have resulted from a leak in the inhalation chamber used for
10 the first dose group. Boecker and Cuddihy (1974) observed an early clearance of initial body
11 burden from 35–80% for individual dogs 4 days after exposure. Thomas et al. (1972) exposed
12 Holtzman rats to two concentrations (unreported) of aerosolized [¹⁴⁴Ce]-ceric hydroxide for
13 10 minutes and observed approximately 75–95% clearance of initial body burden within 2 weeks
14 of exposure (Thomas et al., 1972). Sturbaum et al. (1970) reported clearance of 80% of initial
15 cerium body burden by 7 days in Chinese hamsters exposed to [¹⁴⁴Ce]-cerous chloride aerosol
16 for 20 minutes. After the initial clearance of cerium particles from the upper respiratory tract,
17 pulmonary clearance is slower, with reported slow-phase clearance half-times ranging from 100
18 to 190 days in rodents (Lundgren et al., 1974; Thomas et al., 1972; Morgan et al., 1970;
19 Sturbaum et al., 1970). The slow-phase clearance was slightly faster in beagles, with an
20 estimated half-time of 63 days (Boecker and Cuddihy, 1974). Slow-phase clearance from the
21 lung is a combination of cerium dissolution and absorption (Morgan et al., 1970) and mechanical
22 clearance from the respiratory tract (Sturbaum et al., 1970).

23 Elimination of orally administered cerium has been shown to be age dependent in
24 animals, with suckling animals absorbing cerium into the GI tissues (Inaba and Lengemann,
25 1972). This cerium remains in the intestinal cells, is not available systemically, and is eventually
26 eliminated in the feces.

27 Although quantitative estimates of cerium elimination are rare, it appears that the primary
28 route of elimination for cerium, whether inhaled, ingested, or injected, is through the feces, with
29 small (generally <10%) amounts eliminated in the urine (Lustgarten et al., 1976; Durbin et al.,
30 1956). It has been suggested that the fecal excretion of systemically absorbed cerium is due to
31 elimination in the bile (Lustgarten et al., 1976), since hepatic clearance was due primarily to
32 biliary function.

33

34 **3.5. PHYSIOLOGICALLY BASED TOXICOKINETIC MODELS**

35 No physiologically based toxicokinetic (PBTK) models for cerium oxide or other cerium
36 compounds were located in the evaluated literature.

37

4. HAZARD IDENTIFICATION

4.1. STUDIES IN HUMANS—EPIDEMIOLOGY, CASE REPORTS, CLINICAL CONTROLS

4.1.1. Oral Exposure

An association between exposure to rare earth elements, cerium in particular, in food and the development of endomyocardial fibrosis has been suggested (Eapen, 1998; Kutty et al., 1996; Valiathan et al., 1989). Cerium levels were elevated in the endomyocardial tissue samples from patients who died of endomyocardial fibrosis compared with the tissues of controls who died of accidents or congenital heart disease (Valiathan et al., 1989). Although causality has not been conclusively demonstrated, a higher incidence of endomyocardial fibrosis has been reported among a population consuming tubers grown in a region of India with high soil cerium concentrations, compared to subjects consuming tubers grown in a soil with low cerium concentrations. Analysis of the geographic distribution of endemic endomyocardial fibrosis in India suggested a link to high cerium soil concentrations and possibly to magnesium deficiency during childhood (Kutty et al., 1996).

A case-control study was conducted by Gómez-Aracena et al. (2006) to investigate the role of chronic cerium exposure in coronary heart disease. Chronic cerium exposure was represented by toenail cerium concentrations, and the occurrence of first myocardial infarction was the characterization used for coronary heart disease. Odds ratios were calculated by comparing the four exposure groups of 111, 142, 171, and 257 µg/kg toenail cerium concentration to the control group. Gómez-Aracena et al. (2006) found an association between increased toenail cerium concentrations and the risk of first myocardial infarction, when controlling for confounding factors, such as smoking, body mass index history of hypertension, diabetes, family history of coronary heart disease, β-carotene, lycopene, α-tocopherol, selenium, mercury, and scandium. The odds ratio of first myocardial infarction in smokers with a toenail cerium concentration of 257 µg/kg and without adjusting for additional risk factors of myocardial infarction was 1.18 (95% confidence interval [CI]: 0.83–1.66), with a *p* value for trend of 0.020. In nonsmokers, the odds ratio of first myocardial infarction with toenail cerium concentrations of 169 and 227 µg/kg were 2.09 (95% CI: 1.05–4.16) and 2.81 (95% CI: 1.21–6.52), respectively, with a *p* value for trend of 0.011, when controlling for the confounding factors listed above. The results of Gómez-Aracena et al. (2006) suggest a relationship between chronic cerium exposure and increased risk of acute myocardial infarction, with the strongest association observed in nonsmokers.

1 **4.1.2. Inhalation Exposure**

2 Reports have been published describing numerous cases of workers who developed
3 adverse lung effects, such as interstitial lung disease or pneumoconiosis, associated with
4 accumulation of cerium in the lungs after prolonged occupational exposure to cerium fumes or
5 dust (Yoon et al., 2005; Porru et al., 2001; McDonald et al., 1995; Pairen et al., 1995, 1994;
6 Sulotto et al., 1986; Vogt et al., 1986; Pietra et al., 1985; Vocaturo et al., 1983; Sabbioni et al.,
7 1982; Husain et al., 1980; Kappenberger and Bühlmann, 1975; Heuck and Hoschek, 1968).
8 The workers in the above reports had been exposed to cerium for periods of 10–46 years, with
9 exposures most commonly due to fumes from carbon arc lamps. These lamps, widely used in
10 the past in the fields of cinematography and photoengraving, have a central core consisting of
11 approximately 46% cerium oxide and smaller amounts of other rare earth oxides, including
12 lanthanum, neodymium, praseodymium, and samarium (Waring and Watling, 1990). As the
13 core burns, it emits oxide, and, to a lesser extent, fluoride dusts of cerium and the other rare
14 earth elements. Cases of cerium pneumoconiosis not associated with carbon arc lamps all
15 involved exposure to cerium oxide, either during processing or peripheral to its use as an
16 abrasive to grind and polish lenses (McDonald et al., 1995). Exposure concentrations of
17 cerium were not quantified in any of these studies.

18 Dendriform pulmonary ossification (DPO), a rare condition characterized by branching,
19 bony spicules often containing marrow, which are found in the lung parenchyma and are
20 associated with pulmonary fibrosis, was observed in a 38-year-old man who worked as a
21 polisher in a crystal factory for 3 years (Yoon et al., 2005). Diffuse reticulonodular infiltrates in
22 the lung were observed in a chest radiograph, and a computer tomography (CT) scan showed
23 diffuse, tiny, circular, or beak-like densities with branching structures in the interlobular septum.
24 A CT scan with bone-setting showed branching, twig-like ossified masses in the right lower
25 lobe and a few dot-like ossifications in both lower lobes. The lung surface appeared irregular,
26 emphysematous, and mottled with anthracotic pigmentation during an open lung biopsy, with
27 several thorn-like hard materials in the lung parenchyma. Microscopic examination revealed
28 interstitial fibrosis, peripheral emphysema, multiple particles, and pneumonia. The particles
29 (0.1–0.3 μm) were determined to be cerium oxide and phosphates of cerium and lanthanum by
30 energy dispersive X-ray analysis, with particles of quartz, feldspar, mica, kaolinite, halloysite,
31 talc, and TiO_2 infrequently detected. Yoon et al. (2005) characterized this case study as the first
32 to present a case of DPO associated with pneumoconiosis caused by the inhalation of rare earth
33 metals.

34 A 60-year-old male who worked as a movie projectionist and was exposed to rare earth
35 dusts for 12 years presented with diffuse interstitial lung fibrosis, emphysema, and severe
36 obstructive impairment (Porru et al., 2001). An increase in the lung concentration of rare earth
37 elements was evident in the subject with the highest concentration for cerium, compared with the

1 five unexposed controls. Interstitial fibrosis accompanied by vascular thickening, reactive
2 alveolar macrophages, abundant macrophages in the air space, and moderate chronic interstitial
3 inflammation, along with small interstitial clumps of macrophages bearing scant deposits of
4 grayish-black pigment, was observed in a 68-year-old man who was employed as an optical lens
5 grinder for 35 years and smoked for 20 years (McDonald et al., 1995). Pairon et al. (1995)
6 identified particles containing cerium, lanthanum, and phosphorus in the alveolar macrophages
7 from a 58-year-old smoker with dyspnea who had been exposed to asbestos and rare earth dusts
8 as a crystal manufacturer, polisher, and movie projectionist from 1951–1967. Diffuse fibrosis of
9 interalveolar septa and perivascular hyalinized fibrosis was observed from the histologic analysis
10 (Pairon et al., 1995). Mild interstitial fibrosis, peribronchiolar fibrosis, and diffuse interstitial
11 fibrosis with emphysema were seen in a photoengraver or glass polisher, nonsmoking foundry
12 worker, and a movie projectionist and glass polisher, respectively (Pairon et al., 1994).

13 Deposits of carbon arc lamp fumes were evident in the macrophages in the
14 tracheobronchial lymph nodes of a 66-year-old smoking movie projectionist of 25 years,
15 although pneumoconiosis was not considered because respiratory symptoms and radiographic
16 and histologic changes were not apparent (Waring and Watling, 1990). Dark particles, identified
17 as cerium by energy dispersive X-ray analysis, were observed in lung tissue from a 48-year-old
18 smoker employed as a photoengraver for 13 years (Sulotto et al., 1986). A chest X-ray showed a
19 micronodular pattern extending to all lung fields although lung examinations were normal and
20 the patient did not experience respiratory impairment. Vogt et al. (1986) observed five
21 reproduction photographers, exposed to carbon arc fumes for more than a decade, with slowly
22 progressive respiratory function restriction, as well as interstitial lung fibrosis and the
23 accumulation of fine, granular dusts, characterized as rare earth minerals and primarily cerium,
24 in the lung tissue.

25 Pulmonary hypertension with increased vascular resistance was observed in a 58-year-old
26 man who worked in the photoengraving industry for 46 years (Vocaturro et al., 1983; Sabbioni et
27 al., 1982). Rare earth elements, primarily cerium, were observed by neutron activation analysis
28 in the lung and lymph node biopsies, and the concentrations of the elements in these tissues were
29 greater than those in control subjects (Pietra et al., 1985).

30 Profuse discrete nodular shadowing was present in the chest X-ray of a 34-year-old
31 employee at a glass rubbing polish plant, although pulmonary function appeared normal (Husain
32 et al., 1980). The subject refused histologic examination, and an analysis of the occupational
33 dust concentrations revealed high levels of cerium oxide (50%) and other rare earth oxides.

34 A 65-year-old man working in the photographic department of a printing plant
35 demonstrated inactive right apical infiltrates in a chest roentgenogram in 1948; in 1951 diffuse
36 spotty infiltrates were noted; in 1953 and 1959 the infiltrates were more pronounced in the
37 middle and lower fields; and in 1965 slight fibrosis of the surrounding tissue was evident, along

1 with a perifocal emphysema (Heuck and Hoschek, 1968). Heuck and Hoschek (1968) also
2 documented fibrosis and small infiltrates in the lung of a 53-year-old male exposed to carbon arc
3 lamp smoke in printing industries and infiltrates in a 67-year-old man, with chemotherapy-
4 treated tuberculosis in both upper lobes of the lung, who worked with carbon arc lamps for 26
5 years.

6 Collectively, the available studies show that the defining characteristic of cerium
7 pneumoconiosis is accumulation of cerium particles, as well as other rare earth particles, in the
8 lungs and lymphoreticular system. In most cases, the initial indication of disease was the
9 presence of diffuse interstitial or reticulonodular opacities in chest X-rays. Pulmonary function
10 in the affected workers varied from normal to severe restrictive impairment. In several cases,
11 thorium, a common impurity in rare earth minerals, and the naturally occurring radioisotopes of
12 the rareearth elements were quantified, but, in each case, they were found to be present in
13 quantities too small to produce any effect due to radiation. Exposure to silica, which is
14 fibrogenic, may have contributed to the effects observed in the cerium oxide and glass workers
15 but is not a factor for the workers exposed to carbon arc lamp dust, which includes most of the
16 workers found to have fibrosis. Two reviews of the available case studies concluded that there is
17 convincing evidence that accumulation of cerium and other rare earth metals in the lungs is
18 causally related to the development of pulmonary interstitial fibrosis in workers (McDonald et
19 al., 1995; Waring and Watling, 1990). The human data were inadequate to identify potentially
20 sensitive subgroups.

22 **4.2. SUBCHRONIC AND CHRONIC STUDIES AND CANCER BIOASSAYS IN** 23 **ANIMALS—ORAL AND INHALATION**

24 **4.2.1. Oral Exposure**

25 Kawagoe et al. (2005) studied the possible association between cerium exposure and
26 oxidative stress in the mouse liver. Groups of male ICR mice (four per dose group) were
27 administered a diet containing 0, 20, or 200 ppm cerium chloride ($CeCl_3$) for 6 or 12 weeks.
28 This corresponds to doses of approximately 2.6 or 26 mg Ce/kg-day, assuming a reference food
29 consumption of 0.005 kg/day and reference body weight of 0.022 kg (U.S. EPA, 1988).
30 Evaluations conducted at termination included cerium levels in organs (liver, spleen, kidney, and
31 lung), levels of glutathione (GSH) and metallothionein (MT) in liver, kidney, and lung, levels of
32 lipoperoxide in liver and plasma, superoxide dismutase (SOD) activity in blood and liver, as well
33 as cholesterol levels, triglyceride levels, and aspartate aminotransferase (AST) and alanine
34 aminotransferase (ALT) activity in serum. Treatment with cerium did not affect food
35 consumption or body weight but statistically significantly decreased lipoperoxide levels in
36 hepatic tissues (33% at 20 and 38% at 200 ppm, 6 weeks; 29% at 20 ppm, 12 weeks), increased
37 liver GSH levels (200 ppm, 12 weeks) and liver MT activity (20 ppm, 12 weeks; 200 ppm,

1 6 weeks), and decreased plasma SOD activity (20 and 200 ppm, 6 weeks). Cerium in the kidney,
2 liver, lung, and spleen was statistically significantly elevated relative to controls in the organs of
3 mice in the 200 ppm group at 6 and 12 weeks, with the lung and spleen containing higher cerium
4 concentrations. Pathological alterations were not detected in the kidney, liver, lung, or spleen by
5 microscopic observation. According to Kawagoe et al. (2005), the increases in hepatic GSH and
6 MT activity represent a response to cerium-induced oxidative stress. It is unknown whether the
7 decrease in hepatic lipoperoxide is a consequence of the increase in GSH and MT. The study
8 authors suggest that the endpoints showing changes as a result of cerium exposure in this study
9 are indicators of reactive oxygen species generation in the liver.

10 A study (Cheng et al., 2000) in male Wistar rats (six per dose group) dosed orally (further
11 details not reported) with 0, 0.2, 2.0, or 20 mg cerium chloride/kg-day (0.1, 1.1, or 11.4 mg
12 Ce/kg-day) for up to 105 days investigated the effects of CeCl₃ on the structure and oxygen
13 affinity of hemoglobin in vivo. The highest dose, 20 mg/kg-day, produced a slight increase of
14 hemoglobin content in the erythrocytes after 40 days of treatment, with an even greater increase
15 in hemoglobin content after 80 days. The effect on the oxygen affinity of the hemoglobin was
16 demonstrated by oxygen saturation curves for the dosed rats and control rats. Hemoglobin in
17 cerium-treated rats also exhibited altered oxygen affinity up to 80 days of exposure,
18 demonstrated by increased affinity up to 10 mm Hg and a double sigmoidal curve for rats treated
19 for 40 days and increased affinity above 20 mm Hg for rats treated for 80 days. A significant
20 change was not observed at 0.2 mg/kg-day for 105 days of exposure. These results suggest that
21 the oxygen affinity of hemoglobin increases following long-term oral exposure to CeCl₃. When
22 the 80-day cerium feeding period was followed by 15 days of cerium-free exposure, the oxygen
23 affinity did not recover to normal levels. The investigators attributed the altered oxygen affinity
24 up to 80 days of exposure to conformational changes of hemoglobin, hydrolysis of hemoglobin-
25 bound diphosphoglyceric acid, and the partial oxidation of heme-Fe(II) to heme-Fe(III).

26 Kartha et al. (1998) examined the effects of cerium chloride, administered in the drinking
27 water with other rare earth chlorides, on the heart of New Zealand white rabbits (10/group) fed
28 diets with normal or restricted magnesium for 6 months. The rabbits were randomly distributed
29 into 4 groups with a male to female ratio of 1:1 within each group. Group 1 was exposed to a
30 magnesium-sufficient diet only, group 2 was exposed to a magnesium-sufficient diet and cerium,
31 group 3 was exposed to a magnesium-restricted diet, and group 4 was exposed to a magnesium-
32 restricted diet and cerium. The drinking water for the rare earth chloride-exposed rabbits was
33 adulterated with 1 g/L of rare earth chloride of which 56.6% was cerium (lanthanum 11.5%;
34 praseodymium and neodymium 14.5%; samarium 2.6%). Histologic evaluation of the heart at
35 the end of the study showed no cardiac lesions in the groups fed the normal magnesium diet,
36 regardless of whether they consumed water with or without cerium. Cardiac lesions were
37 evident in 6/10 rabbits from group 3, the magnesium-restricted diet, and in 9/10 rabbits from

1 group 4, the magnesium-restricted diet and cerium chloride-exposed group. Rabbits fed the
2 magnesium-restricted diets, treated with or without cerium, showed endocardial, subendocardial,
3 interstitial, and perivascular fibrosis. The lesions were more severe in those with cerium added
4 to the drinking water (group 4). The results suggest that cerium may intensify the effect of
5 magnesium deficiency on heart tissue. A no-observed-adverse-effect level (NOAEL) or lowest-
6 observed-adverse-effect level (LOAEL) for cerium cannot be established for cerium chloride in
7 this study because the cardiac lesions were observed in rabbits that were fed a magnesium-
8 restricted diet.

9 The only chronic oral exposure study to stable cerium was reported by Kumar et al.
10 (1996), who exposed mixed-sex (male to female ratio of 1:1) groups (n = 5–9) of Sprague-
11 Dawley rats to cerium chloride and a magnesium-sufficient or magnesium-deficient diet for
12 13 months. The rats were randomly distributed into four groups, with groups 1 and 2 fed a
13 magnesium-sufficient diet and groups 3 and 4 fed a magnesium-deficient diet. The rats in group
14 1 and 3 were exposed to 0 ppm cerium chloride, while the rats in groups 2 and 4 were exposed to
15 35 ppm cerium chloride in the drinking water. This cerium chloride exposure corresponds to a
16 concentration of 19 mg/L, which in turn, assuming a reference water consumption of 0.046 and
17 0.038 L/day and reference body weight of 0.523 and 0.338 kg (U.S. EPA, 1988) in males and
18 females, respectively, corresponds to an average daily dose in males of 1.7 mg/kg-day and in
19 females of 2.1 mg/kg-day. At 13 months, the animals were sacrificed and cardiac tissue was
20 collected for elemental analysis and histology. No statistically significant changes in serum or
21 cardiac levels of magnesium or calcium were reported. Cerium levels in cardiac tissue were
22 statistically significantly elevated in group 4. Cerium-treated animals had a significantly greater
23 level of collagen in cardiac tissue, relative to group 1, with an enhanced effect in animals fed a
24 magnesium-deficient diet (groups 3 and 4). No other endpoints were evaluated. This study was
25 not adequate to identify a NOAEL or LOAEL due to the limited number of evaluated endpoints
26 and the investigation of a single dose group.

28 **4.2.2. Inhalation Exposure**

29 No chronic inhalation studies on cerium toxicity are available. However, the National
30 Toxicology Program (NTP) is considering an evaluation of the chronic inhalation toxicity of
31 cerium oxide.

32 A subchronic inhalation study using cerium oxide (CeO₂; ceric oxide) was conducted in
33 7-week-old Sprague-Dawley rats (BRL, 1994). Cerium oxide is the form of cerium typically
34 encountered in industrial exposures (Reinhardt and Winkler, 1986). This study is an
35 unpublished study; accordingly, it was externally peer reviewed by EPA in August 2006
36 (external peer review report available at www.epa.gov/iris).

37 Groups of 15 male and 15 female Sprague-Dawley CD rats were given nose-only

1 exposure to a dry powder aerosol (mass median aerodynamic diameter [MMAD] = 1.8–2.2 μm ,
2 geometric standard deviation [GSD] = 1.8–1.9) of cerium oxide at concentrations of 0, 0.005,
3 0.0505, or 0.5075 mg/L (0, 5, 50.5, or 507.5 mg/m^3) 6 hours/day, 5 days/week for 13 weeks.
4 The cerium oxide test material was 99% rare earth oxide with a maximum of 75 ppm Fe_2O_3 . Of
5 the 99% rare earth oxide, 99.95% was cerium oxide with a maximum of 25 ppm of both Pr_6O_{11}
6 and Nd_2O_3 . Praseodymium and neodymium are also rare earth metals.

7 A functional observational battery was performed on all rats, as well as activity level
8 testing, hematology, clinical biochemistry, urinalysis, ophthalmological examination, and a gross
9 pathological examination of selected tissues weighed and retained for histopathologic
10 examination. No deaths or clinical signs related to cerium oxide were noted. Food consumption
11 and body weight gain were marginally, but statistically not significantly reduced in males at
12 507.5 mg/m^3 and were considered the result of cerium oxide exposure.

13 A functional observational battery detected a statistically significant ($p < 0.05$) 17%
14 decrease in forelimb grip strength at week 13 in females exposed to 507.5 mg/m^3 . No other
15 changes were found in the functional observational battery. Motor activity, measured by
16 photocells in a figure-8 enclosure, was unaffected by cerium oxide exposure. The
17 ophthalmology examination was normal.

18 Hematological analysis revealed a statistically significant ($p < 0.05$) increase in absolute
19 neutrophil counts of 105% in 6-week males at 507.5 mg/m^3 , 130% in 6-week females at
20 50.5 mg/m^3 , 85% in 13-week males at 50.5 mg/m^3 , 75% in 13-week males at 507.5 mg/m^3 ,
21 210% in 13-week females at 5 mg/m^3 , 177% in 13-week females at 50.5 mg/m^3 , and 233% in 13-
22 week females at 507.5 mg/m^3 . Differential white blood cell counts were largely restricted to
23 altered neutrophil counts (as shown in Table 4-1), with the exception being increased absolute
24 lymphocyte and eosinophil counts in 13-week males at 50.5 mg/m^3 by 36% and 187%,
25 respectively. Differential white blood cell counts revealed changes in relative percentages of
26 neutrophils and lymphocytes. The relative percentage of neutrophils and lymphocytes were
27 significantly increased by 102 and 80% in 6-week and 13-week males, respectively, at
28 507.5 mg/m^3 . In 13-week males, there was a corresponding 19% decrease in the relative
29 percentage of lymphocytes. The 118% relative increase in neutrophils was accompanied by a
30 12% decrease in lymphocytes in 6-week females at 50.5 mg/m^3 . In 13-week females, the 5, 50.5,
31 and 507.5 mg/m^3 doses were associated with, respectively, 130, 130, and 164% relative increase
32 in the percentage of neutrophils and 12, 10, and 14% decreases in lymphocytes. Clinical
33 chemistry and urinalysis were normal.

34

Table 4-1. Hematological changes in male and female Sprague-Dawley rats following inhalation of cerium oxide aerosol 6 hours/day, 5 days/week for 13 weeks

Dose (mg/m ³)	Absolute neutrophils		Absolute lymphocytes		Absolute eosinophils		Relative neutrophils		Relative lymphocytes	
	Mean	SD	Mean	SD	Mean	SD	Mean	SD	Mean	SD
<i>Males, week 6</i>										
0	1,509	1,697	10,057	2,455	122	299	11.3	7.5	85.4	8.3
5	1,440	720	9,891	2,602	102	136	12.5	5.9	84.6	6.5
50.5	2,560	882	11,478	1,944	83	125	17.7	5.6	79	5.7
507.5	3,088 ^a	1,162	10,296	3,670	110	175	22.8 ^a	6.8	73.7 ^a	6.9
<i>Females, week 6</i>										
0	724	494	8,791	2,355	75	119	7.8	5.7	90.8	6.0
5	1,230	765	7,537	2,068	67	57	13.0	6.3	84.5	6.7
50.5	1,680 ^b	651	8,083	1,737	92	88	17 ^a	6.4	80.1 ^b	7.5
507.5	1,328	562	8,916	3,678	87	104	13.3	4.3	84.9	4.5
<i>Males, week 13</i>										
0	1,541	909	6,243	1,575	62	94	18.4	6.4	77.5	8.7
5	1,638	622	6,254	1,521	85	83	20.1	6.9	76.1	7.9
50.5	2,844 ^a	1,038	8,481 ^b	3,009	175 ^b	173	25.4	10.9	70.7	11.0
507.5	2,698 ^a	1,101	5,238	1,910	73	98	33.2 ^a	9.5	62.7 ^a	11.0
<i>Females, week 13</i>										
0	325	162	3,624	1,365	47	58	8.3	4.2	85.7	5.5
5	1,006 ^c	797	3,794	1,200	61	55	19.1 ^a	10.0	75.3 ^a	10.7
50.5	899 ^c	414	3,802	1,228	36	44	19.1 ^a	7.8	76.9 ^b	8.5
507.5	1,081 ^c	399	3,722	1,137	32	42	21.9 ^a	6.3	73.9 ^a	7.1

^aStatistically significantly different from control (Dunnett's test; $p < 0.01$).

^bStatistically significantly different from control (Dunnett's test; $p < 0.05$).

^cStatistically significantly different from control (Dunn's test; $p < 0.01$).

Source: BRL (1994).

At necropsy, there were treatment-related increases in the weight of the lungs and spleen that correlated with gross and microscopic findings. Absolute and relative lung weights were statistically significantly ($p \leq 0.001$) increased in both males and females at 50.5 and 507.5 mg/m³ (Tables 4-2 and 4-3). Lung weights, relative to brain weights, were also statistically significantly increased in male and female rats at 50.5 and 507.5 mg/m³. Relative spleen weight was statistically significantly ($p \leq 0.05$) increased in males at 507.5 mg/m³ (Table 4-4). A statistically significant increase in absolute (29%) and relative (28%) thymus weight in male mice at 50.5 mg/m³ was not considered by the study authors to be related to cerium oxide treatment. The increase in absolute and relative thymus weight was observed in only the mid-dose male rats and a dose-response relationship was not observed in male or female rats. Thus, the increase in thymus weight in male rats was not determined to be a biologically significant effect of cerium oxide exposure.

**Table 4-2. Absolute lung weight in rats exposed to cerium oxide aerosol
6 hours/day, 5 days/week for 13 weeks**

Dose (mg/m ³)	Males			Females		
	Mean weight (g)	SD	% Change	Mean weight (g)	SD	% Change
0	1.611	0.1088	–	1.145	0.0871	–
5	1.760	0.1398	9	1.311	0.1490	14
50.5	2.574 ^a	0.3499	60	1.651 ^a	0.2900	44
507.5	4.662 ^a	0.5161	189	3.173 ^a	0.3235	177

^aStatistically significantly different from control (Dunn’s test; $p < 0.01$).
Source: BRL (1994).

**Table 4-3. Relative lung weight in rats exposed to cerium oxide aerosol
6 hours/day, 5 days/week for 13 weeks**

Dose (mg/m ³)	Males			Females		
	Mean weight (g%)	SD	% Change	Mean weight (g%)	SD	% Change
0	0.334	0.0256	–	0.477	0.0443	–
5	0.371	0.0270	11	0.544	0.0557	14
50.5	0.528 ^a	0.0775	58	0.697 ^a	0.1124	46
507.5	1.024 ^a	0.1621	207	1.358 ^a	0.1432	185

^aStatistically significantly different from control (Dunn’s test; $p < 0.01$).
Source: BRL (1994).

**Table 4-4. Relative spleen weight in rats exposed to cerium oxide aerosol
6 hours/day, 5 days/week for 13 weeks**

Dose (mg/m ³)	Males			Females		
	Mean weight (g%)	SD	% Change	Mean weight (g%)	SD	% Change
0	0.162	0.0190	-	0.216	0.0281	-
5	0.169	0.0286	4	0.242	0.0259	12
50.5	0.178	0.0266	10	0.226	0.0445	5
507.5	0.188 ^a	0.0298	16	0.222	0.0252	3

^aStatistically significantly different from control (Dunnett’s test; $p < 0.05$).
Source: BRL (1994).

Gross examination found discoloration or pale areas, pale foci, and uncollapsed parenchyma in the lungs of male and female rats (Table 4-5). Pale areas and discoloration in the lung were evident at 50.5 and 507.5 mg/m³ in male and female rats, respectively, with uncollapsed parenchyma evident at 50.5 and 507.5 mg/m³ in male rats and 507.5 mg/m³ in female rats. Pale foci in the lungs were only seen in female rats exposed to 5 mg/m³. The incidence of enlargement or pale discoloration of the mandibular, bronchial, mediastinal, and pancreatic lymph nodes is shown in Table 4-6. Enlargement or pale discoloration of both lymph nodes that drain the lungs (the bronchial and mediastinal lymph nodes) was evident in both

1 males and females at ≥ 5 mg/m³ cerium oxide. The mandibular and pancreatic lymph nodes did
 2 not display a dose-response trend of enlargement or discoloration. The study authors (BRL,
 3 1994) judged the mandibular lymph node enlargement, which was observed in control and
 4 cerium-exposed rats, not to be an effect of the cerium oxide exposure, while the pancreatic,
 5 bronchial, and mediastinal lymph node enlargement and/or discoloration were considered to be
 6 related to the cerium oxide treatment.

7
 8 **Table 4-5. Results of gross pathological examination of lungs of rats**
 9 **exposed to cerium oxide 6 hours/day, 5 days/week for 13 weeks**

Lung	Dose (mg/m ³)							
	Male				Female			
	0	5	50.5	507.5	0	5	50.5	507.5
Pale foci	0/15	0/15	0/15	0/15	0/15	4/15	0/15	0/15
Pale areas or discoloration	0/15	0/15	15/15	15/15	0/15	0/15	15/15	15/15
Uncollapsed parenchyma	0/15	0/15	2/15	15/15	0/15	0/15	0/15	15/15

10 Source: BRL (1994).

11
 12
 13 **Table 4-6. Results of gross pathological examination of bronchial,**
 14 **mediastinal, and pancreatic lymph nodes of rats exposed to cerium oxide**
 15 **6 hours/day, 5 days/week for 13 weeks**

Lymph nodes	Dose (mg/m ³)							
	Male				Female			
	0	5	50.5	507.5	0	5	50.5	507.5
Mandibular								
enlargement	4/15	2/15	5/15	4/15	2/15	4/15	3/15	1/15
discoloration	–	–	–	–	–	–	–	–
Bronchial								
enlargement	0/15	4/15	15/15	15/15	0/15	1/15	14/15	15/15
discoloration	0/15	13/15	15/15	15/15	0/15	15/15	15/15	15/15
Mediastinal								
enlargement	0/15	2/15	9/15	9/15	0/15	1/15	8/15	8/15
discoloration	0/15	2/15	9/15	10/15	1/15	10/15	9/15	10/15
Pancreatic								
enlargement	1/15	0/15	0/15	0/15	2/15	0/15	1/15	0/15
discoloration	–	–	–	–	0/15	0/15	0/15	1/15

16 Source: BRL (1994).

17
 18 Histologic examination revealed dose-related alveolar epithelial and lymphoid
 19 hyperplasia and pigment accumulation in the lungs, lymph nodes, and larynx of male and female
 20 rats at ≥ 5 mg/m³. The incidence data are presented in Table 4-7. The pigment, which was also
 21 found in other parts of the respiratory tract, including the nasal cavities and the trachea, and the
 22 liver and spleen, was considered by the study authors to be the test compound or a product
 23 thereof. The lymphoid hyperplasia in the lymph nodes following cerium oxide exposure was
 24 characterized by the study pathologist as an increase in the number of lymphocytes, with lymph

1 node paracortices and cortices expansion. The authors reported that the severity of the
2 hyperplasia in a given tissue, lung or lymph node, was correlated with the amount of pigment
3 accumulated in the tissue but did not present any supporting data. BRL (1994) considered these
4 findings to be consistent with antigenic stimulation by cerium oxide; however, they did not
5 discuss the possibility of non-antigenic stimulation. The metaplasia evident in the larynx was
6 interpreted by the study pathologist as adaptive and reversible. Lesions were not observed in the
7 testes or ovaries of the high-dose group.

8 The NOAEL and LOAEL values for the toxicological effects observed are included in
9 Table 4-8. This study identified a LOAEL of 5 mg/m³ in rats, based on the increased incidence
10 of lymphoid hyperplasia in the bronchial lymph nodes of male and female rats. A NOAEL was
11 not identified.

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Table 4-7. Incidences of histopathologic effects in rats exposed to cerium oxide aerosol 6 hours/day, 5 days/week for 13 weeks

Exposure (mg/m ³)	Control	5	50.5	507.5
<i>Males</i>				
Larynx				
metaplasia	0/15	3/15	9/15 ^a	13/15 ^a
pigment accumulation	0/15	6/15 ^a	9/15 ^a	12/15 ^a
Lung				
lymphoid hyperplasia	0/15	0/15	0/15	12/15 ^a
alveolar epithelial hyperplasia	0/15	1/15	11/15 ^a	14/15 ^a
pigment accumulation	0/15	15/15 ^a	15/15 ^a	15/15 ^a
Bronchial lymph node				
lymphoid hyperplasia	0/15	11/13 ^a	15/15 ^a	15/15 ^a
pigment accumulation	0/15	13/13 ^a	15/15 ^a	15/15 ^a
Mediastinal lymph node				
lymphoid hyperplasia	0/0	2/2	9/10	9/9
pigment accumulation	0/0	2/2	8/10	9/9
Mandibular lymph node				
lymphoid hyperplasia	0/15	0/3	0/5	2/15
pigment accumulation	0/15	0/3	0/5	6/15 ^a
Pancreatic lymph node				
lymphoid hyperplasia	–	–	–	–
pigment accumulation	–	–	–	–
Spleen				
pigment accumulation	0/15	0/15	0/15	6/15 ^a
<i>Females</i>				
Larynx				
metaplasia	0/15	3/15	6/15 ^a	9/15 ^a
pigment accumulation	0/15	0/15	7/15 ^a	9/15 ^a
Lung				
lymphoid hyperplasia	0/15	0/15	1/15	7/15 ^a
alveolar epithelial hyperplasia	0/15	0/15	5/15 ^a	15/15 ^a
pigment accumulation	0/15	15/15 ^a	15/15 ^a	15/15 ^a
Bronchial lymph node				
lymphoid hyperplasia	0/15	13/15 ^a	15/15 ^a	15/15 ^a
pigment accumulation	0/15	14/15 ^a	15/15 ^a	15/15 ^a
Mediastinal lymph node				
lymphoid hyperplasia	0/1	10/10	9/9	9/10
pigment accumulation	0/1	10/10	9/9	9/10
Mandibular lymph node				
lymphoid hyperplasia	0/15	0/5	0/3	0/15
pigment accumulation	0/15	0/5	0/3	6/15 ^a
Pancreatic lymph node				
lymphoid hyperplasia	0/2	0/0	1/1	0/1
pigment accumulation	0/2	0/0	1/1	1/1
Spleen				
pigment accumulation	0/15	0/0	0/0	3/15

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^aSignificantly different from vehicle control group ($p < 0.05$ by Fisher's exact test)
Source: BRL, 1994.

Table 4-8. NOAEL and LOAEL values for toxicological effects

1

observed in BRL, 1994.

Toxicological effect	Sex	NOAEL (mg/m ³)	LOAEL (mg/m ³)
<i>Organ weight changes</i>			
Absolute lung weight	M, F	5	50.5
Relative lung weight	M, F	5	50.5
Absolute spleen weight	M, F	507.5	–
Relative spleen weight	M	50.5	507.5
	F	507.5	–
<i>Gross pathological lesions in lung</i>			
Pale areas	M, F	5	50.5
Discoloration	M, F	50.5	507.5
Uncollapsed parenchyma	M	5	50.5
	F	50.5	507.5
<i>Gross pathological lesions in the lymph nodes</i>			
Mandibular enlargement	M, F	–	5
Bronchial enlargement	M, F	–	5
Bronchial discoloration	M, F	–	5
Mediastinal enlargement	M, F	–	5
Mediastinal discoloration	M, F	–	5
<i>Histopathologic lesions</i>			
Metaplasia, larynx	M, F	–	5
Pigment accumulation, larynx	M	–	5
	F	5	50.5
Lymphoid hyperplasia, lung	M	50.5	507.5
	F	5	50.5
Alveolar epithelial hyperplasia, lung	M, F	5	50.5
Pigment accumulation, lung	M, F	–	5
Lymphoid hyperplasia, bronchial lymph node	M, F	–	5
Pigment accumulation, bronchial lymph node	M, F	–	5
Pigment accumulation, spleen	M	50.5	507.5
Pigment accumulation, mandibular lymph node	M	50.5	507.5
Lymphoid hyperplasia, mediastinal lymph node	M, F	–	5
Pigment accumulation, mediastinal lymph node	M, F	–	5

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Intratracheal instillation of 50 mg of a dust suspension of cerium oxide produced mild changes in the lung and spleen of white rats examined 8 months later (Mogilevskaya and Raikhlin, 1967). There was a moderate proliferative response in the lungs and bronchi, with occasional increased lymphocyte numbers and very slight development of connective tissue and collagen fibers, but diffuse or nodular fibrotic changes were lacking. In the spleen, macrophages and large multinucleated cells accumulated. No histologic changes were found in any of the other major organs.

Lundgren et al. (1996) conducted an investigation into the pulmonary carcinogenicity of beta-particle radiation from inhaled ¹⁴⁴CeO₂ in F344/N rats, with stable cerium oxide serving as the control group (n = 1,049). The radiation doses ranged from 3.6 to 37 Gy. The control rats

1 received stable cerium oxide in a single inhalation dose of comparable mass concentration to the
2 dose groups, although the concentration was not specified by the study authors. The control rats
3 were held for life-span observation and were evaluated histologically. The histologic evaluation
4 showed nonneoplastic lesions, such as inflammation (5.1%), fibrosis (5.6%), alveolar-epithelial
5 hyperplasia (4.5%), and alveolar macrophage hyperplasia (7.1%), in the lungs of control rats.
6 Seven primary lung neoplasms were also apparent in the control rats and included four alveolar
7 or papillary adenomas; one alveolar, papillary, or tubular adenocarcinoma; one squamous cell
8 carcinoma; and one fibro- or osteosarcoma. Control rats, unexposed to cerium oxide, in a
9 separate study of inhaled ¹⁴⁴CeO₂ in F344/N rats demonstrated an incidence of lung tumors of
10 6/110 (Lundgren et al., 1992). The observed tumors included two papillary adenomas, two
11 papillary adenocarcinomas, one adenosquamous carcinoma, and one mesothelioma.

12

13 **4.3. REPRODUCTIVE/DEVELOPMENTAL STUDIES—ORAL AND INHALATION**

14 No studies were located regarding reproductive/developmental effects of cerium in
15 humans by any route of exposure. The only relevant information from a study in animals is the
16 administration of 0, 200, or 800 mg/kg-day cerium to male mice (five to eight per group) in the
17 diet for 30 or 45 days. This exposure increased the rate of unspecified sperm abnormalities but
18 did not statistically significantly affect testes weight or alter serum levels of testosterone (Yu et
19 al., 2001, English translation of abstract). The increased rate of the apparent sperm
20 abnormalities was dose and time dependent, with a 17 and 22% increase in abnormalities 30
21 days postexposure for 200 and 800 mg/kg-day, respectively, and a 28 and 32% increase in
22 abnormalities 45 days postexposure at the same dose levels, respectively.

23

24 **4.4. OTHER DURATION- OR ENDPOINT-SPECIFIC STUDIES**

25 As indicated in Section 3.1.1, cerium compounds are poorly absorbed from the GI tract of
26 animals; thus, parenteral administration of cerium compounds has been the preferred route of
27 exposure to study the systemic effects of these compounds.

28

29 **4.4.1. Acute Toxicity Studies (Oral and Inhalation)**

30 Single dose and/or acute oral data are limited but include a report of splenic lesions,
31 including hypertrophy, reticuloendothelial hyperplasia, and hyperactive lymphoid follicles, and
32 GI irritation, characterized by gastritis and enteritis with focal hemorrhage and necrosis of the
33 mucosa in the stomach and duodenum, in mice given single gavage doses of 1,000 or
34 1,163 mg/kg (1:3 cerium chloride-sodium citrate complex) (Stineman et al., 1978). Open field
35 behavior was not affected and the 7-day LD₅₀ was 1,291 mg/kg cerium (95% CI: 1,198–1,440).
36 Bruce et al. (1963) identified an oral LD₅₀ of 4,200 mg/kg for cerium nitrate in female Sprague-
37 Dawley rats (n=30).

1 There is a report of piloerection, hunched posture, unsteady gait, and dark coloring of
2 eyes in male and female rats, along with lethargy, abnormal respiration, and prostration in
3 females, given single gavage doses of 5,000 mg/kg cerium as cerium sulfide (Rhodia Inc., 1998).
4 Ji and Cui (1988) list an oral LD₅₀ in female mice of 1,178 mg/kg (95% CI: 1,043–1,331) for
5 cerium nitrate (Ce[NO₃]₃) and 622 mg/kg (95% CI: 550–702) for cerium oxides.

6 Data on the toxicity of single exposures to airborne cerium in animals are limited. A
7 review by the HEI (2001) reported that the LC₅₀ for cerium oxide in rats was >50 mg/m³ in a
8 study by Rhone-Poulenc (1983), but the study was unavailable for examination and no further
9 details were reported.

11 **4.4.2. Acute Studies (Injection)**

12 **4.4.2.1. Neurobehavioral and Neurodevelopmental Effects**

13 D'Agostino et al. (1982, 1978a, b) studied the neurodevelopmental effects of cerium
14 following subcutaneous injection in mice. The three separate publications present the same
15 experimental design. Pregnant mice were administered a single dose of 80 mg/kg cerium
16 (sodium/citrate complex) or citrate (control) on gestation day (GD) 7 or 12 or postnatal day
17 (PND) 2. In order to differentiate the gestational effects from lactational effects or changes in
18 maternal behavior, a full cross-foster study design was employed. A cross-fostering design
19 distributed the offspring (3 males and 3 females per dam) of mothers receiving cerium or citrate
20 during gestation to lactating dams exposed to either cerium or citrate. Body weight and gross
21 activity of the neonates were assessed on PNDs 7 or 12 (D'Agostino et al., 1978a, b) or PNDs 8
22 or 13 (D'Agostino et al., 1982), whereas open-field behavior, accelerating rotarod performance,
23 and passive avoidance learning were assessed on PNDs 60–65. Maternal offspring retrieval
24 latency was measured on PND 3. The growth and behavioral data were analyzed via a
25 multivariate analysis.

26 Pups exposed to cerium in utero on GDs 7 and 12 demonstrated a statistically significant
27 decrease in body weight on PNDs 7 and 12 (D'Agostino et al., 1978a, b), whereas the body
28 weight of offspring of cerium-exposed dams on GD 7 did not differ significantly from that of
29 control offspring (D'Agostino et al., 1982). Offspring of dams exposed to cerium on GD 12
30 showed significantly decreased body weights on PNDs 8 and 13 (D'Agostino et al., 1982). Pups
31 exposed to cerium in utero were retrieved by dams and replaced in the nest significantly faster
32 than controls. When the dams were injected on PND 2, neonatal weights on PNDs 7 and 12 and
33 8 and 13 were statistically significantly reduced (D'Agostino et al., 1982, 1978a, b). The study
34 authors suggested that these effects may be due to altered maternal behavior (e.g., ineffective
35 suckling or lack of grooming) rather than cerium being transmitted to offspring in the milk. In
36 addition, offspring of rats exposed to cerium on GD 7 showed a higher frequency of rearings
37 than controls when evaluated on PNDs 60–70 (D'Agostino et al., 1982, 1978a, b). Rats exposed

1 to cerium on GD 7 demonstrated a decrease in activity, measured by the circular runway, on
2 PND 12 (D'Agostino et al., 1978a, b). Other behavioral differences were not reported. No
3 possible mechanism of action was discussed to explain the neurodevelopmental effects of cerium
4 administration. The lack of information on maternal health status following administration of
5 cerium limits the usefulness of these studies.

7 **4.4.2.2. Neurological Effects**

8 Morganti et al. (1978) examined the open-field and exploratory behavior of 6- to 8-week-
9 old male Swiss ICR mice following subcutaneous administration of cerium. Three days after
10 receiving a dose of 20 mg/kg cerium (from a 1:3 cerium chloride-cerium citrate complex),
11 10 mice were observed in the open-field and exploratory apparatuses and then sacrificed. This
12 procedure, including injection, was repeated every 3 days until the last group of mice had
13 received a total of 200 mg/kg cerium. Control mice were injected with sodium citrate. The
14 results showed that cerium exposure statistically significantly ($p \leq 0.05$) depressed ambulations,
15 marginally depressed explorations, but did not affect rearings. Behavioral measures were not
16 correlated with cerium levels in tissues. According to Morganti et al. (1978), the lack of
17 correlation between the behavioral measures and the cerium levels in tissue indicated that the
18 effect of cerium is not a one-step process depending directly on the level of cerium in a single
19 tissue but instead involves a biological chain of events.

20 In a different study by the same group of investigators (Stineman et al., 1978) in which
21 male Swiss mice received single, subcutaneous injections of cerium (136 or 173 mg/kg), there
22 was an inverse relationship between open field behavior (ambulations and rearings) and levels of
23 cerium in blood, brain, lung, stomach, intestine, and kidney. The levels of cerium in the brain
24 had the strongest association with decreased open field behavior, followed by the lung.
25 However, splenic levels of cerium were positively correlated with behavior, as mice with higher
26 levels of cerium in the spleen exhibited less altered behavior. Stineman et al. (1978) suggested
27 that the spleen may protect against the cerium-induced effects by sequestering cerium from the
28 circulation after the removal of damaged erythrocytes and leukocytes.

29 In a subsequent study (Morganti et al., 1980), 6- to 8-week-old adult male Swiss ICR
30 mice administered single, subcutaneous injections of 136 or 173 mg/kg cerium showed
31 depression of general activity, as measured in an activity wheel study and a passive avoidance
32 study. However, the subcutaneous injection of cerium did not significantly affect two-way
33 active avoidance learning or social behavior, although gross activity was depressed in the social
34 behavior study (Morganti et al., 1980). The study authors' interpretation of the study data was
35 that cerium exposure did not affect simple and complex learning, as measured in the above tests.

37 **4.4.2.3. Hepatic Effects**

1 Numerous studies have examined the effects of cerium on liver parameters following
2 parenteral dosing. The i.v administration of a dose of 3 mg/kg cerium (specific form not
3 specified) to rats resulted in increased serum levels of ornithine-carbonyl transferase (OCT),
4 which reached a peak 1–2 days after dosing (Magnusson, 1962). Blood glucose levels were
5 reduced markedly in females 2–3 days after dosing but recovered by day 4. No macroscopic
6 changes were seen in the liver of males, but fatty liver and fatty degeneration were evident in
7 females 1 day after treatment. Males showed hydropic changes and isolated necrotic cells 1–
8 4 days after dosing. In females, changes to the ultrastructure of the liver cells were seen 12
9 hours after administration of cerium and appeared to reach a maximum 2–3 days later, with only
10 slight ultrastructural changes observed in males.

11 Several additional reports have confirmed and expanded on the Magnusson (1962)
12 findings. Marciniak and Baltrukiewicz (1981, 1977) showed that the increase in serum OCT
13 activity after cerium injection was linear in the range 1.5–4.5 mg/kg cerium. Lombardi and
14 Recknagel (1962) observed that i.v. administration of 5 mg/kg cerium (form not specified) to
15 female Sprague-Dawley rats produced a fourfold increase in liver triglyceride levels 24 hours
16 after treatment. Salas et al. (1976) showed that a single i.v. dose of 10 mg/kg cerium chloride
17 administered to female Sprague-Dawley rats decreased the total hepatic adenosine triphosphate
18 level by 12 hours postexposure and depleted liver glycogen levels and increased liver
19 triglyceride levels within 48 hours. Ultrastructurally, the rough endoplasmic reticulum was the
20 main target of cerium toxicity, with marked dilation and degranulation, as well as the appearance
21 of free ribosomes in the cytoplasm, 24 hours postexposure. The hepatic changes returned to
22 normal between the 5th and 8th day postexposure.

23 Arvela and Karki (1971) observed fatty degeneration on the first day following i.v.
24 administration of 2 mg/kg cerium chloride in Sprague-Dawley rats. On the third day after i.v.
25 administration, there was a 53% reduction in the level of CYP450 in the liver of male Sprague-
26 Dawley rats. Arvela et al. (1991) showed that C57BL/6N male mice were more resistant to the
27 hepatotoxic effects, including necrosis, cell membrane disintegration, and microsteatosis, of
28 cerium than were DBA/2 mice. However, the concentration of cerium in the livers of C57BL/6N
29 mice was about 50% higher at 72 hours than in the livers of DBA/2 mice. The authors also
30 showed that in C57BL/6N mice the slight to moderate liver injury 72 hours post administration
31 was associated with an increase in coumarin 7-hydroxylase (COH) activity; however, at doses
32 that caused severe damage in DBA/2 mice, COH activity 72 hours post administration was
33 drastically reduced. It was also reported that the total amount of CYP450 in the liver was
34 significantly increased in both strains after a dose of 1 mg/kg cerium, but higher doses tended to
35 decrease CYP450 content, producing a biphasic relationship. A follow-up study found that
36 cerium increased the amount of hepatic CYP2A5 mRNA only in DBA/2 mice (Salonpää et al.,
37 1992), which have been shown to be more sensitive to cerium exposure than the C57BL/6N

1 strain. Since the CYP2A5 gene encodes P450 isozymes catalyzing COH activity in mice, the
2 study authors suggested that some association exists between the development of liver damage
3 and COH induction.

4 Strubelt et al. (1980) reported dose-dependent increases in AST (reported using the older
5 name of glutamic-oxaloacetic transaminase) and ALT (reported using the older name of
6 glutamic-pyruvic transaminase) and sorbitol dehydrogenase (SDH) in male Wistar rats after i.v.
7 administration of 3, 5, 7, and 10 mg/kg cerium nitrate.

8 Sex differences in sensitivity were also reported by Wiener-Schmuck et al. (1990).
9 Intravenous injection of 1.3 mg/kg of cerium chloride to rats caused lipid deposition, damage to
10 mitochondria, and invaginations of the nuclear membrane in hepatocytes 48 hours after dosing.
11 These changes, which were accompanied by increased activities of serum transaminases, were
12 reversible and occurred only in females. However, when isolated hepatocytes of female rats
13 were incubated in medium with cerium chloride for 20 hours, there was no sign of cell damage.
14 The lack of toxicity of cerium to isolated hepatocytes in vitro suggests that hepatocellular lesions
15 and related changes observed following injection of cerium result from an indirect cause.
16 Wiener-Schmuck et al. (1990) suggested that deposition of injected rare earths in Kupffer cells
17 leads to blockage of the reticuloendothelial system in the liver, inducing damage to
18 macrophages, which release mediators that in turn damage the hepatocytes.

19 A more recent paper showed that administration of the metal chelator ethylene glycol
20 bis(2-aminoethylether)tetraacetic acid (EGTA) to mice after dosing with cerous sulfate for 7
21 days decreased the severity of the histologic effects of cerium on the liver (Shrivastava and
22 Mathur, 2004). The histologic effects that resulted from this 7-day subcutaneous exposure of 0.5
23 mL cerous sulfate included lymphocytic infiltration, hepatocyte hypertrophy, cytoplasmic
24 vacuolation, and stumpy Kupffer cells. In addition to the histologic effects, the activity of ALT
25 and AST were statistically significantly increased, with maximum increases of 350 and 98%,
26 respectively, and hepatic acid, alkaline phosphatases, and succinic dehydrogenase were
27 statistically significantly decreased, 42, 69, and 46%, respectively. The decrease in severity of
28 the histologic effects would indicate that the presence of circulating cerium is necessary for
29 hepatotoxicity but does not provide information on a possible mechanism of hepatotoxicity.

30 In summary, administration of cerium to rodents caused lipid deposition, mitochondrial
31 damage, and invaginations of the nuclear membrane in hepatocytes, as well as adverse
32 morphological effects in the liver, characterized by fatty liver, fatty degeneration, and necrosis.
33 In addition, histologic effects included lymphocytic infiltration, hepatocyte hypertrophy,
34 cytoplasmic vacuolation, and stumpy Kupffer cells. Arvela and Karki (1971) suggested that
35 liver toxicity may result indirectly from induction of CYP2A5 and COH.

36 37 **4.4.2.4. Cardiovascular Effects**

1 Information regarding effects of cerium on the human heart derives mainly from a series
2 of studies (see Section 4.1.1) in which naturally high levels of cerium in the soil in certain
3 geographical regions appear to be correlated with higher levels of cerium in serum and cardiac
4 tissue of individuals with endomyocardial fibrosis (Eapen, 1998; Kutty et al., 1996; Valiathan et
5 al., 1989).

6 The i.v. administration of a single dose of 1.3 mg/kg cerium as cerium chloride to female
7 Sprague-Dawley rats resulted in a statistically significant twofold increase in protein synthesis
8 ($p \leq 0.001$) and transcription ($p \leq 0.01$) in cardiac muscle relative to controls 24 hours after
9 injection (Kumar et al., 1995). This was consistent with findings by the same group of
10 investigators who reported that incubation of cardiac fibroblasts in vitro with 100 nM cerium
11 increased RNA synthesis approximately 64%, but the rate of DNA synthesis was unaffected
12 (Shivakumar et al., 1992). However, it should be noted that higher concentrations of cerium in
13 the medium were inhibitory. This was taken as evidence suggesting that cerium at very low
14 levels may act at the level of transcription to stimulate collagen and non-collagen protein
15 synthesis. This, in turn, may contribute to the accumulation of collagen in endocardial fibrosis.
16 In a follow-up study, Kumar and Shivakumar (1998) reported that a single i.v. dose of 1.3 mg/kg
17 cerium increased lipid peroxidation by 30% in cardiac tissue of Sprague-Dawley rats and
18 increased proliferation of cardiac fibroblasts by 23%. Treatment with cerium also statistically
19 significantly decreased collagen degradation by 7% and increased the rate of deposition of newly
20 synthesized collagen by 27% in cardiac tissue 48 hours after cerium administration (Kumar and
21 Shivakumar, 1998).

22 In summary, the limited data from oral exposure experiments in rats and rabbits and i.v.
23 administration studies in rats suggest that cerium increases collagen accumulation in cardiac
24 muscle by increasing synthesis and decreasing degradation by unknown mechanisms.

26 **4.4.2.5. Hematological Effects**

27 The i.v. administration of a single dose of 10 mg/kg cerium as the chloride, citrate
28 complex, or ethylenediamine tetraacetic acid (EDTA) complex to anesthetized dogs every
29 10 minutes for 10 total doses, significantly increased prothrombin levels and coagulation time
30 within minutes of the injection (Graca et al., 1964). The magnitude of the increased prothrombin
31 levels and coagulation time was greatest for cerium chloride, followed by the citrate complex,
32 and finally the EDTA complex. No other hematological endpoint was significantly affected by
33 cerium exposure. Talbot et al. (1965) implanted a pellet of cerium metal under the skin of
34 C57BL mice and collected blood samples from five male and five female mice every 6 months
35 for hematological determinations. They stated that there were no significant differences between
36 coagulation times of cerium-implanted mice and controls; however, a table shows that after
37 6 months, coagulation time in implanted male mice was approximately twice that in controls. In

1 this study, the only statistically significant difference ($p < 0.05$) between cerium implanted and
2 nonimplanted mice was a decrease in total leukocyte counts in males and females at 6, 12, and
3 18 months relative to controls. However, analysis of differential counts showed no significant
4 differences between treated and control mice. In another study, Shrivastava and Mathur (2004)
5 injected subcutaneous doses of cerous sulfate ($\text{Ce}_2[\text{SO}_4]_3$) daily for 7 days into male mice and
6 reported 18 and 37% decreases in hemoglobin and red blood cell counts, respectively, and 93
7 and 39% increases in sedimentation rate and hematocrit, respectively, all of which appeared to
8 reach a maximum on day 14 (7 days after the last injection) and appeared to return to control
9 levels approximately 60 days posttreatment.

10 The information available from these few studies is insufficient to determine a possible
11 mechanism of action by which cerium might be causing hematological alterations. Alterations in
12 prothrombin and coagulation times could be secondary to liver dysfunction, but more
13 information is necessary to confirm this hypothesis.

14 15 **4.4.2.6. Renal Effects**

16 The i.v. administration of a single dose of 2 mg/kg of cerium chloride to adult male
17 DBA/2 and C57BL/6 mice resulted in a more than fourfold increase in COH activity in the
18 kidneys of DBA/2 mice, but less than a twofold increase in C57BL/6 mice (Salonpää et al.,
19 1992). The effect was maximized in DBA/2 mice 4 hours after dosing, and the enzyme activity
20 returned to predosing levels 6 hours after treatment. Cerium also increased CYP2A5 mRNA in
21 the kidneys of DBA/2 mice sevenfold 6 hours after dosing and sixfold 1 day after the injection,
22 but no such increase occurred in C57BL/6 mice.

23 Injection of subcutaneous doses of 1 mM cerous sulfate to male mice for 7 days produced
24 statistically significant ($p < 0.05$) decreases up to 62 and 42% in the activities of renal acid and
25 alkaline phosphatases, respectively, and up to 31% in succinic dehydrogenase activity
26 (Shrivastava and Mathur, 2004). Cerium also induced histologic damage to the kidneys,
27 consisting of hypertrophy in the epithelial cells, deformed Bowman's capsules, exfoliated nuclei
28 in tubular lumen, and leukocyte infiltration. Maximum injury was observed on day 21 (14 days
29 after the last injection). As with the findings regarding liver toxicity, mice administered EGTA
30 after cerium exhibited less severe necrotic effects in the kidney than mice not receiving EGTA.
31 The mechanism of kidney toxicity of cerium is unknown.

32 In summary, cerium exposure initiated an increase in COH activity and CYP2A5
33 expression and decreased acid and alkaline phosphatases and succinic dehydrogenase activities,
34 as well as kidney epithelial hypertrophy, exfoliated nuclei in the tubular lumen, and leukocyte
35 infiltration.

36 37 **4.4.3. Genotoxicity**

1 No information was located regarding genotoxic effects of cerium or cerium compounds
2 in humans and only three studies were identified with pertinent information in mammals and in
3 lower organisms (Sharma and Taluker, 1987; Shimizu et al., 1985; Nishioka, 1975). The
4 available information is insufficient to ascertain the genotoxicity of cerium.

5 Shimizu et al. (1985) examined the potential mutagenicity of cerium oxide in five strains
6 of *Salmonella typhimurium* (TA98, TA100, TA1535, TA1537, and TA1538) and in *Escherichia*
7 *coli* WP2uvrA. Tests were conducted with and without metabolic activation (S9 fraction from
8 male Sprague-Dawley rats). Eight different concentrations were tested, ranging from 1 to
9 5,000 µg/plate. No increase in the number of revertant colonies per plate was observed at any
10 dose level; in some cases the highest concentration produced growth inhibition. In a short
11 communication, Nishioka (1975) reported that cerium chloride did not induce DNA damage in
12 two strains of *Bacillus subtilis* using the rec-assay. Cerium nitrate was reported to induce
13 chromosomal breaks and reduce the mitotic index in rat bone marrow in vivo (Sharma and
14 Talukder, 1987).

16 **4.5. MECHANISTIC DATA AND OTHER STUDIES IN SUPPORT OF THE MODE OF** 17 **ACTION**

18 **4.5.1. In Vitro Studies**

19 The cytotoxicity of soluble cerium chloride and insoluble cerium oxide was assayed in
20 Sprague-Dawley rat pulmonary alveolar macrophages (PAMs) and compared to the cytotoxic
21 and fibrogenic cadmium chloride and oxide (Palmer et al., 1987). Cell viability, effect on
22 lysosomal enzyme release, and cell morphology were investigated. Cerium chloride was
23 cytotoxic to rat PAMs with an LC₅₀ (concentration inducing 50% cell death) of 29 µM, while
24 cerium oxide was less toxic, with an LC₅₀ of approximately 4,700 µM; the LC₅₀ values in this
25 assay for cadmium chloride and oxide were 28 and 15 µM, respectively. Cerium chloride and
26 oxide did not affect lysosomal enzyme release, although the sensitivity of this assay was
27 questioned by the study authors. Cerium oxide induced an increase in cells with a featureless
28 surface and a decrease in cells consistent with the control population. Cells typified by blebs on
29 the cell surface and/or surface structure were absent. The induction of cells with a featureless
30 surface was considered by the study authors to be minimal. Cerium chloride was not evaluated
31 for effect on cell morphology due to an experimental error during cell culture preparation.
32 Cerium chloride was more cytotoxic than cerium oxide and of similar cytotoxicity to cadmium
33 chloride, which had an LC₅₀ of 28 µM.

34 Shivakumar and Nair (1991) conducted an in vitro study to examine the effect of cerium
35 on protein synthesis in cultured rat heart cells and human lung fibroblasts exposed to normal and
36 reduced levels of magnesium in the growth medium. Cerium exposure resulted in the inhibition
37 of protein synthesis in rat heart cells and human lung fibroblasts, evident by the decreased

1 amount of [³H]-tyrosine incorporated into heart cell and fibroblast proteins, 73 and 76%,
2 respectively; the cell cultures grown on low-magnesium medium displayed a more pronounced
3 decrease in protein synthesis, with [³H]-tyrosine incorporation decreased 51 and 40% in heart
4 cell and fibroblast proteins, respectively. However, the mechanism of protein synthesis
5 inhibition is unknown.

6 Another in vitro study was conducted to investigate the toxicity of lanthanides, which
7 includes cerium, towards cultured rat alveolar macrophages (Lizon and Fritsch, 1999). Rat
8 alveolar macrophages were acquired by pulmonary lavage of 2-month-old male Sprague-Dawley
9 rats and were cultured for 1 day on culture medium and then for 3 days on medium containing
10 soluble cerium concentrations of 1×10^{-4} and 5×10^{-6} M. The fraction of apoptotic cells
11 increased with concentration and as a function of time, with observations at 24, 48, and 78 hours.
12 As cerium concentration increased from 10^{-6} to 5×10^{-5} M, the fraction of normal cells
13 decreased and the fraction of post-apoptotic and unstained cells increased. The LC₅₀ for cerium
14 was approximately 5×10^{-5} M. Cerium exposure to cultured rat alveolar macrophages resulted
15 in significant alveolar macrophage death.

16 Studies with cardiac fibroblasts from neonatal rats in vitro showed that the stimulation of
17 fibroblast proliferation was accompanied by an increase in the generation of free radicals. Preeta
18 and Nair (1999) isolated cells from the hearts of 3–4-day-old neonatal Wistar rats. Fibroblasts
19 were selected from the cultured isolated heart cells via selective adhesion and reincubated in
20 fresh medium. To assess fibroblast growth, the selected fibroblasts were exposed to cerium
21 concentrations of 0.1, 0.5, 1, 10, and 100 μ M and were harvested 96 hours postexposure.
22 Growth dynamics were evaluated at 24, 48, and 96 hours postexposure. Immunohistochemical
23 labeling for proliferating cell nuclear antigen (PCNA) was also conducted to determine if an
24 increase in cell number was due to cell proliferation. Intracellular free-radical generation was
25 determined by spectrophotometric assay with reduced nitroblue tetrazolium, with the cardiac
26 fibroblasts exposed to the same cerium concentrations as in the growth assessment. In addition,
27 the role of free radicals in cell proliferation was investigated using cardiac fibroblasts exposed to
28 0.5 μ M cerium, SOD (100 U/mL), and catalase (120 U/mL) with cell counts measured after
29 96 hours.

30 The results of this investigation showed that cardiac fibroblast proliferation followed a
31 concentration-dependent response to cerium exposure, with increased proliferation at low
32 concentrations and decreased proliferation at high concentrations (Preeta and Nair, 1999).
33 Increased proliferation was evident from 0.1 to 1 μ M, with a statistically significant ($p < 0.01$)
34 peak at 0.5 μ M, while decreased proliferation was evident from 10–100 μ M. A statistically
35 significant increase in the proliferation of PCNA immunoreactive cells was evident at low
36 cerium concentrations (0.1–1.0 μ M), while a decrease was evident at higher concentrations. The
37 addition of SOD inhibited the increased PCNA expression. The reduction of nitroblue

1 tetrazolium to formazan, which peaked at 0.5 μM , showed the increase in free radicals from the
2 fibroblasts exposed to cerium. An increase in free radical production resulted from the fibroblast
3 exposure to cerium at 0.5 μM , and the cerium-induced stimulatory response was inhibited by the
4 addition of SOD. This in vitro study demonstrates that low concentrations of cerium stimulate
5 cardiac fibroblast proliferation in association with an increase in intracellular generation of free
6 radicals.

7 A comparative study of cardiac and pulmonary fibroblasts in vitro by Nair et al. (2003)
8 was conducted to investigate the mitogenic effect of cerium. Fibroblasts were isolated from
9 heart and lung tissue of 3- to 4-day-old neonatal Wistar rats through selective adhesion. The
10 cardiac and pulmonary fibroblasts were exposed to fetal bovine serum, a known nonspecific
11 mitogen, which resulted in cell proliferation in association with intracellular free radical
12 generation in both types of fibroblasts. Exposure to SOD resulted in significant reductions in
13 intracellular superoxide anion content and cell density for both types of fibroblasts. The cardiac
14 fibroblast proliferation was stimulated by exposure to cerium at 0.5 μM for 96 hours, whereas
15 lung fibroblast proliferation was not stimulated by 0.5 μM cerium exposure for 96 hours. These
16 data indicate that cardiac fibroblasts may be more sensitive to cerium exposure than lung
17 fibroblasts.

18 Du et al. (2001) exposed packed human erythrocytes to cerium chloride at concentrations
19 ranging from 4.9×10^{-5} to 3.9×10^{-3} M Ce^{3+} to investigate the aggregation of membrane
20 proteins after exposure to cerium. At the highest concentration tested, 3.9×10^{-3} M, aggregation
21 of membrane proteins was clearly evident, with aggregation increasing gradually with increasing
22 concentration from 4.9×10^{-5} to 3.9×10^{-3} M. Further analysis, using SDS-PAGE of the
23 membrane proteins and light scattering measurements, showed that the membrane protein
24 aggregation was mainly due to non-covalent cross-linking and to a lesser extent oxidative cross-
25 linking through disulfide bond formation.

26 27 *Nanoparticle studies*

28 In addition to the above in vitro studies, data on nano-sized cerium particles provide
29 information on absorption and cytotoxicity relevant to the mode of action. Limbach et al. (2005)
30 measured the uptake of cerium oxide nanoparticles at four different sizes, 20–50, 40–80, 80–150,
31 and 250–500 nm, into cultured human lung fibroblasts. The cultured fibroblasts absorbed the
32 nanoparticle cerium linearly with exposure time, with absorption occurring at concentrations as
33 low as 100 ng/g. Particles were not found outside of the vesicles or flowing freely in the
34 cytoplasm and were present exclusively in the form of agglomerates. The size of the cerium
35 nanoparticle greatly affected the amount of cerium incorporated into the cell, with better
36 absorption of larger nanoparticles. Particle size was a more important factor in absorption than
37 particle number and total surface area (Limbach et al., 2005).

1 Brunner et al. (2006) evaluated the cytotoxicity of nanoparticle CeO₂ to human
2 mesothelioma and rodent fibroblast cell lines by measuring metabolic activity and cell
3 proliferation. Cerium oxide was tested at exposures of 0, 3.75, 7.5, and 15 ppm for 6 days and 0,
4 7.5, 15, and 30 ppm for 3 days. The cerium oxide particles had a specific surface area derived
5 particle size of 6 nm and a hydrodynamic particle size of 19 nm, a specific surface area of
6 $124 \pm 3\% \text{ m}^2/\text{g}$, and a GSD of 1.49. Metabolic activity was spectroscopically measured as the
7 total mitochondrial activity through the conversion of the leuko form of a formazan-type dye to
8 the active dye, and cell proliferation was measured by the total DNA content of the cells
9 measured spectroscopically after converting DNA with an intercalating dye into a highly
10 fluorescent complex (Brunner et al., 2006). The human mesothelioma cells were more sensitive
11 to CeO₂ than the rodent fibroblast cells, with cell activity and DNA content decreased
12 approximately 50% after 3 days. The mesothelioma and fibroblasts were, however, not
13 completely killed at 30 ppm. After 6 days, the cell activity was not significantly altered and
14 DNA content was increased slightly in the mesothelioma cells; in the rat fibroblasts, the DNA
15 content increased slightly and the cell activity was not significantly affected.

16 The impact of a model water dispersion of 7 nm CeO₂ nanoparticles (specific surface
17 area of $400 \text{ m}^2/\text{g}$) on *E. coli* and cytotoxicity, assessed by counting colony-forming units (CFUs),
18 was investigated at concentrations ranging from 0 to 730 mg/L by Thill et al. (2006). Cerium
19 was almost completely adsorbed to the bacteria cell surface at a concentration of approximately
20 30 mg/L, and above this concentration an increasing amount of cerium was found in the
21 supernatant with a maximum adsorbed concentration of approximately 48 mg/L. The adsorption
22 appeared to be due to electrostatic attraction between the cerium oxide nanoparticles and the cell
23 membrane. The percentage of CFUs was strongly affected as the CeO₂ nanoparticle
24 concentration increased, with a 50% survival rate at around 5 mg/L and no survival above
25 230 mg/L. Thill et al. (2006) also showed that the speciation of the adsorbed cerium was
26 modified and the nanoparticles reduced. The study authors concluded that direct contact
27 between the *E. coli* and the CeO₂ nanoparticles needs to be established for CeO₂ cytotoxicity to
28 occur and that the reduction of the nanoparticles occurs at or close to the surface of the bacteria
29 and may be associated with cytotoxicity.

30 The toxicity of cerium nanoparticles was also investigated using the human
31 bronchioalveolar carcinoma-derived cell line A549 (Lin et al., 2006). The A549 cell line was
32 exposed to $20 \pm 3 \text{ nm}$ cerium oxide nanoparticles at 0, 3.5, 10.5, or 23.3 $\mu\text{g}/\text{mL}$ for 24, 48, or
33 72 hours. The cells were evaluated for cytotoxicity, as well as for intracellular reactive oxygen
34 species generation, lactate dehydrogenase (LDH) activity, dichlorodihydrofluorescein diacetate
35 (DCF) fluorescence, GSH levels, α -tocopherol levels, and malondialdehyde (MDA) levels. Cell
36 viability was decreased at all three dose levels and exposure durations and followed a dose- and
37 time-dependent decrease, with viability at 3.5, 10.5, and 23.3 $\mu\text{g}/\text{mL}$ at 72 hours decreased 12,

1 22, and 46%, respectively. LDH levels, indicative of cell membrane damage, increased 15, 32,
2 and 71% at 24, 48, and 72 hours, respectively, and were greatest at 23.3 $\mu\text{g}/\text{mL}$. DCF
3 fluorescence, indicative of oxidative stress, increased 70, 139, and 181% after exposure to 3.5,
4 10.5, and 23.3 $\mu\text{g}/\text{mL}$ cerium oxide, respectively. Antioxidant levels were decreased, with
5 cellular GSH levels reaching a maximal decrease of approximately 70% at 48 hours, with a
6 possible recovery of GSH levels at 72 hours, and α -tocopherol levels decreased 38, 76, and 88%
7 at 3.5, 10.5, and 23.3 $\mu\text{g}/\text{mL}$, respectively. MDA levels, indicative of lipid peroxidation, were
8 significantly increased in a dose- and time-dependent manner in the 3.5, 10.5, and 23.3 $\mu\text{g}/\text{mL}$
9 dose groups at 48 and 72 hours. Lin et al. (2006) demonstrated the induction of significant
10 oxidative stress at levels of 3.5, 10.5, and 23.3 $\mu\text{g}/\text{mL}$ of 20 nm cerium oxide particles. The
11 elevated reactive oxygen species levels, increased lipid peroxidation, increased membrane
12 damage, and reduced antioxidant levels are evidence of the increased oxidative stress from
13 cerium oxide nanoparticle exposure.

14 In addition to the evidence of cytotoxicity, there is evidence of neuroprotective and
15 cardioprotective effects of nanoparticle cerium exposure. Schubert et al. (2006) exposed the
16 HT22 hippocampal nerve cell line, derived from the rodent nervous system, to CeO_2
17 nanoparticles and monitored the intracellular generation of reactive oxygen species with a
18 nonfluorescent compound that fluoresces when in contact with reactive oxygen species. The
19 cerium oxide nanoparticles were characterized as single, monodisperse crystals. The CeO_2
20 nanoparticles were, for the most part, nontoxic to the HT22 cell line, with the exception of the
21 1 μm CeO_2 nanoparticles at concentrations ≥ 20 $\mu\text{g}/\text{mL}$. The 6 and 12 nm, as well as 1 μm ,
22 cerium particles were protective of oxidative stress to HT22 cells, and a difference in level of
23 protection offered by the 6 nm, 12 nm, and 1 μm sized particles could not be produced. It was
24 also shown that 12 nm CeO_2 particles at 20 and 200 $\mu\text{g}/\text{mL}$ were able to rapidly reduce pools of
25 reactive oxygen species formed by the cells after 8 hours of exposure to glutamate. Cerium
26 nitrate and cerium chloride, when tested at the same concentrations as cerium oxide, displayed
27 no protective effects. Schubert et al. (2006) were able to provide evidence that cerium oxide
28 nanoparticles have antioxidant properties that promote nerve cell survival under oxidative stress
29 conditions.

30 The cardioprotective effects of nano-sized CeO_2 were evident in cultured cells from MCP
31 mice administered 15 nmol of 7 nm CeO_2 intravenously twice a week for two weeks (Niu et al.,
32 2007). The cardiac-specific expression of monocyte chemoattractant protein (MCP-1) in mice
33 causes ischemic cardiomyopathy. The treatment of MCP mice with nanoparticle CeO_2 inhibited
34 monocyte/macrophage infiltration into the myocardial interstitial space, suppressed
35 proinflammatory cytokine production in the myocardium, and limited myocardial oxidative
36 stress.

37

1 **4.5.2. Ex Vivo Studies**

2 Manju et al. (2003) utilized isolated papillary muscle in an effort to assess the mechanical
3 response of the myocardium to varying levels of cerium. Cerium chloride concentrations of 0,
4 0.1, 0.5, 1, 5, 10, 20, 50, and 100 μM were applied to papillary muscle isolated from Sprague-
5 Dawley rats and the force of contraction was recorded using a force transducer. The role of SOD
6 was also investigated by superfusing the muscle with SOD prior to cerium chloride exposure and
7 recording of contractile force. A statistically significant ($p \leq 0.01$) reduction in contractile force
8 was evident at cerium chloride exposures as low as 0.1 μM , with the lowest dose reducing the
9 force of contraction approximately 15% (Manju et al., 2003). Complete recovery of contractile
10 force was apparent at or below 0.5 μM when cerium exposure was removed. The addition of
11 SOD completely inhibited the effect of cerium exposure, up to 5 μM , on the contractile force of
12 the isolated papillary muscle. These results provide further evidence for the involvement of
13 reactive oxygen species in the effects of cerium on the heart.
14

15 **4.6. SYNTHESIS OF MAJOR NONCANCER EFFECTS**

16 **4.6.1. Oral**

17 No studies evaluating the oral toxicity of cerium in humans were located, but an
18 association between exposure to cerium in food and the development of endomyocardial fibrosis
19 has been suggested (Eapen et al., 1998; Kutty et al., 1996; Valiathan et al., 1989). Long-term
20 studies in animals are limited to a 13-month drinking water study in rats, which investigated the
21 effects of a single dose group (Kumar et al., 1996), a 6-month drinking water study in rabbits in
22 which the administered dose consisted of a mixture of rare earth chlorides (Kartha et al., 1998), a
23 12-week dietary study in mice (Kawagoe et al., 2005), and a 105-day gavage study in rats
24 (Cheng et al., 2000). The study of Kumar et al. (1998) suggested that cerium may increase the
25 levels of collagen in the heart, while the findings of Kartha et al. (1998) suggested that cerium
26 may intensify the adverse cardiac effects of magnesium deficiency. Kawagoe et al. (2005)
27 suggested that cerium may increase oxidative stress in tissues of mice, and Cheng et al. (2000)
28 reported that cerium increased the oxygen affinity of hemoglobin in rats.

29 A single dose study in mice observed splenic lesions and GI irritation in treated mice
30 (Stineman et al., 1978). Other acute oral data consist of a report of piloerection, hunched
31 posture, unsteady gait, and dark coloring of eyes in male and female rats (along with lethargy,
32 abnormal respiration, and prostration in females only) given single gavage doses of 5,000 mg/kg
33 cerium as cerium sulfide (Rhodia Inc., 1998); a 7-day LD_{50} in mice of 1,291 mg/kg cerium as the
34 chloride/citrate complex (Stineman et al., 1978); and an LD_{50} of 1,178 mg/kg for cerium nitrate
35 and 622 mg/kg for cerium oxide, also in mice (Ji and Cui, 1988).

36 The few long-term studies available in animals identify cardiac tissue and hemoglobin
37 oxygen affinity as possible adverse health effects, but the animal studies are of limited scope and

1 insufficient duration and experimental design. Toxicokinetics data have shown that orally
2 administered cerium compounds are poorly absorbed, and this may be reflected in the results of
3 the studies conducted with these compounds by the oral route of exposure.

4 5 **4.6.2. Inhalation**

6 Inhalation data in humans consist of reports describing numerous cases of workers who
7 developed pneumoconiosis associated with accumulation of cerium in the lungs after prolonged
8 occupational exposure to cerium fumes or dust (Yoon et al., 2005; Porru et al., 2001; McDonald
9 et al., 1995; Paireon et al., 1995, 1994; Sulotto et al., 1986; Vogt et al., 1986; Pietra et al., 1985;
10 Vocaturo et al., 1983; Sabbioni et al., 1982; Husain et al., 1980; Kappenberger and Bühlmann,
11 1975; Heuck and Hoschek, 1968). In these cases, the exposure was to cerium oxide, and cerium-
12 induced pneumoconiosis was characterized by accumulation of cerium particles (and other rare
13 earth particles) in the lungs and lymphoreticular system. Exposure was not quantified in any of
14 these cases. The available data in humans are inadequate to identify potentially sensitive
15 subgroups.

16 Information regarding long-term inhalation exposure in animals is derived from a single
17 subchronic study in rats (BRL, 1994). Sprague-Dawley rats were exposed nose only to cerium
18 oxide aerosol 6 hours/day, 5 days/week for 13 weeks. Endpoints evaluated included a functional
19 observational battery, hematology and clinical chemistry, urinalysis, and gross and microscopic
20 morphology of tissues. The results revealed statistically significant increases in absolute and
21 differential neutrophil counts in the blood, treatment-related increases in the absolute and
22 relative weight of the lungs in both males and females dosed at 50.5 and 507.5 mg/m³ and in the
23 relative spleen weight of male rats at 507.5 mg/m³, discoloration or pale areas and uncollapsed
24 parenchyma in the lungs of male and female rats at ≥50 mg/m³ and pale foci in female rats at
25 5 mg/m³, and dose-related alveolar epithelial and lymphoid hyperplasia and pigment
26 accumulation in the lungs, lymph nodes, and larynx of males and females at ≥5 mg/m³. The
27 lowest exposure level, 5 mg/m³, was a LOAEL for lymphoid hyperplasia of the lymph nodes. A
28 NOAEL was not identified.

29 Acute inhalation data are limited to the determination of an LC₅₀ greater than 50 mg/m³
30 for cerium oxide in rats (Rhone-Poulenc, 1983) and a report of mild histologic alterations,
31 including slight connective tissue and collagen fiber development and increased lymphocyte
32 numbers, in the lung, as well as the accumulation of macrophages and large, multinucleated cells
33 in the spleen, of rats that received an intratracheal instillation of cerium oxide 8 months prior to
34 examination (Mogilevskaya and Raikhlin, 1967).

35 36 **4.6.3. Mode-of-Action Information**

37 **4.6.3.1. Respiratory Tissues**

1 The mode of action for cerium toxicity following chronic human inhalation exposures is
2 uncertain, since limited pathological data are available from the human case reports. In animals,
3 the observed pathology has been attributed to immune responses to cerium dust loads, which
4 overwhelmed innate pulmonary clearance mechanisms, namely clearance by pulmonary
5 macrophages (BRL, 1994). The accumulation of insoluble cerium particles in the respiratory
6 tract of humans and animals following chronic and subchronic inhalation exposures,
7 respectively, suggests that impaired clearance may influence pulmonary toxicity for both
8 species. In animals, correspondence of pulmonary and lymphoid hyperplasia with the
9 accumulation of cerium treatment-related pigmentation in the same tissues suggests that the
10 mode of action for cerium inhalation toxicity may be mediated by cytokine and fibrogenic
11 effects resulting from pulmonary macrophage activation followed by macrophage
12 immobilization.

13 The concept of dust overloading of the lungs refers to inhalation exposures that are
14 sufficiently intense as to overwhelm the pulmonary clearance mechanisms, specifically,
15 macrophagic phagocytosis in the alveolar spaces with mucociliary escalation in the bronchial
16 airways (Morrow, 1988). The concept of particle overload applies, specifically, to particles of
17 relatively low cytotoxicity that secondarily induce pulmonary toxicity via chronic activation of
18 immune-responsive cells (Oberdorster, 1995). In addition to immune cellular activation, the
19 uncleared particles may traverse the pulmonary epithelial boundary to the interstitial spaces,
20 where fibrogenesis may occur (Oberdorster, 1995).

21 Cullen et al. (2000) demonstrated overwhelmed pulmonary clearance in male Wistar rats
22 following inhalation exposure to another poorly soluble metal oxide (titanium dioxide; TiO₂) in
23 which the particles were of similar size and concentration to that used for cerium oxide in the
24 BRL (1994) study. Cullen et al. (2000) investigated pulmonary effects in male Wistar rats
25 following the 4- and 7-month inhalation exposure of TiO₂ particles with a MMAD of 2.1 μm
26 administered at 25 mg/m³ for 209 calendar days and 50 mg/m³ for 118 calendar days. The
27 lymph node burdens, measured in mg/day, for TiO₂ demonstrated the initiation of overload
28 retardation after approximately 50 days at 50 mg/m³ and 150 days at 25 mg/m³. In addition,
29 inhalation of TiO₂ led to an accumulation of pulmonary macrophages around dust deposition
30 sites, and most of the macrophages contained phagocytized dust particles. The results observed
31 in Cullen et al. (2000) for TiO₂ support the proposed overload of pulmonary clearance during the
32 13-week inhalation exposure to cerium oxide particles of similar size and solubility observed in
33 BRL (1994). Cullen et al. (2000) did not include exposure concentrations as low as the BRL
34 cerium oxide study (i.e., 5 mg/m³); however at the lowest concentration of TiO₂ examined,
35 overload was also observed, although time to overload was longer at the lower concentration
36 (140 days at 25 mg/m³). The demonstration of pulmonary overload by Cullen et al. (2000) in an
37 exposure time frame and at a concentration of TiO₂ particles with a similar MMAD to the cerium

1 oxide particles in the BRL (1994) study supports the proposed mode of action of overwhelmed
2 pulmonary clearance for the relatively insoluble cerium oxide.

3 Overwhelmed pulmonary clearance, as marked by increased translocation of dust to
4 lymph nodes, was observed for TiO₂ but not for barium sulfate (BaSO₄) under similar conditions
5 in the study by Cullen et al. (2000). Although pulmonary overload is a mode of action that may
6 apply to other relatively insoluble particles of low cytotoxicity, it is clear from the Cullen et al.
7 (2000) study that there are strong differences between exposure to the dust of different
8 chemicals, such that exposure to TiO₂ leads to overloading of the lung, whereas overloading was
9 not observed for BaSO₄.

10 Data were unavailable to definitively identify toxic mechanisms of cerium-induced nasal
11 and bronchial metaplasia and hyperplasia. In a review of lanthanide toxicity, Haley (1991)
12 proposed that rare earth metals, including cerium, may exert toxicity both from innate chemical
13 characteristics as well as pulmonary dust burden. A suggested mechanism explaining both
14 inflammatory and fibrogenic responses involves the activation of PAMs following dust overload.

15 Tissue-destructive release products of activated PAMs include acid hydrolases, elastases,
16 collagenases, and several reactive oxidant species (Haley, 1991). Additionally, the release of
17 fibroblast growth factor and fibronectin may stimulate the proliferation of fibroblasts, and PAMs
18 may release neutrophil attractant factors (Haley, 1991). The subsequently enlisted neutrophils
19 can release several oxidants and proteases that are known to result in connective tissue damage,
20 possibly stimulating fibrogenesis (Hunninghake et al., 1984). Although BRL (1994) did not
21 collect data on macrophage activation or the release of chemokines, the significant increase in
22 mature neutrophils in rats exposed to high levels of cerium observed in this study is consistent
23 with stimulation of neutrophils by PAMs. The exposure-related increase in pigment
24 accumulation and lymphoid hyperplasia in lymph nodes draining the lungs (i.e., bronchial and
25 mediastinal lymph nodes) and the absence of significant effects in pancreatic and mandibular
26 lymph nodes further supports the role of pulmonary macrophages.

27 The immobilization of PAMs by excessive cerium dust loads, as hypothesized by
28 Morrow (1992, 1988), results in macrophages carrying a heavy cerium load to lose the ability to
29 move either toward the mucociliary escalator system, for subsequent clearance to the gut, or to
30 the lymphoid vasculature. A dense tissue population of activated, yet immobile, macrophages
31 may serve to induce significant cell damage by effectively increasing the concentration of
32 inflammatory cytokines and fibrogenic growth factors within the pulmonary epithelium.
33 Reduction in the ability to clear insoluble cerium from the lung spaces due to macrophage
34 immobilization is consistent with reports of extracellular presence of pigment in mid- and high-
35 dose rats relative to low-dose animals (BRL, 1994). The presence of inflammatory cytokines
36 and growth factors was not examined.

1 The ability of immobilized macrophages to induce toxicity via concentration of immune
2 signaling requires minimal cytotoxicity of the macrophages themselves. In vitro experiments
3 have shown that cytotoxicity of rat pulmonary macrophages is high for soluble cerium (cerium
4 chloride) (Lizon and Fritsch, 1999; Palmer et al., 1987) but quite low for the insoluble salt
5 (cerium oxide) (Palmer et al., 1987). This suggests that the pulmonary responses to inhaled
6 cerium reported in human cases and animal bioassays (which predominantly involved insoluble
7 cerium oxide) may not have caused significant macrophage cell death. This finding is consistent
8 with pathological findings in human case reports, animal studies, and in vivo assays of PAMs in
9 which cerium-related granule pigmentation was visible in the lung and lymphoid tissues,
10 suggesting that the cerium sequestration and concentration occurred in viable PAMs.

11 The precipitation and concentration of both soluble and insoluble cerium within cytosolic
12 lysosomes has been demonstrated (Berry et al., 1997; Berry, 1996). The high phosphate
13 concentration and extensive enzymatic hydrolysis activity (acid phosphatase) within lysosomes
14 was shown to effectively precipitate cerium as cerium phosphate. Rats exposed to 5 µg insoluble
15 cerium oxide via intratracheal dosage for 30 days displayed rat alveolar macrophages that
16 contained very fine needles or granules in the lysosomes (Berry et al., 1997). The needle or
17 granule inclusions contained both phosphorus and cerium. After 2 days of intratracheal dosage,
18 the rat alveolar macrophage lysosomes contained cerium particles approximately 1 µm in length
19 and displayed a crystalline structure. Rats exposed to soluble cerous chloride displayed fine
20 granules and needles in the lysosomes (Berry et al., 1997). Berry (1996) showed that cerium
21 precipitated with phosphorus in the lysosomes of hepatocytes and of bone marrow and splenic
22 macrophages of rats after intraperitoneal injections of cerium. Granular or needle-like
23 precipitate deposits were observed in the lysosomes of alveolar macrophages of rats exposed to
24 an aerosol solution of cerium chloride (Galle et al., 1992). The authors (Berry et al., 1997; Galle
25 et al., 1992) suggest that the precipitation and concentration of cerium within the lysosomes may
26 serve to inhibit diffusion of the metal to other tissues.

27 The presence of insoluble cerium particles in the alveolar macrophages of rats exposed
28 via the intratracheal route is consistent with the appearance of cerium-related particles within the
29 alveolar macrophages of a worker in an occupation associated with cerium oxide exposure
30 (McDonald et al., 1995). The particles in the human alveolar macrophages ranged from small,
31 pinpoint particles to oblong or needle-shaped. Pairon et al. (1995) revealed particles of
32 phosphorus, calcium, lanthanum, and cerium in human alveolar macrophages following
33 occupational exposure and identified interstitial macrophages that contained both phosphorus
34 and rare earth elements. Cerium was essentially always associated with phosphorus in the
35 biological samples studied by Pairon et al (1995).

36 The presence of the insoluble cerium particles in the rat alveolar macrophages (Berry et
37 al., 1997) and in the lysosomes of hepatocytes and splenic macrophages (Berry, 1996) is also

1 consistent with the involvement of macrophages in the appearance of cerium-related
2 pigmentation in the rat lung, liver, and spleen following inhalation exposures (BRL, 1994).

3 Shivakumar and Nair (1991) have shown that cerium exposure resulted in the inhibition
4 of protein synthesis in human lung fibroblasts, while lung fibroblast proliferation was not
5 stimulated by 0.5 μ M cerium exposure (Nair et al., 2003). However, cell types other than
6 fibroblasts may be involved in the inflammatory response associated with cytokine and growth
7 factor release that leads to a fibrotic response (Nair et al., 2003).

8 The hypothesized mode of action for the pulmonary effects observed following cerium
9 oxide exposure is the overloading of the PAMs by cerium oxide particles, leading to the release
10 of inflammatory cytokines and fibrogenic growth factors and subsequent cell damage.

11 12 **4.6.3.2. Other Tissues**

13 No unifying mode of action for cerium in non-respiratory tissues was identified in the
14 studies available. Studies conducted utilized primarily parenteral routes of exposure and suggest
15 that the production of free radicals may be involved in cerium toxicity, as well as actions at the
16 RNA transcription level. However, details of potential mode(s) of action in non-respiratory
17 tissues are lacking.

18 Cerium administered subcutaneously was found to have minor effects on some tests of
19 spontaneous motor behavior in young adult mice (Morganti et al., 1980, 1978; Stineman et al.,
20 1978) and in mice exposed in utero (D'Agostino et al., 1982, 1978a, b); however, a mode of
21 action was not discussed or apparent in these studies. Interestingly, behavioral measures were
22 not correlated with cerium levels in tissues, which suggested an unknown, indirect mode of
23 action involving a biological chain of events (Morganti et al., 1978).

24 Liver effects have been reported in several studies in rats and mice following i.v.
25 administration of cerium compounds. The effect is characterized by fatty liver, fatty
26 degeneration, and necrosis (Magnusson, 1962; Lombardi and Recknagel, 1962). Female rats
27 appeared more sensitive than male rats (Wiener-Schmuck et al., 1990; Magnusson, 1962).
28 Incubation of hepatocytes from female rats with cerium did not result in damage to the cells
29 (Wiener-Schmuck et al., 1990), suggesting that the damage is not a result of direct contact of
30 cerium with the hepatocytes but that cerium exposure may trigger a series of events that
31 ultimately result in liver damage. This seems consistent with the results of a study in two strains
32 of mice having different sensitivities to cerium toxicity that showed that cerium levels in the
33 liver of the more resistant strain were 50% higher than those in the more sensitive strain (Arvela
34 et al., 1991). The latter study and an additional study by Salonpää et al. (1992) presented
35 evidence suggesting that some association exists between the development of liver damage and
36 COH induction. Enzyme induction was accompanied by increased expression of CYP2A5
37 mRNA, but the mechanism by which cerium increases CYP2A5 expression is unknown.

1 Cerium exposure has been found to increase the levels of collagen in the heart of animals
2 in an oral study (Kumar et al., 1996) and following i.v. dosing (Kumar and Shivakumar, 1998).
3 Cerium exposure decreased collagen degradation and increased the rate of deposition of newly
4 synthesized collagen in cardiac tissue of rats (Kumar and Shivakumar, 1998). The increase in
5 collagen synthesis is consistent with a marked increase in mRNA in cardiac fibroblasts incubated
6 with cerium (Shivakumar et al., 1992), pointing to an action at the level of transcription.
7 Cerium-induced production of free radicals was also associated with stimulation of fibroblast
8 proliferation in studies in vitro (Nair et al., 2003; Preeta and Nair, 1999). While cerium-induced
9 collagen proliferation can explain the occurrence of cardiac fibrosis, the mechanism by which
10 cerium triggers this event at the molecular level remains unknown.

11 In vitro studies demonstrated that low cardiac fibroblast proliferation is stimulated by
12 exposure to cerium (0.5 µM) (Nair et al., 2003), that low concentrations of cerium stimulate
13 cardiac fibroblast proliferation in association with an increase in intracellular generation of free
14 radicals (Preeta and Nair, 1999), and that cerium exposure results in the inhibition of protein
15 synthesis in rat heart cells (Shivakumar and Nair, 1991). Kuruvilla and Kartha (2006)
16 demonstrated that cerium reduced the incorporation of [H3]-thymidine into DNA of cardiac
17 fibroblasts grown in endocardial endothelial cells conditioned medium. These results suggest
18 that the cardiac lesions in endomyocardial fibrosis may result from the direct stimulation, and
19 not through toxic effects on the endocardial endothelium, of subendomyocardial fibroblasts by
20 cerium (Kuruvilla and Kartha, 2006).

21 Cerium inhibited contraction of isolated rat ventricular papillary muscle, an effect that
22 could be partially prevented by the free radical scavenger SOD (Manju et al., 2003), which adds
23 support for the involvement of free radicals in the cerium-induced cardiac effects.
24

25 **4.7. EVALUATION OF CARCINOGENICITY**

26 **4.7.1. Summary of Overall Weight of Evidence**

27 Data were unavailable regarding the carcinogenicity of cerium compounds in humans or
28 experimental animals. In accordance with U.S. EPA (2005a) *Guidelines for Carcinogen Risk*
29 *Assessment*, there is “inadequate information to assess the carcinogenic potential” of cerium in
30 humans.
31

32 **4.7.2. Synthesis of Human, Animal, and Other Supporting Evidence**

33 No relevant human or animal data are available. In addition, the available information is
34 insufficient to ascertain the mutagenicity of cerium compounds. The NTP has recently begun an
35 evaluation of the chronic inhalation toxicity of cerium oxide, including a cancer bioassay. The
36 date of completion and public availability of the results is unknown but may be expected in
37 2009.

1 A study in various strains of *S. typhimurium* demonstrated negative evidence of
2 mutagenicity under the conditions of the assay (Shimizu et al., 1985). Cerium chloride did not
3 induce DNA damage in two strains of *B. subtilis* by using the rec-assay (Nishioka, 1975), but
4 cerium nitrate was reported to induce chromosomal breaks and reduce the mitotic index in rat
5 bone marrow in vivo, and cerium sulfate was reported to cause differential destaining of
6 chromosomal segments in plants (Sharma and Talukder, 1987).

7 8 **4.8. SUSCEPTIBLE POPULATIONS AND LIFE STAGES**

9 **4.8.1. Possible Childhood Susceptibility**

10 No studies were located regarding possible childhood susceptibility to cerium
11 compounds. Kutty et al. (1996) stated that high cerium soil concentrations and magnesium
12 deficiency during childhood may lead to endomyocardial fibrosis, although causation has not
13 been established. Increased GI uptake of cerium in preweaning, suckling animals compared with
14 adults is apparent, due primarily to pinocytosis in intestinal cells, but it is unclear if this has any
15 toxic consequences, since the cerium remains in the intestinal cells, may be minimally available
16 systemically, and is eventually eliminated in the feces as the intestinal cells die and are replaced
17 (Kargacin and Landeka, 1990; Kostial et al., 1989a, b; Inaba and Lengemann, 1972).

18 19 **4.8.2. Possible Gender Differences**

20 No information was located regarding gender differences in humans in response to
21 exposure to cerium compounds. Information addressing gender differences in animals is from a
22 study by Magnusson (1962) in which i.v. administration of cerium to rats resulted in noticeably
23 more severe adverse liver effects in females than in males. This was confirmed in another study
24 in rats also treated intravenously with cerium (Wiener-Schmuck et al., 1990). There is no
25 explanation for this gender-related difference in susceptibility, and it is unknown whether it also
26 applies to endpoints other than the liver. Additionally, it appears that female rats may be more
27 susceptible to the hematological changes (Table 4-1) observed as the result of cerium oxide
28 inhalation (BRL, 1994).

29 30 **4.8.3. Possible Susceptible Populations**

31 A variety of health effects could result from the accumulation of a sufficient amount of
32 persistently retained particles in the lung (Morrow, 1988). Smoking has suppressive effects on
33 pulmonary clearance in humans (Morrow et al., 1992), and the co-exposure of cerium with
34 cigarette smoke, or particulates that accumulate in the lungs, may potentially lead to more severe
35 adverse pulmonary effects in exposed populations. Chen et al. (2006) found that pulmonary
36 inflammation, in this case from lipopolysaccharide treatment, may play an integral role in
37 enhancing the extrapulmonary translocation of particles.

1 Particle size is another factor that influences overload and toxicity that may be
2 particularly relevant for human exposure if human exposure is associated with fumes from
3 carbon arc lamps, as fumes include smaller particles from the low nm range to $<1\ \mu\text{m}$
4 aggregates. Impaired lung clearance and lung effects in rats exposed to ultrafine particles (<100
5 nm) occur at lower mass concentrations than in rats exposed to fine particles ($<10\ \mu\text{m}$) (Baan et
6 al., 2006). Translocation of particles to the interstitium is a function of number of particles, and
7 appears to be dependent on the dose and particle size (Ferin et al., 1992). Excessive
8 translocation into the interstitium may cause damage to epithelial cells, pulmonary edema, and
9 eventual fibrosis (Ferin 1994, Oberdorster et al., 1990). Populations exposed to ultra-fine cerium
10 oxide may have a higher than expected toxicity when compared to cerium oxide particles of a
11 larger size, as ultrafine particles have higher than expected toxicity when compared to similar
12 particles of a larger size (Ferin, 1994).

13
14

5. DOSE-RESPONSE ASSESSMENTS

5.1. ORAL REFERENCE DOSE (RfD)

5.1.1. Choice of Principal Study and Critical Effect—with Rationale and Justification

The available human and animal studies demonstrate that cerium may have an effect on cardiac tissue and hemoglobin oxygen affinity. However, an RfD for cerium was not derived because the available studies were not suitable for quantitation of effects for various reasons, including unknown exposure concentrations in the available human studies, lack of a dose-response, uncertain biological significance (e.g., increased oxygen affinity for hemoglobin, changes in measures of oxidative stress), and inadequate study design (e.g., effects noted only under conditions of a restricted diet).

An association between exposure to cerium in food and the development of endomyocardial fibrosis has been suggested (Eapen et al., 1998; Kutty et al., 1996; Valiathan et al., 1989). In addition, Gómez-Aracena et al. (2006) suggest a relationship between chronic cerium exposures, characterized by cerium toenail concentrations, and increased risk of acute myocardial infarction. The data set for long-term animal studies consists of a 13-month drinking water study in rats (Kumar et al., 1996), a 6-month drinking water study in rabbits (Kartha et al., 1998), a 12-week dietary study in mice (Kawagoe et al., 2005), and a 105-day gavage study in rats (Cheng et al., 2000).

Kumar et al. (1996) demonstrated increased, highly variable cerium levels in cardiac tissue. Cerium chloride-treated rats, relative to untreated controls, had an increased level of collagen in the cardiac tissue, with an enhanced effect in rats fed a magnesium-deficient diet. This study suggested that cerium might increase the levels of collagen in the heart. This study was conducted on a small number of rats at one dose level (control and dosed rats) and evaluated few endpoints, and the observed effects were highly variable and not statistically significant.

Kartha et al. (1998) suggested that cerium chloride may intensify the adverse cardiac effects of magnesium deficiency. Cardiac lesions were apparent in 6/10 rabbits fed a magnesium-deficient diet with no cerium exposure and 9/10 rabbits fed a magnesium-deficient diet with cerium exposure. Rabbits fed magnesium-restricted diets, treated with or without cerium, showed endocardial, subendocardial, interstitial, and perivascular fibrosis and the lesions were more severe in those with cerium added to the drinking water. Cardiac lesions were absent from the groups fed the normal magnesium diet regardless of whether they consumed water with or without cerium. This study was conducted in an adequate number of animals but used only one dose group. The authors reported that cerium may intensify the cardiotoxicity associated with a magnesium-deficient or restricted diet but did not elicit a cardiac effect when tested under conditions of a normal magnesium diet.

1 Cheng et al. (2000) reported that cerium chloride exposure in rats produced a slight
2 increase of hemoglobin content in erythrocytes after 40 days of treatment with an even greater
3 increase in hemoglobin content after 80 days of exposure. The effect on the oxygen affinity of
4 hemoglobin was demonstrated by altered oxygen saturation curves for the dosed rats compared
5 to control rats. Hemoglobin in cerium-treated rats exhibited altered oxygen affinity up to 80
6 days of exposure, demonstrated by increased affinity up to 10 mm Hg and a double sigmoidal
7 curve for 40-day rats and increased affinity above 20 mm Hg for 80-day rats.

8 Kawagoe et al. (2005) demonstrated that cerium chloride statistically significantly
9 decreased liver lipoperoxide levels, increased liver GSH levels and liver MT activity, and
10 decreased plasma SOD activity in mice. Cerium concentrations in the kidney, liver, lung, and
11 spleen were statistically significantly elevated relative to controls, with the lung and spleen
12 containing the highest levels. The study authors suggested that the endpoints showing changes
13 as a result of cerium exposure in this study are indicators of reactive oxygen species generation
14 and resultant oxidative stress but indicated that their toxicological significance was uncertain.
15 The authors did not report any other effects in the liver.

16 17 **5.2. INHALATION REFERENCE CONCENTRATION (RfC)**

18 **5.2.1. Choice of Principal Study and Critical Effect—with Rationale and Justification**

19 Exposure to cerium compounds in the environment is most likely through cerium (ceric)
20 oxide. There are numerous case reports of workers who developed pneumoconiosis or
21 interstitial lung disease associated with the accumulation of cerium particles in the lungs after
22 prolonged occupational exposure to cerium fumes or dust (Yoon et al., 2005; Porru et al., 2001;
23 McDonald et al., 1995; Pairon et al., 1995, 1994; Sulotto et al., 1986; Vogt et al., 1986; Pietra et
24 al., 1985; Vocaturo et al., 1983; Sabbioni et al., 1982; Husain et al., 1980; Kappenberger and
25 Bühlmann, 1975; Heuck and Hoschek, 1968). In these cases, the inhalation exposure was
26 primarily to cerium oxide, and cerium-induced pneumoconiosis was characterized by
27 accumulation of cerium particles (and other rare earth particles) in the lungs and lymphoreticular
28 system. Cerium exposure was associated with interstitial, peribronchial, and perivascular
29 fibrosis, a restriction of respiratory function and/or pulmonary hypertension (Porru et al., 2001;
30 Pairon et al., 1995, 1994; Vogt et al., 1986; Vocaturo et al., 1983; Sabbioni et al., 1982). The
31 available human studies were not selected for the derivation of an inhalation RfC because the
32 cerium exposures were not available in any of the case reports.

33 Information regarding long-term inhalation exposure in animals is derived from a single
34 subchronic study in rats (BRL, 1994), which was chosen as the principal study. Sprague-Dawley
35 rats (n = 30) were exposed nose only to cerium oxide aerosol 6 hours/day, 5 days/week for
36 13 weeks. Endpoints evaluated included a functional observational battery, hematology and
37 clinical chemistry, urinalysis, and gross and microscopic morphology of tissues, as discussed in

1 Section 4.2.2. The following effects were observed: (1) a statistically significant increase in
2 absolute and differential neutrophil counts in the blood in both males and females at 50.5 and
3 507.5 mg/m³ at both 6 and 13 weeks; (2) a statistically significant increase in absolute and
4 relative lung weight in both male and female rats at 50.5 and 507.5 mg/m³; (3) relative spleen
5 weight was statistically significantly increased in male rats at 507.5 mg/m³; (4) discoloration or
6 pale areas and uncollapsed parenchyma in the lungs in male and female rats at 50.5 and
7 507.5 mg/m³, with pale foci in female rats at 5 mg/m³; (5) an increased incidence of lymphoid
8 hyperplasia, alveolar epithelial hyperplasia, and pigment accumulation in the lungs at
9 507.5 mg/m³, 50.5 mg/m³, and 5 mg/m³, respectively, in both males and females; (6) an
10 increased incidence of lymphoid hyperplasia and pigment accumulation in the bronchial lymph
11 nodes of both males and females at 5 mg/m³; and (7) an increased incidence of metaplasia and
12 pigment accumulation in the larynx in males at 50.5 mg/m³ and 5 mg/m³, respectively, and in
13 females at 50.5 mg/m³.

14 The BRL (1994) study is an unpublished study; accordingly, it was externally peer
15 reviewed by EPA in August 2006 to establish its suitability for a dose-response evaluation of
16 cerium toxicity.

17 Based on the results of the human case reports, the BRL (1994) study, and the mode of
18 action analysis (Section 4.6.3.1), cerium toxicity may be the result of nonspecific stimulation of
19 PAMs that are activated and immobilized by the accumulation of insoluble cerium particles. The
20 subchronic BRL (1994) study in rats displayed increased lung weight; discoloration or pale
21 areas, pale foci, and uncollapsed parenchyma in the lungs; enlargement or pale discoloration of
22 the bronchial, mediastinal, and pancreatic lymph nodes; and dose-related alveolar epithelial and
23 lymphoid hyperplasia and pigment accumulation in the lungs and lymph nodes. These effects
24 are similar to the pneumoconiosis described in the human case reports, which were characterized
25 by the accumulation of cerium particles in the lungs and lymphoreticular system and histologic
26 effects throughout the lung. In addition, Hahn et al. (2001, 1999) demonstrated the retention of
27 cerium-aluminosilicate particles in the tracheobronchial lymph nodes of dogs for several years
28 following a single inhalation exposure.

29 Clearance of foreign substances from the lung by macrophages involves removal to the
30 stomach and GI tract, the lymphatics and lymph nodes, and the pulmonary vasculature (Witschi
31 and Last, 2001). As part of that process, macrophages that have phagocytized cerium particles
32 and have retained their mobility, would be removed from the lung and may accumulate in the
33 lymph nodes. The point in the neutrophilic response when normal clearance is overwhelmed
34 represents a shift to PAMs that have been activated and immobilized by the absorption of cerium
35 particles. This shift is reflected by the alveolar epithelial and lymphoid hyperplasia and pigment
36 accumulation in the lungs and lymph nodes of male and female rats (BRL, 1994). The
37 overloading of the PAMs by cerium, leading to the release of inflammatory cytokines and

1 fibrogenic growth factors, and the subsequent cell damage are the hypothesized mode of action
2 presented in Section 4.6.3.1. Therefore, the critical effect of cerium oxide exposure is increased
3 incidence of lymphoid hyperplasia in the bronchial lymph nodes of male and female rats, which
4 represents the most sensitive effect following cerium oxide exposure.

6 **5.2.2. Methods of Analysis—Including Models (PBTK, BMD, etc.)**

7 A NOAEL/LOAEL approach was used to derive the RfC for cerium oxide. Benchmark
8 dose (BMD) modeling was not utilized because the incidences of lymphoid hyperplasia in the
9 bronchial lymph nodes of male and female rats approached 100% at the lowest dose tested
10 (5 mg/m³) and were not amenable to modeling. Thus, the RfC is based on the LOAEL of
11 5 mg/m³ as the point of departure. Additionally, there was an increase in the incidence of
12 lymphoid and alveolar epithelial hyperplasia in the lung at 50.5 mg/m³. The selected point of
13 departure is considered to be protective of the pulmonary effects.

14 The human equivalent concentration (HEC) was calculated from the point of departure
15 by adjusting to a continuous exposure (24 hours a day, 7 days a week) and multiplying by a
16 dosimetric adjustment factor (DAF), which, in this case, was the regional deposited dose ratio
17 (RDDR) for the pulmonary region of the lung. Adjustment to a continuous exposure was
18 calculated as follows:

$$\begin{aligned} \text{LOAEL}_{\text{ADJ}} &= \text{LOAEL} \times (6 \text{ hours}) / (24 \text{ hours}) \times (5 \text{ days}) / (7 \text{ days}) \\ &= 5 \text{ mg/m}^3 \times 0.25 \times 0.71 \\ &= 0.89 \text{ mg/m}^3 \end{aligned}$$

24 The RDDR was calculated using the RDDR v.2.3 program (Table 5-1), as described in
25 *Methods for Derivation of Inhalation Reference Concentrations and Application of Inhalation*
26 *Dosimetry* (U.S. EPA, 1994b). The pulmonary region of the lung was selected as the deposition
27 site because the critical effect and the mode-of-action data indicate that the pulmonary region of
28 the lung is the initial location of cerium oxide toxicity.

29 The human parameters used were body weight, 70 kg; minute volume, 13.8 L;
30 extrathoracic surface area, 200 cm²; tracheobronchial surface area, 3,200 cm²; and pulmonary
31 surface area, 54.0 m². The parameters used for the rat were body weight, 345 g (calculated by
32 averaging the mean body weights for each dose group for the course of the experiment); minute
33 volume, 234.18 mL; extrathoracic surface area, 15 cm²; tracheobronchial surface area,
34 22.50 cm²; and pulmonary surface area, 0.34 m². The GSD for the cerium oxide particles in the
35 BRL (1994) study ranged from 1.8 to 1.9 and the MMAD for the particles was approximately
36 2.0 μm. The RDDRs calculated from the above model parameters vary depending upon the

1 deposition site in the lung (Table 5-1). The RDDR for pulmonary depositon is 0.536. The
 2 calculation is as follows:

3
 4 $LOAEL_{HEC} = LOAEL_{ADJ} \times RDDR$
 5 $LOAEL_{HEC} = 0.89 \text{ mg/m}^3 \times 0.536$
 6 $LOAEL_{HEC} = 0.48 \text{ mg/m}^3$

7
 8 **Table 5-1. Output from RDDR v.2.3 used in the analysis in Section 5.2.2**

Species		Body weight (g)	VE (ml)	Extrathoracic		Tracheobronchial		Pulmonary	
				SA (cm ²)	dep	SA (cm ²)	dep	SA (m ²)	dep
Rat		345	234.2	15.000	0.542	22.500	0.042	0.340	0.049
Human		70000	13800.0	200.000	0.352	3200.000	0.085	54.000	0.245
	Ratio	0.005	0.017	0.075	1.540	0.007	0.490	0.006	0.199
	RDDR			0.348		1.183		0.536	
				Thoracic		Total RT		Extrarespiratory	
				SA (cm ²)	dep	SA (cm ²)	dep	SA (m ²)	dep
Rat				0.342	0.091	0.344	0.632	345	0.632
Human				54.320	0.125	54.340	0.682	70000	0.682
	Ratio			0.006	0.724	0.006	0.927	0.005	0.927
	RDDR			0.738		2.487		3.191	

9
 10 MMAD = 2.00; Sigma g = 1.85.

11
 12
 13 **5.2.3. RfC Derivation—Including Application of Uncertainty Factors (UFs)**

14 The LOAEL_{HEC} value of 0.48 mg/m³ for lymphoid hyperplasia in the bronchial lymph
 15 nodes of male and female CD rats (BRL, 1994) was used to derive the RfC for cerium oxide. A
 16 total UF of 3,000 was applied to this point of departure: 3 for extrapolation from animals to
 17 humans (UF_A: animal to human), 10 for consideration of interindividual variability (UF_H: human
 18 variability), 10 for extrapolation from a subchronic study (UF_S), 3 for LOAEL-to-NOAEL
 19 extrapolation (UF_L), and 3 for database deficiencies (UF_D). The rationale for the application of
 20 the UFs is described below.

21 A factor of 3 was selected to account for uncertainties in extrapolating from rats to
 22 humans (UF_A). This value is adopted by convention where an adjustment from an animal-
 23 specific LOAEL_{ADJ} to a LOAEL_{HEC} has been incorporated. Application of a full UF of 10 would
 24 depend on two areas of uncertainty (i.e., toxicokinetic and toxicodynamic uncertainties). In this
 25 assessment, the toxicokinetic component is mostly addressed by the determination of a HEC as
 26 described in the RfC methodology (U.S. EPA, 1994b). The toxicodynamic uncertainty is also
 27 accounted for to a certain degree by the use of the applied dosimetry method.

28 A factor of 10 was used to account for variation in susceptibility among members of the
 29 human population (UF_H). Insufficient information is available to predict potential variability in
 30 susceptibility among the population to inhaled cerium oxide and cerium compounds.

1 A factor of 10 was used to account for uncertainty in extrapolating from a subchronic to
2 chronic (UF_S) exposure duration, since the BRL (1994) study, which was selected as the
3 principal study, is a subchronic study. The critical effect, increased incidence of lymphoid
4 hyperplasia in the bronchial lymph nodes, may be more pronounced at longer durations.

5 A factor of 3 was used to account for uncertainty in extrapolating from a LOAEL to a
6 NOAEL (UF_L) because the critical effect selected to determine the point of departure for this
7 analysis, lymphoid hyperplasia in the bronchial lymph nodes, represents a sensitive, precursor
8 effect that occurs early in the series of critical events leading to more severe effects in the lung.
9 Specifically, lymphoid hyperplasia in the bronchial lymph nodes represents the point at which
10 normal clearance of particles from the lung by alveolar macrophages becomes overwhelmed and
11 particles are no longer cleared effectively. This delayed clearance leads to increased
12 accumulation of cerium oxide particles in the respiratory tract, an inflammatory response, and
13 subsequent cell proliferation.

14 The involvement of additional lymph nodes (e.g., enlargement and discoloration of the
15 mediastinal lymph nodes) and other tissues (e.g., enlargement and discoloration of the pancreatic
16 lymph nodes), along with alveolar epithelial hyperplasia in the lung at higher exposure
17 concentrations, supports the hypothesis that the clearance process may be increasingly
18 overwhelmed at higher concentrations and that the point of departure for the derivation of the
19 RfC (i.e., the LOAEL) is based on a sensitive effect. The ability of cerium-oxide-exposed rats to
20 recover after exposure has not been studied, but would demonstrate the level of persistence of
21 the bronchial lymphoid hyperplasia observed in BRL (1994) study. Case studies in workers
22 exposed to cerium oxide (Yoon et al., 2005; Porru et al., 2001; McDonald et al., 1995; Pairen et
23 al., 1995, 1994; Sulotto et al., 1986; Vogt et al., 1986; Pietra et al., 1985; Vocaturo et al., 1983;
24 Sabbioni et al., 1982; Husain et al., 1980; Kappenberger and Bühlmann, 1975; Heuck and
25 Hoschek, 1968) have identified interstitial lung disease as a possible outcome of exposure to
26 cerium oxide in the workplace. The BRL (1994) study and proposed mode of action for cerium
27 oxide-induced lung effects supports the conclusion that lymphoid hyperplasia in the bronchial
28 lymph nodes of humans may be an early manifestation of the nonspecific response to cerium
29 oxide.

30 A UF of 3 was used to account for deficiencies in the cerium oxide database. The
31 database includes multiple case reports of inhalation exposure to workers and a single 13-week
32 subchronic inhalation study in rats. The effects from the subchronic rat inhalation study that are
33 used for the derivation for the RfC (i.e., bronchiolar lymph node hyperplasia) may be early
34 indicators of the more overt toxicity that is found in humans (i.e., interstitial lung disease)
35 exposed to cerium oxide in the workplace. The database does not include an exposure and
36 recovery study that could demonstrate the persistence or, conversely, the adaptive nature of the
37 lymphoid hyperplasia in the bronchial lymph nodes.

1 Toxicity via the inhalation route is expected to be a portal-of-entry effect. Cerium oxide
2 is a relatively insoluble metal oxide and absorption or translocation from the lung to the
3 circulation is expected to be minimal at low doses. The pulmonary effects observed in the
4 human case reports and in the BRL (1994) study are likely due to the physical deposition of
5 cerium oxide particles in the lung and the immunological reaction to the particles, and are not
6 due to a chemical reaction of cerium oxide with lung tissues. The lymphoid hyperplasia in the
7 bronchial lymph nodes is an immunological response to the cerium oxide particles and is not due
8 to cytotoxicity with regenerative cell growth. The observed immunological response is a portal-
9 of-entry effect and systemic circulation and effects are not expected because of the insoluble
10 nature of the cerium oxide particles.

11 In considering the impact of database deficiencies on the derivation of the RfC,
12 substantial weight was given to the available data demonstrating similarities between the effects
13 observed in humans following prolonged exposure to cerium oxide and the effects observed in
14 rats in the subchronic principal study, along with the data on deposition and absorption of cerium
15 oxide in the lung. Thus, these data support the assumption that the respiratory system may be
16 the most sensitive target of toxicity following inhalation exposure to cerium and that inhalation
17 exposure to cerium may primarily involve portal-of-entry effects.

18 The database for cerium oxide lacks both a two-generation reproductive toxicity bioassay
19 and a developmental toxicity bioassay. Systemic effects following the inhalation of cerium
20 oxide, with an MMAD of approximately 2.0 μm and a GSD from 1.8 to 1.9, are not likely to be
21 observed outside the lung. While it is recognized that the investigation of systemic effects
22 following cerium oxide exposure has not been the focus of existing studies, there is no reason to
23 expect that reproductive, developmental, or other systemic effects would occur, and a UF of 3 is
24 sufficient for the absence of data on these effects. The analysis of the scientific information
25 available for cerium as a whole supports the utilization of a database UF of 3.

26 The chronic RfC for cerium oxide was calculated as follows:

27
28
$$\begin{aligned} \text{RfC} &= \text{LOAEL}_{\text{HEC}} \div \text{UF} \\ &= 0.48 \text{ mg/m}^3 \div 3000 \\ &= 2 \times 10^{-4} \text{ mg/m}^3 \text{ (rounded to one significant figure)} \end{aligned}$$

31
32 Note that the RfC was quantified for cerium oxide particles with an MMAD of
33 approximately 2.0 μm and a GSD from 1.8 to 1.9, and may not appropriately characterize the
34 potential toxicity from exposures to cerium oxide particles with smaller MMADs and GSDs,
35 including nano-sized cerium particles. The use of the RfC for cerium compounds other than
36 cerium oxide is not recommended as the similarity between this form of cerium and other cerium
37 compounds is unknown.
38

1 **5.2.4. RfC Comparison Information**

2 Figure 5-1 is an exposure-response array, which presents NOAELs, LOAELs, and the
3 dose range tested corresponding to selected health effects from the BRL (1994) study, some of
4 which were considered candidates for chronic RfC derivation. Figure 5-2 presents the point of
5 departure, calculated as a human equivalent dose, applied uncertainty factors, and derived
6 chronic inhalation reference values for selected endpoints from Figure 5-1. This comparison is
7 intended to provide information for additional endpoints associated with cerium oxide inhalation.

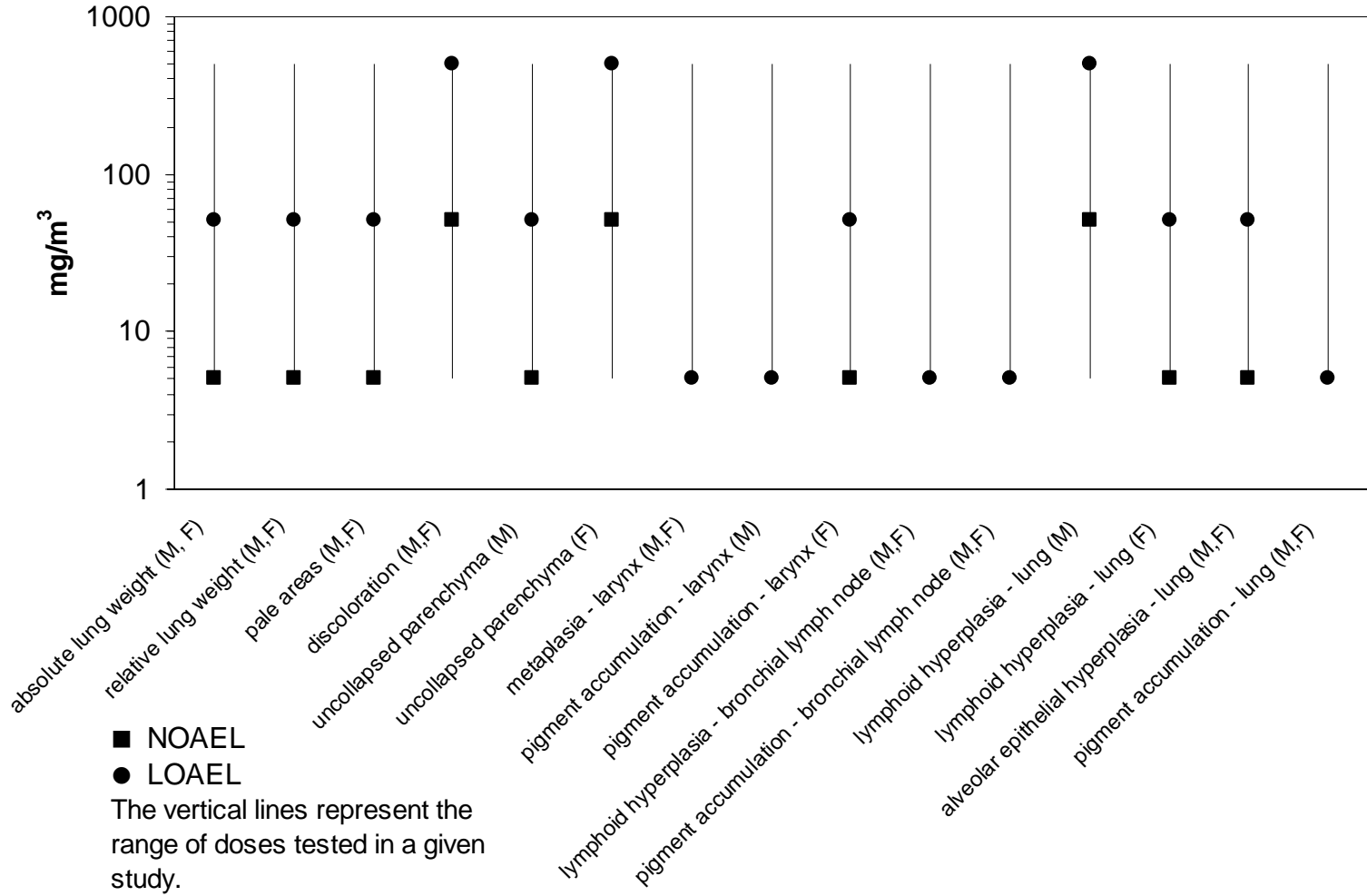


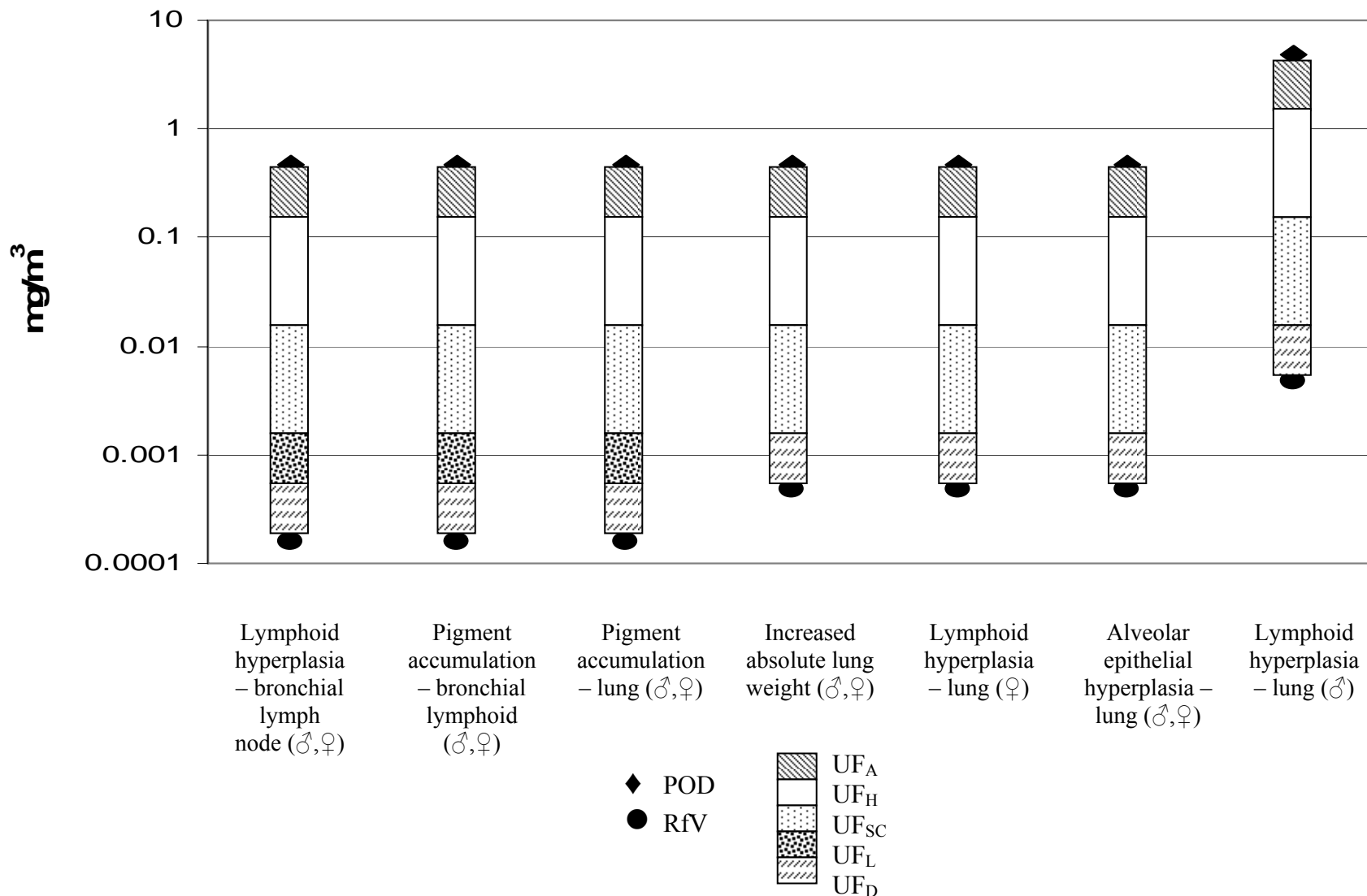
Figure 5-1. Exposure-response array of selected toxicity effects from the BRL (1994) study

Source: BRL, 1994.

2
3
4
5

1
2

Figure 5-2. Points of Departure for selected endpoints from Figure 5-1 with corresponding applied uncertainty factors and derived chronic inhalation RfVs.



1 **5.2.5. Previous RfC Assessment**

2 A previous IRIS assessment was not available for cerium oxide and cerium compounds.

3
4 **5.3. UNCERTAINTIES IN THE INHALATION REFERENCE CONCENTRATION**
5 **(RfC)**

6 Risk assessments need to portray associated uncertainty. The following discussion
7 identifies uncertainties associated with the RfC for cerium oxide. As presented earlier in this
8 chapter (Sections 5.2.2 and 5.2.3), UFs were applied to the point of departure for the RfC.
9 Factors accounting for uncertainties associated with a number of steps in the analyses were
10 adopted to account for extrapolating from an animal bioassay to human exposure, a diverse
11 population of varying susceptibilities, extrapolating from a subchronic to chronic exposure
12 duration, extrapolating from a LOAEL to a NOAEL, and database deficiencies.

13 A limited range of animal toxicology data is available for the hazard assessment of
14 cerium oxide, as described throughout the previous sections (see Sections 4 and 5). For the oral
15 route, human studies showing an association between exposure to cerium in food and the
16 development of endomyocardial fibrosis are available. Long-term studies in animals are limited
17 to a 13-month drinking water study in rats, which investigated the effects of a single dose group;
18 a 6-month drinking water study in rabbits in which the administered dose consisted of a mixture
19 of rare earth chlorides; a 12-week dietary study in mice; and a 105-day gavage study in rats. An
20 RfD for cerium was not derived since the available studies were not suitable for quantitation of
21 effects for various reasons, including unknown exposure concentrations in the available human
22 studies, lack of a dose response, uncertain biological significance (e.g., increased oxygen affinity
23 for hemoglobin, changes in measures of oxidative stress), and inadequate study design (e.g.,
24 effects noted only under conditions of a restricted diet).

25 The inhalation database includes a single, 13-week subchronic inhalation bioassay in rats,
26 with numerous case reports of workers who developed pneumoconiosis associated with
27 accumulation of cerium particles in the lungs after prolonged occupational exposure to cerium
28 fumes or dust. Critical data gaps have been identified and uncertainties associated with data
29 deficiencies are more fully discussed below.

30 Consideration of the available dose-response data to determine an estimate of inhalation
31 exposure that is likely to be without an appreciable risk of adverse health effects over a lifetime
32 led to the selection of the 13-week subchronic inhalation bioassay in Sprague-Dawley rats (BRL,
33 1994) and increased incidence of lymphoid hyperplasia in the bronchial lymph nodes of male
34 and female rats as the principal study and critical effect for deriving the RfC for cerium oxide.
35 The presence of lymphoid hyperplasia in the bronchial lymph nodes following cerium oxide
36 exposure may be part of a clearance process in which macrophages that have phagocytosed
37 cerium particles are removed to the lymphatic drainage. Foreign substances that are introduced

1 to the lung are cleared to the stomach and GI tract, the lymphatics and lymph nodes, and the
2 pulmonary vasculature (Witschi and Last, 2001). However, the database of cerium studies does
3 not include an exposure and recovery study that may demonstrate the persistence or, conversely,
4 the adaptive nature of the lymphoid hyperplasia in the bronchial lymph nodes. In the absence of
5 evidence indicating otherwise, these effects are assumed to represent cerium-induced toxicity.

6 The derived RfC was quantified using a LOAEL for the point of departure. A point of
7 departure based on a LOAEL or NOAEL is, in part, a reflection of the particular exposure
8 concentration or dose at which a study was conducted. It lacks characterization of the dose-
9 response curve and for this reason is less informative than a point of departure defined as an
10 effect level concentration (i.e., benchmark concentration [BMC]) obtained from benchmark
11 dose-response modeling. In this assessment, the exposure-related increase in pigment
12 accumulation and lymphoid hyperplasia in lymph nodes draining lungs, the bronchial and
13 mediastinal lymph nodes, in combination with the hypothesized mode of action, the overloading
14 of the PAMs by cerium oxide particles, further supports the role of pulmonary macrophages in
15 the nonspecific response. Thus, the lymphoid hyperplasia in the bronchial lymph nodes, which
16 was observed in 80% of the exposed rats, may be an early manifestation of the nonspecific
17 response to cerium oxide.

18 Extrapolating from animals to humans embodies further issues and uncertainties. The
19 effect and its magnitude associated with the concentration at the point of departure in rodents are
20 extrapolated to human response. Pharmacokinetic models are useful to examine species
21 differences in pharmacokinetic processing; however, dosimetric adjustment using
22 pharmacokinetic modeling was not possible for the toxicity observed following inhalation
23 exposure to cerium oxide. For the RfC, HECs were calculated from the point of departure by
24 multiplying the $LOAEL_{ADJ}$ by a DAF. The calculated DAF in this assessment is an RDDDR. The
25 RDDDR was calculated using the RDDDR v.2.3 program, as described in *Methods for Derivation of*
26 *Inhalation Reference Concentrations and Application of Inhalation Dosimetry* (U.S. EPA,
27 1994b).

28 Heterogeneity among humans is another uncertainty associated with extrapolating doses
29 from animals to humans. Uncertainty related to human variation needs consideration, also, in
30 extrapolating dose from a subset or smaller sized population, say of one sex or a narrow range of
31 life stages typical of occupational epidemiologic studies, to a larger, more diverse population.
32 Human variation may be larger or smaller; however, cerium-specific data to examine the
33 potential magnitude of over- or underestimation is unavailable.

34 Critical data gaps have been identified with uncertainties associated with database
35 deficiencies with regards to chronic toxicity, especially evidence demonstrating persistence of
36 lymphoid hyperplasia, and reproductive and developmental toxicity associated with inhalation
37 exposure to cerium. The available oral cerium exposure information in both humans and rats

1 identifies cardiac tissue and hemoglobin oxygen affinity as possible adverse health effects, but
2 the animal studies are of insufficient duration and experimental design and the human studies do
3 not provide an adequate exposure characterization. The lack of a sufficient study to derive an
4 RfD given the possible adverse effects demonstrated represents a critical data gap in the oral
5 database. A chronic, cerium oxide inhalation exposure study in animals is unavailable; however,
6 the workers in the human case reports were exposed to cerium oxide for periods of 10 to
7 46 years, and, while the case reports do not contain the necessary exposure analysis for dose
8 response assessment, they do provide evidence of adverse respiratory effects following long-
9 term exposure to cerium oxide. There are limited data available addressing possible
10 reproductive or neurodevelopmental toxicity following exposure to cerium. The accumulation of
11 insoluble cerium particles in the respiratory tract of humans and animals following chronic and
12 subchronic inhalation exposures, respectively, suggests that impaired clearance may influence
13 pulmonary toxicity in both rats and humans and limit systemic availability; therefore, possible
14 reproductive or developmental toxicity may be expected to occur at doses higher than those at
15 which portal-of-entry effects occurred.

16

17 **5.4. CANCER ASSESSMENT**

18 Studies addressing the carcinogenic effects of cerium or cerium compounds upon which
19 to base a cancer assessment are unavailable. Lundgren et al. (1996) observed lung neoplasms
20 (7/1,049) in cerium oxide exposed control F344/N rats in an investigation of the carcinogenicity
21 of the beta-particle emitter, ¹⁴⁴Ce. The incidence of lung neoplasms in the stable cerium oxide
22 exposed rats was not compared to a treatment-free group or to historical background levels in
23 F344/N rats.

24 As discussed in Section 4.7, data were unavailable regarding the carcinogenicity of stable
25 cerium in humans or experimental animals. In accordance with U.S. EPA (2005a) guidelines for
26 carcinogen risk assessment, there is “inadequate information to assess the carcinogenic
27 potential” of cerium oxide and cerium compounds. Lack of carcinogenicity data precludes
28 derivation of an oral slope factor or inhalation unit risk. Genotoxicity evidence was insufficient
29 to assess the genotoxic potential. This overall lack of information represents a data gap and does
30 not allow for a quantitative assessment of the carcinogenicity of cerium oxide and cerium
31 compounds. A previous cancer assessment was not available for cerium oxide and cerium
32 compounds.

1 **6. MAJOR CONCLUSIONS IN THE CHARACTERIZATION OF HAZARD**
2 **AND DOSE RESPONSE**

3
4
5 **6.1. HUMAN HAZARD POTENTIAL**

6 Cerium (CAS No. 7440-45-1) exposure is mostly in the form of mischmetal for
7 metallurgical purposes (Kilbourn, 2003). Cerium is the major component of mischmetal (50–
8 75% by weight for the most common grades), a commercial mixture of metallic light lanthanides
9 prepared by the electrolysis of mixed lanthanide chlorides and fluorides obtained from bastanite
10 or monazite (Kilbourn, 2003; Reinhardt and Winkler, 2002). Mischmetal reacts with the
11 impurities found in metals to form solid compounds, thereby reducing the effect of these
12 impurities on the properties of the metal (Reinhardt and Winkler, 2002). Cerium oxide (CeO₂;
13 ceric oxide) is the most important of the commercial cerium compounds. It is used either in the
14 pure form or in a concentrate as a polishing agent for glass mirrors, plate glass, television tubes,
15 ophthalmic lenses, and precision optics (Kilbourn, 2003; Reinhardt and Winkler, 2002). Cerium
16 oxide is used as a glass constituent to prevent solarization and discoloration (especially in the
17 faceplates of television screens) (Reinhardt and Winkler, 2002). Cerium oxide is also used as a
18 diesel fuel-borne catalyst to reduce particulate matter emissions (HEI, 2001). Cerium is not
19 expected to exist in elemental form in the environment since it is a very reactive metal (Lewis,
20 2001). Cerium compounds are not expected to volatilize and will exist in the particulate form if
21 released into the air.

22 Toxicokinetic studies in rodents have examined the absorption, distribution, metabolism,
23 and elimination of cerium. In adult rats, cerium compounds are very poorly absorbed following
24 oral exposure, while suckling animals exhibit higher absorption and retention of cerium in the GI
25 tissues (Kostial et al., 1989a, b; Inaba and Lengemann, 1972). Following inhalation exposure,
26 cerium, a poorly-soluble particle, behaves like other airborne particles, depositing within the
27 respiratory tract based on aerodynamic character (for review, see Schulz et al., 2000). Cerium
28 has been detected in lung tissues and in alveolar macrophages of subjects believed to have been
29 exposed occupationally (Vocaturro et al., 1983; Sabbioni et al., 1982), with cerium concentrations
30 in the lung tissues 2,800–207,000 times higher than those found in the urine, blood, or nails
31 (Pietra et al., 1985). The early clearance of the radioactive cerium administered dose suggests
32 that the majority of cerium aerosol is deposited in the airways, where it is subject to removal via
33 the mucociliary escalator, swallowing, and elimination in the feces.

34 Once absorbed into the body, cerium tends to accumulate primarily in the bone, liver,
35 heart, and lung. Cerium has been observed to be localized in the cell, particularly in the
36 lysosomes, where it is concentrated and precipitated in an insoluble form in association with
37 phosphorus. As an element, cerium is neither created nor destroyed within the body. The
38 particular cerium compound (e.g., cerium chloride, cerium oxide) may be altered as a result of

1 various chemical reactions within the body, particularly dissolution, but data have not
2 demonstrated a change in the oxidation state of the cerium molecule (Berry et al., 1989, 1988).

3 Following inhalation exposure, the initial rapid elimination of cerium from the body is
4 due primarily to transport up the respiratory tract by the mucociliary escalator and eventual
5 swallowing of the material, as with other poorly soluble particles (Boecker and Cuddihy, 1974).
6 After the initial clearance of cerium particles from the upper respiratory tract, pulmonary
7 clearance is slower, with reported slow-phase clearance half-times ranging from 100 to 190 days
8 in rodents (Lundgren et al., 1974; Thomas et al., 1972; Morgan et al., 1970; Sturbaum et al.,
9 1970). Elimination of orally administered cerium has been shown to be age dependent in
10 animals, with suckling animals absorbing cerium into the GI tissues (Inaba and Lengemann,
11 1972).

12 An epidemiological study reports a higher incidence of endomyocardial fibrosis among a
13 population consuming tubers grown in a high cerium soil in India (Eapen, 1998; Kutty et al.,
14 1996; Valiathan et al., 1989), and a case control study found an association between increased
15 toenail cerium concentrations and the risk of first myocardial infarction (Gómez-Aracena et al.,
16 2006). The few long-term animal studies available included dietary, gavage, and drinking water
17 administrations, although clear indicators of adverse effects were not evident.

18 Numerous case reports have been published describing cases of workers who developed
19 adverse lung effects, such as interstitial lung disease or pneumoconiosis, associated with
20 accumulation of cerium in the lungs after prolonged occupational exposure to cerium fumes or
21 dust (Yoon et al., 2005; Porru et al., 2001; McDonald et al., 1995; Pairon et al., 1995, 1994;
22 Sulotto et al., 1986; Vogt et al., 1986; Pietra et al., 1985; Vocaturo et al., 1983; Sabbioni et al.,
23 1982; Husain et al., 1980; Kappenberger and Bühlmann, 1975; Heuck and Hoschek, 1968). The
24 human cases of cerium exposure demonstrate the accumulation of cerium particles in the lungs
25 and lymphoreticular system, with pulmonary function varying from normal to severe restriction
26 and interstitial fibrosis in one case and granulomas in another. Interstitial fibrosis accompanied
27 by vascular thickening, reactive alveolar macrophages, abundant macrophages in the airspace,
28 and moderate chronic interstitial inflammation, along with small interstitial clumps of
29 macrophages bearing scant deposits of grayish-black pigment, was observed in a 68-year-old
30 man who was employed as an optical lens grinder for 35 years (McDonald et al., 1995).
31 Additionally, particles characterized as cerium were identified in alveolar macrophages,
32 macrophages in the tracheobronchial lymph nodes, and lung and lymph node tissue (Porru et al.,
33 2001; Pairon et al., 1995; Waring and Watling, 1990; Sulotto et al., 1986; Vogt et al., 1986;
34 Pietra et al., 1985). Cerium exposure was associated with interstitial, peribronchial, and
35 perivascular fibrosis, a restriction of respiratory function, and/or pulmonary hypertension (Porru
36 et al., 2001; Pairon et al., 1995, 1994; Vogt et al., 1986; Vocaturo et al., 1983; Sabbioni et al.,
37 1982).

1 Data from a subchronic toxicity test in Sprague-Dawley rats (BRL, 1994) identified an
2 increased incidence of lymphoid hyperplasia in the bronchial lymph nodes as the critical effect
3 for noncancer effects. Histologic examination revealed dose-related alveolar epithelial and
4 lymphoid hyperplasia and pigment accumulation in the lungs, lymph nodes, and larynx of male
5 and female rats at ≥ 5 mg/m³.

6 The mode of action for cerium oxide inhalation toxicity may be mediated by cytokine
7 and fibrogenic effects resulting from pulmonary macrophage activation followed by macrophage
8 immobilization. The accumulation of insoluble cerium oxide particles in the respiratory tract of
9 humans and rodents following chronic and subchronic exposure, respectively, suggests that
10 impaired clearance may influence pulmonary toxicity for rats and humans. A population of
11 immobilized, activated macrophages may serve to induce significant cell damage by effectively
12 increasing the concentration of inflammatory cytokines and fibrogenic growth factors within the
13 pulmonary epithelium.

14 15 **6.2. DOSE RESPONSE**

16 **6.2.1. Noncancer/Oral**

17 The database for oral exposure to cerium is limited to geographical distribution studies
18 evaluating the possible association between exposure to cerium in food and the development of
19 endomyocardial fibrosis (Eapen et al., 1998; Kutty et al., 1996; Valiathan et al., 1989), and long-
20 term animal studies, including a 13-month drinking water study in rats (Kumar et al., 1996), a
21 6-month drinking water study in rabbits (Kartha et al., 1998), a 12-week dietary study in mice
22 (Kawagoe et al., 2005), and a 105-day gavage study in rats (Cheng et al., 2000).

23 The geographical distribution studies suggest that there is an association between
24 exposure to cerium in food and the development of endomyocardial fibrosis but are unsuitable
25 for RfD derivation. The above animal studies are unsuitable for derivation of an RfD for various
26 reasons. The 13-month study by Kumar et al. (1996) was conducted on a small number of rats at
27 only one dose level (control and dosed rats) and evaluated few endpoints, and the health effects
28 were highly variable and not statistically significant. Kartha et al. (1998) conducted a 6-month
29 drinking water study in rabbits that utilized a rare earth chloride mixture in the drinking water,
30 which suggests that cerium may enhance the effect of magnesium deficiency in heart tissue. The
31 Cheng et al. (2000) study was limited in scope, since it investigated the effects of cerium
32 chloride exposure on the structure and affinity of hemoglobin in rats. The evaluations conducted
33 in Kawagoe et al. (2005), showing statistically significant changes as a result of cerium exposure
34 (e.g., decreased lipoperoxide levels), increased GSH levels and MT activity, and decreased SOD
35 activity, are of unknown biological significance and may represent oxidative stress in response to
36 cerium exposure.

1 The available oral cerium exposure information in both humans and rats identifies
2 cardiac abnormalities and changes in hemoglobin oxygen affinity as possible effects, but the
3 animal studies are of insufficient duration and experimental design and the human studies do not
4 provide an adequate exposure characterization. The lack of a sufficient study to derive an RfD
5 given the effects demonstrated represents a critical data gap in the oral database.
6

7 **6.2.2. Noncancer/Inhalation**

8 There are numerous case reports of workers who developed pneumoconiosis, associated
9 with accumulation of cerium particles in the lungs, after prolonged occupational exposure to
10 cerium fumes or dust (Yoon et al., 2005; Porru et al., 2001; McDonald et al., 1995; Pairon et al.,
11 1995, 1994; Sulotto et al., 1986; Vogt et al., 1986; Pietra et al., 1985; Vocaturo et al., 1983;
12 Sabbioni et al., 1982; Husain et al., 1980; Kappenberger and Bühlmann, 1975; Heuck and
13 Hoschek, 1968). Information regarding long-term inhalation exposure in animals is derived
14 from a single subchronic study in rats (BRL, 1994). Sprague-Dawley rats were exposed nose
15 only to cerium oxide aerosol 6 hours/day, 5 days/week for 13 weeks. Endpoints evaluated
16 included a functional observational battery, hematology and clinical chemistry, urinalysis, and
17 gross and microscopic morphology of tissues. The critical effect of cerium oxide exposure is
18 increased incidence of alveolar lymphoid hyperplasia in the bronchial lymph nodes of male and
19 female rats with a LOAEL of 5 mg/m³.

20 Consideration of the available dose-response data to determine an estimate of inhalation
21 exposure that is likely to be without an appreciable risk of adverse health effects over a lifetime
22 led to the selection of the 13-week subchronic inhalation bioassay in Sprague-Dawley rats (BRL,
23 1994) and increased incidence of lymphoid hyperplasia in the bronchial lymph nodes of male
24 and female rats as the principal study and critical effect for deriving the RfC for cerium oxide.
25 The presence of lymphoid hyperplasia in the bronchial lymph nodes following cerium oxide
26 exposure may be part of a clearance process in which macrophages that have phagocytosed
27 cerium particles are removed to the lymphatic drainage. Foreign substances that are introduced
28 to the lung are cleared to the stomach and GI tract, lymphatics and lymph nodes, and pulmonary
29 vasculature (Witschi and Last, 2001). However, the database of cerium studies does not include
30 an exposure and recovery study that may demonstrate the persistence of the lymphoid
31 hyperplasia in the bronchial lymph nodes. Thus, in the absence of evidence demonstrating
32 otherwise, these effects are believed to represent cerium-induced toxicity.

33 The RfC was derived using a LOAEL for the point of departure. A point of departure
34 based on a LOAEL or NOAEL is, in part, a reflection of the particular exposure concentration or
35 dose at which a study was conducted. It lacks characterization of the dose-response curve and
36 for this reason is less informative than a point of departure defined as an effect level
37 concentration (i.e., BMC) obtained from benchmark dose-response modeling. In this

1 assessment, the exposure-related increase in pigment accumulation and lymphoid hyperplasia in
2 lymph nodes draining lungs (the bronchial and mediastinal lymph nodes), in combination with
3 the hypothesized mode of action (the overloading of the PAMs by cerium oxide particles),
4 further supports the role of pulmonary macrophages in the nonspecific response. Thus, the
5 lymphoid hyperplasia in the bronchial lymph nodes is an early manifestation of the nonspecific
6 response to cerium oxide.

7 Extrapolating from animals to humans embodies further issues and uncertainties. The
8 effect and its magnitude associated with the concentration at the point of departure in rodents are
9 extrapolated to human response. Pharmacokinetic models are useful to examine species
10 differences in pharmacokinetic processing; however, dosimetric adjustment using
11 pharmacokinetic modeling was not possible for the toxicity observed following inhalation
12 exposure to cerium oxide. For the RfC, an HEC was calculated from the point of departure by
13 multiplying the $LOAEL_{ADJ}$ by a DAF. The calculated DAF in this assessment is an RDDR. The
14 RDDR was calculated using the RDDR v.2.3 program, as described in *Methods for Derivation of*
15 *Inhalation Reference Concentrations and Application of Inhalation Dosimetry* (U.S. EPA,
16 1994b).

17 Heterogeneity among humans is another uncertainty associated with extrapolating doses
18 from animals to humans. Uncertainty related to human variation needs consideration, also, in
19 extrapolating dose from a subset or smaller-sized population, say of one sex or a narrow range of
20 life stages typical of occupational epidemiologic studies, to a larger, more diverse population.
21 Human variation may be larger or smaller; however, cerium-specific data to examine the
22 potential magnitude of over- or underestimation are unavailable.

23 The RfC of 2×10^{-4} mg/m³ was calculated from a $LOAEL_{HEC}$ of 0.48 mg/m³ for
24 increased incidence of lymphoid hyperplasia in the bronchial lymph nodes in male and female
25 rats following subchronic cerium oxide inhalation exposure (BRL, 1994). A total UF of 3,000
26 was used: 3 for interspecies extrapolation, 10 for intraspecies variability, 10 for subchronic to
27 chronic extrapolation, 3 for extrapolating from a LOAEL to a NOAEL, and 3 for database
28 deficiencies.

29 A factor of 3 was selected to account for uncertainties in extrapolating from rats to
30 humans, which is adopted by convention where an adjustment from an animal-specific
31 $NOAEL_{ADJ}$ to a $NOAEL_{HEC}$ has been incorporated. Insufficient information is available to
32 predict potential variability in susceptibility among the population, thus a human variability UF
33 of 10 was applied. A 10-fold UF was used to account for uncertainty in extrapolating from a
34 subchronic to chronic exposure duration. A 3-fold UF was applied to account for the
35 extrapolation from a LOAEL to NOAEL. The critical effect for this analysis, lymphoid
36 hyperplasia in the bronchial lymph nodes, may represent the point at which normal clearance of
37 particles from the lung by alveolar macrophages becomes overwhelmed and particles are no

1 longer cleared effectively, leading to an increasing accumulation of particles in the lung and
2 airways, an inflammatory response, and subsequent cell proliferation. However, due to the
3 absence of evidence demonstrating the persistence of lymphoid hyperplasia, these effects are
4 believed to represent cerium-induced toxicity. Data gaps have been identified with uncertainties
5 associated with database deficiencies with regards to reproductive and developmental toxicity
6 associated with cerium inhalation exposure. A database UF of 3 was applied with special
7 consideration of the information pertaining to the deposition and absorption of cerium oxide, the
8 effects observed in humans following prolonged exposure, the mode-of-action data, and the
9 effects observed in animals in BRL (1994), in addition to the lack of reproductive and
10 developmental studies.

11 The overall confidence in this RfC assessment is low. Confidence in the principal study
12 (BRL, 1994) is medium. EPA conducted an external peer review to evaluate the accuracy of the
13 experimental procedures, results, and interpretation and discussion of the results presented in this
14 study. The peer reviewers considered the BRL (1994) study to be adequate and the study
15 conclusions to be supported by the data. The peer reviewers were not specifically asked to
16 comment on their confidence in the study. In addition, the results observed in the BRL (1994)
17 study were consistent with the observed effects in the human case reports (Yoon et al., 2005;
18 Porru et al., 2001; McDonald et al., 1995; Pairen et al., 1995, 1994; Sulotto et al., 1986; Vogt et
19 al., 1986; Pietra et al., 1985; Vocaturo et al., 1983; Sabbioni et al., 1982; Husain et al., 1980;
20 Kappenberger and Bühlmann, 1975; Heuck and Hoschek, 1968). Confidence in the database is
21 low. The database lacks chronic exposure information on cerium via any route of exposure and
22 multigenerational developmental and reproductive toxicity studies. However, there is evidence
23 of cerium pneumoconiosis in humans exposed to cerium compounds, and the anticipated critical
24 effects observed are point-of-entry effects that would be expected in humans. Reflecting
25 medium confidence in the principal study and low confidence in the database, confidence in the
26 RfC is low.

27

28 **6.2.3. Cancer/Oral and Inhalation**

29 Under the *Guidelines for Carcinogen Risk Assessment* (U.S. EPA, 2005a), the database
30 for cerium oxide and cerium compounds is inadequate to assess human carcinogenic potential
31 and to calculate quantitative cancer risk estimates.

7. REFERENCES

- 1
2
- 3 Arvela, P; Karki, NT. (1971) Effect of cerium on drug metabolism activity in rat liver. *Experientia* 27(10):1189–
4 1190.
- 5 Arvela, P; Kraul, H; Stenback, F; et al. (1991) The cerium-induced liver injury and oxidative drug metabolism in
6 *Dbal/2* and *C57bl/6* mice (Finland). *Toxicology* 69(1):1–9.
- 7 Berry, JP. (1996) The role of lysosomes in the selective concentration of mineral elements. A microanalytical study.
8 *Cell Mol Biol* 42(3):395–411.
- 9 Berry, JP; Meignan, M; Escaig, F; et al. (1988) Inhaled soluble aerosols insolubilised by lysosomes of alveolar cells.
10 Application to some toxic compounds; electron microprobe and ion microprobe studies. *Toxicology* 52(1–2):127–
11 139.
- 12 Berry, JP; Masse, R; Escaig, F; et al. (1989) Intracellular localization of cerium. A microanalytical study using an
13 electron microprobe and ionic microanalysis. *Hum Toxicol* 8(6):511–520.
- 14 Berry, JP; Zhang, L; Galle, P; et al. (1997) Role of alveolar macrophage lysosomes in metal detoxification.
15 *Microscop Res Technol* 36:313–323.
- 16 Boecker, BB; Cuddihy, RG. (1974) Toxicity of ^{144}Ce Inhaled as $^{144}\text{CeCl}_3$ by the beagle: metabolism and dosimetry.
17 *Radiat Res* 60(1):133–154.
- 18 BRL (Bio-Research Laboratories). (1994) Final report for a 90-day inhalation neurotoxicity and toxicity study by
19 exposure to a dry powder aerosol of ceric oxide in the albino rat with cover letter dated 013095. Produced by Bio-
20 Research Laboratories, Montreal, Canada for Rhone-Poulenc Inc. Submitted under TSCA Section 8E; EPA Doc. No.
21 89-950000107; NTIS No. OTS0556254. [An external peer review was conducted by EPA in August 2006 to
22 evaluate the accuracy of experimental procedures, results, and interpretation and discussion of the findings
23 presented. A report of this peer review is available through the EPA's IRIS Hotline, at (202) 566-1676 (phone),
24 (202) 566-1749 (fax), or hotline.iris@epa.gov (e-mail address) and on the IRIS website (www.epa.gov/iris).]
- 25 Bruce, DW; Hietbrink, BE; DuBois, KP. (1963) The acute mammalian toxicity of rare earth nitrates and oxides.
26 *Toxicol and App Pharmacol* 5:750-759.
- 27 Brunner, TJ; Wick, P; Manser, P; et al. (2006) In vitro cytotoxicity of oxide nanoparticles: comparison to asbestos,
28 silica, and the effect of particle solubility. *Environ Sci Technol* 40(14):4374–4381
- 29 Chen, J; Tan, M; Nemmar, A; et al. (2006) Quantification of extrapulmonary translocation of intratracheal-instilled
30 particles in vivo in rats: effect of lipopolysaccharide. *Toxicology* 222:195–201.
- 31 Cheng, Y; Li, Y; Li, R; et al. (2000) Orally administered cerium chloride induces the conformational changes of rat
32 hemoglobin, the hydrolysis of 2,3-DPG and the oxidation of heme-Fe(II), leading to changes of oxygen affinity.
33 *Chem Biol Interact* 125(3):191–208.
- 34 Cotton, FA; Wilkinson G; Murillo CA. (1999) *Advanced inorganic chemistry*. 6th edition. New York, NY: John
35 Wiley and Sons, Inc.; p. 1125.
- 36 Cullen, RT; Tran, CL; Buchanan, D; et al. (2000) Inhalation of poorly soluble particles. I. differences in
37 inflammatory response and clearance during exposure. *Inhal Tox* 12:1089-1111.
- 38 D'Agostino, R; Olson, FC; Massaro, EJ. (1978a) Growth and behavioral abnormalities in offspring of mice treated
39 with cerium (citrate) or platinum (hexachloroplatinate) during pregnancy or lactation. *Proc Int Congr Toxicol*
40 1:608–609.

- 1 D'Agostino, R; Olson, FC; Massaro, EJ; et al. (1978b) Alterations of growth and behavioral parameters of offspring
2 of female mice exposed to cerium (citrate) or platinum (hexachloroplatinate) during pregnancy or lactation. In:
3 Hemphill, DD; ed. Trace substances in environmental health, XI: proceedings of the University of Missouri's 11th
4 annual conference on trace substances in environmental health. Columbia, MO: University of Missouri; pp. 257–
5 263.
- 6 D'Agostino, RB; Lown, BA; Morganti, JB; et al. (1982) Effects of in utero or suckling exposure to cerium (citrate)
7 on the postnatal development of the mouse. *J Toxicol Environ Health* 10:449–458.
- 8 Du, X; Zhang, T; Li, R; et al. (2001) Nature of cerium(III)- and lanthanum(III)-induced aggregation of human
9 erythrocyte membrane proteins. *J Inorg Biochem* 84(1–2):67–75.
- 10 Durbin, PW; Williams, MH; Gee, M; et al. (1956) Metabolism of the lanthanons in the rat. *Proc Soc Exper Biol*
11 *Med* 91:78–85.
- 12 Eapen, JT; Kartha, CC; Rathinam, K; et al. (1996) Levels of cerium in the tissues of rats fed a magnesium-restricted
13 and cerium-adulterated diet. *Bull Environ Contam Toxicol* 56(2):178–182.
- 14 Eapen, JT. (1998) Elevated levels of cerium in tubers from regions endemic for endomyocardial fibrosis (EMF).
15 *Bull Environ Contam Toxicol* 60:168–170.
- 16 Eisele, GR; Mraz, FR; Woody, MC. (1980) Gastrointestinal uptake of ¹⁴⁴Ce in the neonatal mouse, rat and pig.
17 *Health Phys* 39:185–192.
- 18 Evans, LJ. (1989) Chemistry of metal retention by soils. *Environ Sci Technol* 23:1046–1054.
- 19 Fedulov AV; Leme, A; Yang, Z; et al. (2008) Pulmonary exposure to particles during pregnancy causes increased
20 neonatal asthma susceptibility. *Amer Journ Resp Cell Molec Biol* 38:57-67.
- 21 Ferin, J. (1994) Pulmonary retention and clearance of particles. *Toxicol Lett* 72(1-3):121-125.
- 22 Galle, P; Berry, JP; Galle, C. (1992) Role of alveolar macrophages in precipitation of mineral elements inhaled as
23 soluble aerosols. *Environ Health Perspect* 97:145–147.
- 24 Gómez-Aracena, J; Riemersma, RA; Guitierrez-Bedmar, M; et al. (2006) Toenail cerium levels and risk of a first
25 acute myocardial infarction: The EURAMIC and heavy metals study. *Chemosphere* 64: 112–120.
- 26 Graca, JG; Davison, FC; Feavel, JB. (1964) Comparative toxicity of stable rare earth compounds. III. Acute toxicity
27 of intravenous injections of chlorides and chelates in dogs. *Arch Environ Health* 89:555–564.
- 28 Hahn, FF; Muggenberg, BA; Guilmette, RA; et al. (1999) Comparative stochastic effects of inhaled alpha- and beta-
29 particle-emitting radionuclides in beagle dogs. *Radiat Res* 152:S19-S22
- 30 Hahn, FF; Muggenburg, BA; Snipes, MB; et al. (2001) The toxicity of insoluble cerium-144 inhaled by beagle dogs:
31 non-neoplastic effects. *Radiat Res* 155(1):95–112.
- 32 Haley, PJ. (1991) Pulmonary toxicity of stable and radioactive lanthanides. *Health Phys* 61(6):809–820.
- 33 Hedrick, JB. (2004) Rare earths. *Minerals yearbook. Vol. I. Metals and minerals*. U.S. Geological Survey, U.S.
34 Department of the Interior, Reston, VA. Available online at
35 <http://minerals.usgs.gov/minerals/pubs/commodity/myb/>.
- 36 HEI (Health Effects Institute). (2001) Evaluation of human health risk from cerium added to diesel fuel. HEI
37 communication 9. North Andover, MA; Flagship Press. Available online at <http://pubs.healtheffects.org/>.
- 38 Heuck, F; Hoschek, R. (1968) Cer-pneumoconiosis. *Am J Roentgenol Radium Ther Nucl Med* 104(4):777–783.

- 1 Hunninghake, GW; Hemken, C; Brady, M; et al. (1984) Mediators released by human lung cells are potent growth
2 factors for human lung fibroblasts. *Trans Assoc Am Physicians* 97:41–48. (as cited in Haley, 1991).
- 3 Husain, MH; Dick, JA; Kaplan, YS. (1980) Rare earth pneumoconiosis. *J Soc Occup Med* 30:15–19.
- 4 Inaba, J; Lengemann, FW. (1972) Intestinal uptake and whole-body retention of ^{141}Ce by suckling rats. *Health Phys*
5 22:169–175.
- 6 Inaba, J; Nishimura, Y; Takeda, H; et al. (1992) Placental transfer of cerium in the rat with special reference to route
7 of administration. *Radiat Prot Dosimet* 41(2/4):119–122.
- 8 Ji, YJ; Cui, MZ. (1988) Toxicological studies on safety of rare earths used in agriculture. *Biomed Environ Sci* 1:
9 270–276.
- 10 Kanapilly, GM; Luna, RJ. (1975) Deposition and retention of inhaled condensation aerosols of $^{144}\text{CeO}_2$ in Syrian
11 hamsters. In: Boecker, BB; Rupperecht, FC; eds. Annual report of the Inhalation Toxicology Research Institute.
12 Lovelace Foundation for Medical Education and Research, Albuquerque, NM; Report No. LF-52; pp. 79–83.
- 13 Kappenberger, L; Bühlmann, AA. (1975) [Lung lesions caused by “rare earths”]. *Schweiz Med Wochenscher*
14 105:1799–1801. (German with English summary).
- 15 Kargacin, B; Landeka, M. (1990) Effect of glucocorticoids on metal retention in rats. *Bull Environ Contam Toxicol*
16 45(5):655–661.
- 17 Kartha, CC; Eapen, JT; Radhakumary, C; et al. (1998) Pattern of cardiac fibrosis in rabbits periodically fed a
18 magnesium-restricted diet and administered rare earth chloride through drinking water. *Biol Trace Elem Res*
19 63(1):19–30.
- 20 Kawagoe, M; Hirasawa, F; Wang, SC; et al. (2005) Orally administered rare earth element cerium induces
21 metallothionein synthesis and increases glutathione in the mouse liver. *Life Sci* 77:922–937.
- 22 Kilbourn, BT. (2003) Cerium and cerium compounds. In: Kirk-Othmer encyclopedia of chemical technology. New
23 York, NY: John Wiley and Sons, Inc. Available online with subscription at
24 http://www.mrw.interscience.wiley.com/kirk/kirk_search_fs.html.
- 25 Kostial, K; Kargacin, B; Landeka, M. (1989a) Gut retention of metals in rats. *Biol Trace Elem Res* 21:213–218.
- 26 Kostial, K; Kargacin, B; Blanusa, M; et al. (1989b) Location of mercury, cerium, and cadmium in the gut of suckling
27 and weaned rats. *Period Biol* 91(3):321–326.
- 28 Kumar, BP; Shivakumar, K. (1998) Alterations in collagen metabolism and increased fibroproliferation in the heart
29 in cerium-treated rats. *Biol Trace Elem Res* 63:73–79.
- 30 Kumar, BP; D'Souza, SL; Shivakumar, K; et al. (1995) Cerium stimulates protein biosynthesis in rat heart in vivo.
31 *Biol Trace Elem Res* 50(3):237–242.
- 32 Kumar, BP; Shivakumar, K; Kartha, CC; et al. (1996) Magnesium deficiency and cerium promote fibrogenesis in rat
33 heart. *Bull Environ Contam Toxicol* 57(4):517–524.
- 34 Kuruvilla, L; Kartha, CC. (2006) Cerium depresses endocardial endothelial cell-mediated proliferation of cardiac
35 fibroblasts. *Biol Trace Elem Res* 114:85-92.
- 36 Kutty, VR; Abraham, S; Kartha, CC. (1996) Geographical distribution of endomyocardial fibrosis in South Kerala.
37 *Int J Epi* 25(6):1202–1207.
- 38 Levack, VM; Hone, PA; Phipps, AW; et al. (2002) The placental transfer of cerium: experimental studies and
39 estimates of doses to the human fetus from ^{141}Ce and ^{144}Ce . *Int J Radiat Biol* 78:227–235.

- 1 Lewis, RJ; ed. (2001) Hawley's condensed chemical dictionary. New York, NY: John Wiley and Sons, Inc.; pp.
2 229–231.
- 3 Lide, DR; ed. (2005) CRC handbook of chemistry and physics. 86th edition. Boca Raton, FL: CRC Press, Taylor &
4 Francis; pp. 4-9, 4-47, 4-48.
- 5 Limbach, LK; Li, Y; Grass, RN; et al. (2005) Oxide nanoparticle uptake in human lung fibroblast: effects of particle
6 size, agglomeration, and diffusion at low concentrations. *Environ Sci Technol* 39(23):9370–9376.
- 7 Lin, W; Huang, Y; Zhou, X; et al. (2006) Toxicity of cerium oxide nanoparticles in human lung cancer cells. *Int J*
8 *Toxicol* 25:451–457.
- 9 Lizon, C; Fritsch, P. (1999) Chemical toxicity of some actinides and lanthanides towards alveolar macrophages: an
10 in vitro study. *Int J Radiat Biol* 75(11):1459–1471.
- 11 Lombardi, B; Recknagel, RO. (1962) Interference with secretion of triglycerides by the liver as a common factor in
12 toxic liver injury. *Am J Pathol* 40(5):571–586.
- 13 Lundgren, DL; McClellan, RO; Thomas, RL; et al. (1974) Toxicity of inhaled ¹⁴⁴CeO₂ in mice. *Radiat Res* 58:448–
14 461.
- 15 Lundgren, DL; Hahn, FF; Diel, JH; et al. (1992) Repeated inhalation exposure of rats to aerosols of ¹⁴⁴CeO₂. I. Lung,
16 liver, and skeletal dosimetry. *Radiat Res* 132(3):312–24.
- 17 Lundgren, DL; Hahn, FF; Griffith, WC; et al., (1996) Pulmonary carcinogenicity of relatively low doses of beta-
18 particle radiation from inhaled ¹⁴⁴CeO₂ in rats. *Radiat Res* 146(5):525–535.
- 19 Lustgarten, CS; Boecker, BB; Cuddihy, RG; et al. (1976) Biliary excretion of ¹⁴⁴Ce after inhalation of ¹⁴⁴Ce citrate in
20 rats and Syrian hamsters. III. Annual report of the Inhalation Toxicology Research Institute. Lovelace Foundation
21 for Medical Education and Research, Albuquerque, NM; pp. 84–87.
- 22 Magnusson, G. (1962) Acute toxic effects of yttrium, cerium, terbium, holmium and ytterbium on the liver. *Acta*
23 *Pharmacol Toxicol* 20(Suppl.):53–74.
- 24 Manju, L; Remani, K; Nair, RR. (2003) Negative inotropic response to cerium in ventricular papillary muscle is
25 mediated by reactive oxygen species. *Biol Trace Elem Res* 96(1–3):203–208.
- 26 Manoubi, L; Hocine, N; Jaafoura, H; et al. (1998) Subcellular localization of cerium in intestinal mucose, liver,
27 kidney, suprarenal and testicle glands, after cerium administration in the rat. *J Trace Microprobe Techniques*
28 16(2):209–219.
- 29 Marciniak, M; Baltrukiewicz, Z. (1977) Serum ornithine carbamoyltransferase (OCT) in rats poisoned with
30 lanthanum, cerium and praseodymium. *Acta Physiol Pol* 28(6):589–594.
- 31 Marciniak, M; Baltrukiewicz, Z. (1981) Effect of various doses of cerium on the serum level of ornithine-
32 carbamyltransferase in rats. *Acta Physiol Pol* 32(2):205–211.
- 33 McDonald, JW; Ghio, AJ; Sheehan, CE; et al. (1995) Rare earth (cerium oxide) pneumoconiosis—analytical
34 scanning electron microscopy and literature review. *Mod Pathol* 8(8):859–865.
- 35 Mogilevskaya, OY; Raikhlin, NT. (1967) Rare earth metals. In: Izrael'son, ZI; ed. *Toxicology of the rare metals*.
36 Jerusalem, Israel: Israel Program for Scientific Translations; pp. 132–141.
- 37 Monafu, WW; Tandon, SN; Ayvazian, VH; et al. (1976) Cerium nitrate: a new topical antiseptic for extensive burns.
38 *Surgery* 80(4):465–473.
- 39 Morgan, BN; Thomas, RG; McClellan, RO. (1970) Influence of chemical state of cerium-144 on its metabolism
40 following inhalation by mice. *Am Ind Hyg Assoc J* 31(4):479–484.

- 1 Morganti, JB; Lown, BA; Stineman, CH; et al. (1978) Cerium tissue/organ distribution and alterations in open field
2 and exploratory behavior following repeated exposure of the mouse to citrate complexed cerium. *Gen Pharmacol*
3 9(4):257–261.
- 4 Morganti, JB; Lown, BA; Chapin, E; et al. (1980) Effects of acute exposure to cerium (citrate) on selected measures
5 of activity, learning and social behavior of the mouse. *Gen Pharmacol* 11(4):369–373.
- 6 Morrow, PE. (1988) Possible mechanisms to explain dust overloading of the lungs. *Fundam Appl Toxicol* 10:369–
7 384.
- 8 Morrow, PE. (1992) Dust overloading of the lungs: update and appraisal. *Toxicol Appl Pharmacol* 113(1):1–12.
- 9 Mraz, FR; Eisele, GR. (1977) Gastrointestinal absorption and distribution of ¹⁴⁴Ce in the suckling pig. *Health Phys*
10 33:494–495.
- 11 Naharin, A; Lubin, E; Feige, Y. (1969) Transfer of ¹⁴⁴Ce to mouse embryos and offspring via placenta and lactation.
12 *Health Phys* 17:717–722.
- 13 Naharin, A; Feige, Y; Lubin, E; et al. (1974) Internal deposition of ingested cerium in suckling mice. *Health Phys*
14 27(2):207–211.
- 15 Nair, R; Preeta, R; Smitha, G; et al. (2003) Variation in mitogenic response of cardiac and pulmonary fibroblasts to
16 cerium. *Biol Trace Elem Res* 94(3):237–246.
- 17 Nishioka, H. (1975) Mutagenic activities of metal compounds in bacteria. *Mutat Res* 31:185–190.
- 18 Niu et al., 2007, p. 43—could this be Niu, J; Azfer, A; Rogers, LM. (2007) Cardioprotective effects of cerium oxide
19 nanoparticles in a transgenic murine model of cardiomyopathy. *Cardiovasc Res* 73(3):549-559.
- 20 NLM (National Library of Medicine) (2006) Cerium (last modified in September, 2004). ChemIDplus. National
21 Institutes of Health, U.S. Department of Health and Human Services, Bethesda, MD. Available online at
22 <http://toxnet.nlm.nih.gov>.
- 23 NRC (National Research Council). (1983) Risk assessment in the federal government: managing the process.
24 Washington, DC: National Academy Press.
- 25 Oberdörster, G. (1995) Lung particle overload: implications for occupational exposures to particles. *Reg Toxicol*
26 *Pharmacol* 27:123–135.
- 27 O’Neil, MJ; ed. (2001) The Merck index: an encyclopedia of chemicals, drugs, and biologicals. 13th edition.
28 Whitehouse Station, NJ: Merck and Co., Inc.; pp. 89, 342–344.
- 29 Pairon, JC; Roos, F; Iwatsubo, Y; et al. (1994) Lung retention of cerium in humans. *Occup Environ Med* 51(3):195–
30 199.
- 31 Pairon, JC; Roos, F; Sebastien, P; et al. (1995) Biopersistence of cerium in the human respiratory tract and
32 ultrastructural findings. *Am J Indus Med* 27(3):349–358.
- 33 Palmer, RJ; Butenhoff, JL; Stevens, JB. (1987) Cytotoxicity of the rare earth metals cerium, lanthanum, and
34 neodymium in vitro: comparisons with cadmium in a pulmonary macrophage primary culture system. *Environ Res*
35 43(1):142–156.
- 36 Pietra, R; Pietra, R; Sabbioni, E; et al. (1985) Trace elements in tissues of a worker affected by rare-earth
37 pneumoconiosis. A study carried out by neutron activation analysis. *J Radioanal Nucl Chem* 92(2):247–269.
- 38 Porru, S; Placidi, D; Quarta, C; et al. (2001) The potential role of rare earths in the pathogenesis of interstitial lung
39 disease: a case report of movie projectionist as investigated by neutron activation analysis. *J Trace Elem Med Biol*
40 14(4):232–236.

- 1 Preeta, R; Nair, R. (1999) Stimulation of cardiac fibroblast proliferation by cerium: a superoxide anion-mediated
2 response. *J Mol Cell Cardiol* 31(8):1573–1580.
- 3 Reinhardt, K; Winkler, H. (1996) Cerium mischmetal, cerium alloys, and cerium compounds. In: Ullmann's
4 encyclopedia of industrial chemistry. Vol. A6. Weinheim, Germany: Wiley-VCH; pp. 139–152.
- 5 Reinhardt K; Winkler H. (2002) Cerium mischmetal, cerium alloys, and cerium compounds. In: Ullmann's
6 encyclopedia of industrial chemistry. Vol. 7. Weinheim, Germany: Wiley-VCH, pp. 285–300. Available online with
7 subscription at http://www.mrw.interscience.wiley.com/ueic/ueic_search_fs.html.
- 8 Rhodia, Inc. (1998) Initial submission: letter from Rhodia, Inc. to US EPA re acute oral toxicity study in rats of
9 lithium sulfide, sodium sulfide, and cerium sulfide, with attachments and dated 11/23/1998. Produced by
10 Huntingdon Life Sciences Ltd., Cambridgeshire, England, for Rhodia Inc., Cranbury, NJ. Submitted under TSCA
11 Section 8E; EPA Doc. No. 88990000042; NTIS No. OTS0559644.
- 12 Rhone-Poulenc. (1983) Ceric oxide: acute toxicity studies via the respiratory tract in the rat. Rhone-Poulenc French
13 Technology Institute, Courbevoie Ceux, France; Report 302217. (as cited in HEI, 2001).
- 14 Sabbioni, E; Pietra, R; Gaglione, P; et al. (1982) Long-term occupational risk of rare-earth pneumoconiosis: a case
15 report as investigated by neutron activation analysis. *Sci Total Environ* 26:19–32.
- 16 Salas, M; Tuchweber, B; Kovacs, K; et al. (1976) Effect of cerium on the rat liver: an ultrastructural and biochemical
17 study. *Beitr Pathol* 157(1):23–44.
- 18 Salonpää, P; Iscan, M; Pasanen, M; et al. (1992) Cerium-induced strain-dependent increase in Cyp2a-4/5
19 (cytochrome P4502a-4/5) expression in the liver and kidneys of inbred mice. *Biochem Pharmacol* 44(7):1269–1274.
- 20 Schubert, D; Dargusch, R; Raitano, J; et al. (2006) Cerium and yttrium oxide nanoparticles are neuroprotective.
21 *Biochem Biophys Res Commun* 342:86–91
- 22 Schulz, H; Brand, P; Heyder, J. (2000) Particle deposition in the respiratory tract. In: Gehr, P; Heyder, J; eds.
23 Particle-lung interactions. New York, NY: Marcel Dekker; pp. 229–290.
- 24 Sharma, A; Talukder, G. (1987) Effects of metals on chromosomes of higher organisms. *Environ Mutat* 9:191–226.
- 25 Shimizu, H; Suzuki, Y; Takemura, N; et al. (1985) The results of microbial mutation test for forty-three industrial
26 chemicals. *Sangyo Igaku (Jpn J Ind Health)* 27:400–419.
- 27 Shiraishi, Y; Ichikawa, R. (1972) Absorption and retention of ¹⁴⁴Ce and ⁹⁵Zr-⁹⁵Nb in newborn, juvenile and adult
28 rats. *Health Phys* 22:373–378.
- 29 Shivakumar, K; Nair, RR. (1991) Cerium depresses protein synthesis in cultured cardiac myocytes and lung
30 fibroblasts. *Mol Cell Biochem* 100:91–96.
- 31 Shivakumar, K; Nair, RR; Valiathan, MS. (1992) Paradoxical effect of cerium on collagen synthesis in cardiac
32 fibroblasts. *J Mol Cell Cardiol.* 24:775–780.
- 33 Shrivastava, S; Mathur, R. (2004) Effectiveness of ethylene glycol bis (2-aminoethyl ether) tetraacetic acid (EGTA)
34 against cerium toxicity. *Indian J Exp Biol* 42:876–883.
- 35 Stineman, CH; Massaro, EJ; Lown, BA; et al. (1978) Cerium tissue/organ distribution and alterations in open field
36 and exploratory behavior following acute exposure of the mouse to cerium (citrate). *J Environ Pathol Toxicol*
37 2(2):553–570.
- 38 Strubelt, O; Siegers, CP; Younes, M. (1980) The influence of silybin on the hepatotoxic and hypoglycemic effects of
39 praseodymium and other lanthanides. *Drug Res* 30(10):1690–1694.

- 1 Sturbaum, B; Brooks, AL; McClellan, RO. (1970) Tissue distribution and dosimetry of ¹⁴⁴Ce in Chinese hamsters.
2 Radiat Res 44:359–367.
- 3 Sulotto, F; Romano, C; Berra, A; et al. (1986) Rare-earth pneumoconiosis: a new case. Am J Ind Med 9(6):567–
4 575.
- 5 Talbot, RB; Davison, FG; Green, JW; et al. (1965) Effects of subcutaneous implantation of rare earth metals.
6 Produced by the Iowa State University of Science and Technology for the U.S. Atomic Energy Commission;
7 USAEC Report 1170. Available from the National Technical Information Service, Springfield, VA; COO11701.
- 8 Thill, A; Zeyons, O; Spalla, O; et al. (2006) Cytotoxicity of CeO₂ nanoparticles for *Escherichia coli*. Physico-
9 chemical insight of the cytotoxicity mechanism. Environ Sci Technol 40(19):6151–6156.
- 10 Thomas, RL; Scott, JK; Chiffelle, TL. (1972) Metabolism and toxicity of inhaled ¹⁴⁴Ce in rats. Radiat Res 49:589–
11 610.
- 12 U.S. EPA (Environmental Protection Agency). (1986a) Guidelines for the health risk assessment of chemical
13 mixtures. Federal Register 51(185):34014–34025. Available online at <http://www.epa.gov/ncea/raf/rafguid.htm>.
- 14 U.S. EPA (Environmental Protection Agency). (1986b) Guidelines for mutagenicity risk assessment. Federal
15 Register 51(185):34006–34012. Available online at <http://www.epa.gov/ncea/raf/rafguid.htm>.
- 16 U.S. EPA (Environmental Protection Agency). (1987) Recommendations for and documentation of biological values
17 for use in risk assessment. Environmental Criteria and Assessment Office, Office of Health and Environmental
18 Assessment, Cincinnati, OH; EPA/600/6-87/008. Available from the National Technical Information Service,
19 Springfield, VA; PB88-179874/AS, and online at <http://cfpub.epa.gov/ncea/cfm/recordisplay.cfm?deid=34855>.
- 20 U.S. EPA (Environmental Protection Agency). (1991) Guidelines for developmental toxicity risk assessment.
21 Federal Register 56(234):63798–63826. Available online at <http://www.epa.gov/ncea/raf/rafguid.htm>.
- 22 U.S. EPA (Environmental Protection Agency). (1994a) Interim policy for particle size and limit concentration issues
23 in inhalation toxicity: notice of availability. Federal Register 59(206):53799. Available online at
24 <http://www.epa.gov/EPA-PEST/1994/October/Day-26/pr-11.html>.
- 25 U.S. EPA (Environmental Protection Agency). (1994b) Methods for derivation of inhalation reference
26 concentrations and application of inhalation dosimetry. Environmental Criteria and Assessment Office, Office of
27 Health and Environmental Assessment, Cincinnati, OH; EPA/600/8-90/066F. Available from the National Technical
28 Information Service, Springfield, VA, PB2000-500023, and online at
29 <http://cfpub.epa.gov/ncea/raf/recordisplay.cfm?deid=71993>.
- 30 U.S. EPA (Environmental Protection Agency). (1995) Use of the benchmark dose approach in health risk
31 assessment. Risk Assessment Forum, Washington, DC; EPA/630/R-94/007. Available from the National Technical
32 Information Service, Springfield, VA, PB95-213765, and online at
33 http://cfpub.epa.gov/ncea/raf/raf_pubtitles.cfm?detype=document&excCol=archive.
- 34 U.S. EPA (Environmental Protection Agency). (1996) Guidelines for reproductive toxicity risk assessment. Federal
35 Register 61(212):56274–56322. Available online at <http://www.epa.gov/ncea/raf/rafguid.htm>.
- 36 U.S. EPA (Environmental Protection Agency). (1996) Proposed guidelines for carcinogen risk assessment. Federal
37 Register 61:17960–18011. Available online at <http://www.epa.gov/cancerguidelines>.
- 38 U.S. EPA (Environmental Protection Agency). (1998) Guidelines for neurotoxicity risk assessment. Federal
39 Register 63(93):26926–26954. Available online at <http://www.epa.gov/ncea/raf/rafguid.htm>.
- 40 U.S. EPA (Environmental Protection Agency). (2000a) Science policy council handbook: risk characterization.
41 Office of Science Policy, Office of Research and Development, Washington, DC. EPA/100-B-00-002. Available
42 online at <http://www.epa.gov/OSA/spc/pdfs/prhandbk.pdf>.

- 1 U.S. EPA (Environmental Protection Agency). (2000b) Benchmark dose technical guidance document [external
2 review draft]. Risk Assessment Forum, Washington, DC; EPA/630/R-00/001. Available online at
3 [http://cfpub.epa.gov/ncea/cfm/
4 nceapublication.cfm?ActType=PublicationTopics&detype=DOCUMENT&subject
=BENCHMARK+DOSE&subjtype=TITLE&excCol=Archive.](http://cfpub.epa.gov/ncea/cfm/nceapublication.cfm?ActType=PublicationTopics&detype=DOCUMENT&subject=BENCHMARK+DOSE&subjtype=TITLE&excCol=Archive)
- 5 U.S. EPA (Environmental Protection Agency). (2000c) Supplementary guidance for conducting health risk
6 assessment of chemical mixtures. Risk Assessment Forum, Washington, DC; EPA/630/R-00/002. Available online
7 at http://cfpub.epa.gov/ncea/raf/chem_mix.cfm.
- 8 U.S. EPA (Environmental Protection Agency). (2002) A review of the reference dose concentration and reference
9 concentration processes. Risk Assessment Forum, Washington, DC; EPA/630/P-02/002F. Available online at
10 [http://cfpub.epa.gov/ncea/raf/raf_
pubtitles.cfm?detype=document&excCol=archive.](http://cfpub.epa.gov/ncea/raf/raf_pubtitles.cfm?detype=document&excCol=archive)
- 11 U.S. EPA (Environmental Protection Agency). (2005a) Guidelines for carcinogen risk assessment. Federal Register
12 70(66):17765–18717. Available online at <http://www.epa.gov/cancerguidelines>.
- 13 U.S. EPA (Environmental Protection Agency). (2005b) Supplemental guidance for assessing susceptibility from
14 early-life exposure to carcinogens. Risk Assessment Forum, Washington, DC; EPA/630/R-03/003F. Available
15 online at <http://www.epa.gov/cancerguidelines>.
- 16 U.S. EPA (Environmental Protection Agency). (2006a) Science policy council handbook: peer review. 3rd edition.
17 Office of Science Policy, Office of Research and Development, Washington, DC; EPA/100/B-06/002. Available
18 online at <http://www.epa.gov/OSA/spc/2peerrev.htm>.
- 19 U.S. EPA. (2006b) A framework for assessing health risk of environmental exposures to children. National Center
20 for Environmental Assessment, Washington, DC; EPA/600/R-05/093F. Available online at
21 <http://cfpub.epa.gov/ncea/cfm/recordisplay.cfm?deid=158363>.
- 22 Valiathan, MS; Kartha, CC; Eapen, JT; et al. (1989) A geochemical basis for endomyocardial fibrosis. ICMR Centre
23 for Research in Cardiomyopathy, Trivandrum, India. *Cardiovasc Res* 23(7):647–648.
- 24 Vocaturo, G; Colombo, F; Zanoni, M; et al. (1983) Human exposure to heavy metals: rare earth pneumoconiosis in
25 occupational workers. *Chest* 83(5):780–783.
- 26 Vogt, P; Spycher, MA; Ruttner, JR. (1986) [Pneumoconiosis caused by “rare earths” (cer-pneumoconiosis)].
27 *Schweiz Med Wochenscher* 116:1303–1308. (German with English summary).
- 28 Waring, PM; Watling, RJ. (1990) Rare earth deposits in a deceased movie projectionist: a new case of rare earth
29 pneumoconiosis? *Med J Aust* 153(11/12):726–730.
- 30 Wells, WH, Jr; Wells, VL. (2001) The lanthanides, rare earth metals. In: Bingham, E; Cohrssen, B; Powell, C, II;
31 eds. *Patty’s toxicology*. 5th edition. New York, NY: John Wiley and Sons, Inc.; pp 423–458.
- 32 Wiener-Schmuck, M; Lind, I; Polzer, G; et al. (1990) In vivo and in vitro effects of rare earth compounds. *J Aerosol*
33 *Sci* 21(1):S505–S508.
- 34 Witschi, HR; Last, JA. (2001) Toxic responses of the respiratory system. In: Klaassen, CD; ed. *Casarett and Doull’s*
35 *toxicology: the basic science of poisons*. 6th edition. New York, NY: McGraw-Hill; pp. 515–534.
- 36 Wulfsberg, G. (2000) *Inorganic chemistry*. Sausalito, CA: University Science Books; p. 59.
- 37 Yoon, HK; Moon, HS; Park, SH; et al. (2005) Dendriiform pulmonary ossification in patient with rare earth
38 pneumoconiosis. *Thorax* 60(8):701–703.
- 39 Yu, L; Chen, Z; Wang, Y. (2001) Effect of rare earth element cerium (Ce) on abnormality rate of sperm and
40 testosterone secretion in sera of male mice. *Nanjing Nongye Daxue Xuebao* 24(1):77–80. (Chinese with English
41 abstract).

