

1 exposure by itself. Available estimates suggest that exposures to most of these TCE-related
2 compounds are comparable to or greater than TCE itself.

3 4 **6.1.2. Toxicokinetics and Physiologically-Based Pharmacokinetic (PBPK) Modeling** (*see* 5 *Chapter 3 and Appendix A*)

6 TCE is a lipophilic compound that readily crosses biological membranes. Exposures may
7 occur via the oral, dermal, and inhalation route, with evidence for systemic availability from
8 each route. TCE can also be transferred transplacentally and through breast milk ingestion. TCE
9 is rapidly and nearly completely absorbed from the gut following oral administration, and animal
10 studies indicate that exposure vehicle may impact the time course of absorption: oily vehicles
11 may delay absorption whereas aqueous vehicles result in a more rapid increase in blood
12 concentrations. See Section 3.1 for additional discussion of TCE absorption.

13 Following absorption to the systemic circulation, TCE distributes from blood to solid
14 tissues by each organ's solubility. This process is mainly determined by the blood:tissue
15 partition coefficients, which are largely determined by tissue lipid content. Adipose partitioning
16 is high, so adipose tissue may serve as a reservoir for TCE, and accumulation into adipose tissue
17 may prolong internal exposures. TCE attains high concentrations relative to blood in the brain,
18 kidney, and liver—all of which are important target organs of toxicity. TCE is cleared via
19 metabolism mainly in three organs: the kidney, liver, and lungs. See Section 3.2 for additional
20 discussion of TCE distribution.

21 The metabolism of TCE is an important determinant of its toxicity. Metabolites are
22 generally thought to be responsible for toxicity—especially for the liver and kidney. Initially,
23 TCE may be oxidized via cytochrome P450 (CYP) isoforms or conjugated with glutathione by
24 glutathione S-transferase enzymes. While CYP2E1 is generally accepted to be the CYP isoform
25 most responsible for TCE oxidation, others forms may also contribute. There are conflicting
26 data as to which glutathione-S-transferase (GST) isoforms are responsible for TCE conjugation,
27 with one rat study indicating alpha-class GSTs and another rat study indicating mu and pi-class
28 GST. The balance between oxidative and conjugative metabolites generally favors the oxidative
29 pathway, especially at lower concentrations, and inhibition of CYP-dependent oxidation *in vitro*
30 increases glutathione (GSH) conjugation in renal preparations. However, in humans, direct
31 comparison of *in vitro* rates of oxidation and conjugation, as well as *in vivo* data on the amount
32 of the TCE GSH conjugation to dichlorovinyl glutathione in blood, support a flux through the
33 GSH pathway that may be one or more orders of magnitude greater than the <0.1% inferred from
34 excretion of GSH conjugation derived urinary mercapturates. See Section 3.3 for additional
35 discussion of TCE metabolism.

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1 Once absorbed, TCE is excreted primarily either in breath as unchanged TCE or carbon
2 dioxide [CO₂], or in urine as metabolites. Minor pathways of elimination include excretion of
3 metabolites in saliva, sweat, and feces. Following oral administration or upon cessation of
4 inhalation exposure, exhalation of unmetabolized TCE is a major elimination pathway. Initially,
5 elimination of TCE upon cessation of inhalation exposure demonstrates a steep concentration-
6 time profile: TCE is rapidly eliminated in the minutes and hours postexposure, and then the rate
7 of elimination via exhalation decreases. Following oral or inhalation exposure, urinary
8 elimination of parent TCE is minimal, with urinary elimination of the metabolites trichloroacetic
9 acid and trichloroethanol accounting for the bulk of the absorbed dose of TCE. See Section 3.4
10 for additional discussion of TCE excretion.

11 As part of this assessment, a comprehensive Bayesian PBPK model-based analysis of the
12 population toxicokinetics of TCE and its metabolites was developed in mice, rats, and humans
13 (also reported in Chiu et al., 2009). This analysis considered a wider range of physiological,
14 chemical, *in vitro*, and *in vivo* data than any previously published analysis of TCE. The
15 toxicokinetics of the “population average,” its population variability, and their uncertainties are
16 characterized and estimates of experimental variability and uncertainty are included in this
17 analysis. The experimental database included separate sets for model calibration and evaluation
18 for rats and humans; fewer data were available in mice, and were all used for model calibration.
19 The total combination of these approaches and PBPK analysis substantially supports the model
20 predictions. In addition, the approach employed yields an accurate characterization of the
21 uncertainty in metabolic pathways for which available data were sparse or relatively indirect,
22 such as GSH conjugation and respiratory tract metabolism. Key conclusions from the model
23 predictions include (1) as expected, TCE is substantially metabolized, primarily by oxidation at
24 doses below saturation; (2) GSH conjugation and subsequent bioactivation in humans appears to
25 be 10- to 100-fold greater than previously estimated; and (3) mice had the greatest rate of
26 respiratory tract oxidative metabolism compared to rats and humans. The predictions of the
27 PBPK model are subsequently used in noncancer and cancer dose-response analyses for inter-
28 and intraspecies extrapolation of toxicokinetics (see below). See Section 3.5 and Appendix A for
29 additional discussion of and details about PBPK modeling of TCE and metabolites.

31 **6.1.3. Noncancer Toxicity**

32 This section summarizes the weight of evidence for TCE noncancer toxicity. Based on
33 the available human epidemiologic data and experimental and mechanistic studies, it is
34 concluded that TCE poses a potential human health hazard for noncancer toxicity to the central
35 nervous system, the kidney, the liver, the immune system, the male reproductive system, and the

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1 developing fetus. The evidence is more limited for TCE toxicity to the respiratory tract and
2 female reproductive system. The conclusions pertaining to specific endpoints within these
3 tissues and systems are summarized below.

4
5 **6.1.3.1. Neurological Effects (see Sections 4.3 and 4.11.1.1 and Appendix D)**

6 Both human and animal studies have associated TCE exposure with effects on several
7 neurological domains. Multiple epidemiologic studies in different populations have reported
8 abnormalities in trigeminal nerve function in association with TCE exposure. Two small studies
9 did not report an association between TCE exposure and trigeminal nerve function. However,
10 statistical power was limited, exposure misclassification was possible, and, in one case, methods
11 for assessing trigeminal nerve function were not available. As a result, these studies do not
12 provide substantial evidence against a causal relationship between TCE exposure and trigeminal
13 nerve impairment. Laboratory animal studies have also demonstrated TCE-induced changes in
14 the morphology of the trigeminal nerve following short-term exposures in rats. However, one
15 study reported no significant changes in trigeminal somatosensory evoked potential in rats
16 exposed to TCE for 13 weeks. See Section 4.3.1 for additional discussion of studies of
17 alterations in nerve conduction and trigeminal nerve effects. Human chamber, occupational, and
18 geographic based/drinking water studies have consistently reported subjective symptoms such as
19 headaches, dizziness, and nausea which are suggestive of vestibular system impairments. One
20 study reported changes in nystagmus threshold (a measure of vestibular system function)
21 following an acute TCE exposure. There are only a few laboratory animal studies relevant to
22 this neurological domain, with reports of changes in nystagmus, balance, and handling reactivity.
23 See Section 4.3.3 for additional discussion of TCE effects on vestibular function. Fewer and
24 more limited epidemiologic studies are suggestive of TCE exposure being associated with
25 delayed motor function, and changes in auditory, visual, and cognitive function or performance
26 (see Sections 4.3.2, 4.3.4, 4.3.5, and 4.3.6). Acute and subchronic animal studies show
27 disruption of the auditory system, changes in visual evoked responses to patterns or flash
28 stimulus, and neurochemical and molecular changes. Animal studies suggest that while the
29 effects on the auditory system lead to permanent function impairments and histopathology,
30 effects on the visual system may be reversible with termination of exposure. Additional acute
31 studies reported structural or functional changes in hippocampus, such as decreased myelination
32 or decreased excitability of hippocampal CA1 neurons, although the relationship of these effects
33 to overall cognitive function is not established (see Section 4.3.9). An association between TCE
34 exposure and sleep changes has also been demonstrated in rats (see Section 4.3.7). Some
35 evidence exists for motor-related changes in rats/mice exposed acutely/subchronically to TCE,

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1 but these effects have not been reported consistently across all studies (see Section 4.3.6).
2 Gestational exposure to TCE in humans has been reported to be associated with
3 neurodevelopmental abnormalities including neural tube defects, encephalopathy, impaired
4 cognition, aggressive behavior, and speech and hearing impairment. Developmental
5 neurotoxicological changes have also been observed in animals including aggressive behaviors
6 following an *in utero* exposure to TCE and a suggestion of impaired cognition as noted by
7 decreased myelination in the CA1 hippocampal region of the brain. See Section 4.3.8 for
8 additional discussion of developmental neurological effects of TCE. Therefore, overall, the
9 strongest neurological evidence of human toxicological hazard is for changes in trigeminal nerve
10 function or morphology and impairment of vestibular function, based on both human and
11 experimental studies, while fewer and more limited evidence exists for delayed motor function,
12 changes in auditory, visual, and cognitive function or performance, and neurodevelopmental
13 outcomes.

14 15 **6.1.3.2. *Kidney Effects (see Sections 4.4.1, 4.4.4, 4.4.6, and 4.11.1.2)***

16 Kidney toxicity has also been associated with TCE exposure in both human and animal
17 studies. There are few human data pertaining to TCE-related noncancer kidney toxicity;
18 however, several available studies reported elevated excretion of urinary proteins, considered
19 nonspecific markers of nephrotoxicity, among TCE-exposed subjects compared to unexposed
20 controls. While some of these studies include subjects previously diagnosed with kidney cancer,
21 other studies report similar results in subjects that are disease free. Some additional support for
22 TCE nephrotoxicity in humans is provided by a study reporting a greater incidence of end-stage
23 renal disease in TCE-exposed workers as compared to unexposed controls, although some
24 subjects in this study were also exposed to hydrocarbons, JP-4 gasoline, and multiple solvents,
25 including TCE and 1,1,1-trichloroethane. See Section 4.4.1 for additional discussion of human
26 data on the noncancer kidney effects of TCE. Laboratory animal and *in vitro* data provide
27 additional support for TCE nephrotoxicity. TCE causes renal toxicity in the form of cytomegaly
28 and karyomegaly of the renal tubules in male and female rats and mice following either oral or
29 inhalation exposure. In rats, the pathology of TCE-induced nephrotoxicity appears distinct from
30 age-related nephropathy. Increased kidney weights have also been reported in some rodent
31 studies. See Section 4.4.4 for additional discussion of laboratory animal data on the noncancer
32 kidney effects of TCE. Further studies with TCE metabolites have demonstrated a potential role
33 for dichlorovinyl cysteine (DCVC), trichloroethanol, and trichloroacetic acid (TCA) in TCE-
34 induced nephrotoxicity. Of these, available data suggest that DCVC induced renal effects are
35 most similar to those of TCE and that DCVC is formed in sufficient amounts following TCE

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1 exposure to account for these effects. TCE or DCVC have also been shown to be cytotoxic to
2 primary cultures of rat and human renal tubular cells. See Section 4.4.6 for additional discussion
3 on the role of metabolism in the noncancer kidney effects of TCE. Overall, multiple lines of
4 evidence support the conclusion that TCE causes nephrotoxicity in the form of tubular toxicity,
5 mediated predominantly through the TCE GSH conjugation product DCVC.
6

7 **6.1.3.3. Liver Effects (see Sections 4.5.1, 4.5.3, 4.5.4, 4.5.6, and 4.11.1.3, and Appendix E)**

8 Liver toxicity has also been associated with TCE exposure in both human and animal
9 studies. Although there are few human studies on liver toxicity and TCE exposure, several
10 available studies have reported TCE exposure to be associated with significant changes in serum
11 liver function tests, widely used in clinical settings in part to identify patients with liver disease,
12 or changes in plasma or serum bile acids. Additional, more limited human evidence for TCE
13 induced liver toxicity includes reports suggesting an association between TCE exposure and liver
14 disorders, and case reports of liver toxicity including hepatitis accompanying immune-related
15 generalized skin diseases, jaundice, hepatomegaly, hepatosplenomegaly, and liver failure in
16 TCE-exposed workers. Cohort studies examining cirrhosis mortality and either TCE exposure or
17 solvent exposure are generally null, but these studies cannot rule out an association with TCE
18 because of their use of death certificates where there is a high degree (up to 50%) of
19 underreporting. Overall, while some evidence exists of liver toxicity as assessed from liver
20 function tests, the data are inadequate for making conclusions regarding causality. See
21 Section 4.5.1 for additional discussion of human data on the noncancer liver effects of TCE. In
22 rats and mice, TCE exposure causes hepatomegaly without concurrent cytotoxicity. Like
23 humans, laboratory animals exposed to TCE have been observed to have increased serum bile
24 acids, although the toxicological importance of this effect is unclear. Other effects in the rodent
25 liver include small transient increases in DNA synthesis, cytomegaly in the form of “swollen” or
26 enlarged hepatocytes, increased nuclear size probably reflecting polyploidization, and
27 proliferation of peroxisomes. Available data also suggest that TCE does not induce substantial
28 cytotoxicity, necrosis, or regenerative hyperplasia, as only isolated, focal necroses and mild to
29 moderate changes in serum and liver enzyme toxicity markers having been reported. These
30 effects are consistently observed across rodent species and strains, although the degree of
31 response at a given mg/kg/d dose appears to be highly variable across strains, with mice on
32 average appearing to be more sensitive. See Sections 4.5.3 and 4.5.4 for additional discussion of
33 laboratory animal data on the noncancer liver effects of TCE. While it is likely that oxidative
34 metabolism is necessary for TCE-induced effects in the liver, the specific metabolite or
35 metabolites responsible is less clear. However, the available data are strongly inconsistent with

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1 TCA being the sole or predominant active moiety for TCE-induced liver effects, particularly
2 with respect to hepatomegaly. See Section 4.5.6 for additional discussion on the role of
3 metabolism in the noncancer liver effects of TCE. Overall, TCE, likely through its oxidative
4 metabolites, clearly leads to liver toxicity in laboratory animals, with mice appearing to be more
5 sensitive than other laboratory animal species, but there is only limited epidemiologic evidence
6 of hepatotoxicity being associated with TCE exposure.

8 **6.1.3.4. Immunological Effects (see Sections 4.6.1.1, 4.6.2, and 4.11.1.4)**

9 Effects related the immune system have also been associated with TCE exposure in both
10 human and animal studies. A relationship between systemic autoimmune diseases, such as
11 scleroderma, and occupational exposure to TCE has been reported in several recent studies, and a
12 meta-analysis of scleroderma studies resulted in a statistically significant combined odds ratio for
13 any exposure in men (odds ratio [OR]: 2.5, 95% confidence interval [CI]: 1.1, 5.4), with a lower
14 relative risk seen in women in women (OR: 1.2, 95% CI: 0.58, 2.6). The human data at this time
15 do not allow a determination of whether the difference in effect estimates between men and
16 women reflects the relatively low background risk of scleroderma in men, gender-related
17 differences in exposure prevalence or in the reliability of exposure assessment, a gender-related
18 difference in susceptibility to the effects of TCE, or chance. Additional human evidence for the
19 immunological effects of TCE includes studies reporting TCE-associated changes in levels of
20 inflammatory cytokines in occupationally-exposed workers and infants exposed via indoor air at
21 air concentrations typical of such exposure scenarios (see Section 6.1.1, above); a large number
22 of case reports (mentioned above) of a severe hypersensitivity skin disorder, distinct from
23 contact dermatitis and often accompanied by hepatitis; and a reported association between
24 increased history of infections and exposure to TCE contaminated drinking water. See
25 Section 4.6.1.1 for additional discussion of human data on the immunological effects of TCE.
26 Immunotoxicity has also been reported in experimental rodent studies of TCE. Numerous
27 studies have demonstrated accelerated autoimmune responses in autoimmune-prone mice,
28 including changes in cytokine levels similar to those reported in human studies, with more severe
29 effects, including autoimmune hepatitis, inflammatory skin lesions, and alopecia, manifesting at
30 longer exposure periods. Immunotoxic effects have been also reported in B6C3F1 mice, which
31 do not have a known particular susceptibility to autoimmune disease. Developmental
32 immunotoxicity in the form of hypersensitivity responses have been reported in TCE-treated
33 guinea pigs and mice via drinking water pre- and postnatally. Evidence of localized
34 immunosuppression has also been reported in mice and rats. See Section 4.6.2 for additional
35 discussion of laboratory animal data on the immunological effects of TCE. Overall, the human

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1 and animal studies of TCE and immune-related effects provide strong evidence for a role of TCE
2 in autoimmune disease and in a specific type of generalized hypersensitivity syndrome, while
3 there are less data pertaining to immunosuppressive effects.

4
5 **6.1.3.5. *Respiratory Tract Effects (see Sections 4.7.1.1, 4.7.2.1, 4.7.3, and 4.11.1.5)***

6 The very few human data on TCE and pulmonary toxicity are too limited for drawing
7 conclusions (see Section 4.7.1.1), but laboratory studies in mice and rats have shown toxicity in
8 the bronchial epithelium, primarily in Clara cells, following acute exposures to TCE (see
9 Section 4.7.2.1). A few studies of longer duration have reported more generalized toxicity, such
10 as pulmonary fibrosis in mice and pulmonary vasculitis in rats. However, respiratory tract
11 effects were not reported in other longer-term studies. Acute pulmonary toxicity appears to be
12 dependent on oxidative metabolism, although the particular active moiety is not known. While
13 earlier studies implicated chloral produced *in situ* by CYP enzymes in respiratory tract tissue in
14 toxicity, the evidence is inconsistent and several other possibilities are viable. Although humans
15 appear to have lower overall capacity for enzymatic oxidation in the lung relative to mice, CYP
16 enzymes do reside in human respiratory tract tissue, suggesting that, qualitatively, the respiratory
17 tract toxicity observed in rodents is biologically plausible in humans. See Section 4.7.3 for
18 additional discussion of the role of metabolism in the noncancer respiratory tract toxicity of
19 TCE. Therefore, overall, data are suggestive of TCE causing respiratory tract toxicity, based
20 primarily on short-term studies in mice and rats, with available human data too few and limited
21 to add to the weight of evidence for pulmonary toxicity.

22
23 **6.1.3.6. *Reproductive Effects (see Sections 4.8.1 and 4.11.1.6)***

24 A number of human and laboratory animal studies suggest that TCE exposure has the
25 potential for male reproductive toxicity, with a more limited number of studies examining female
26 reproductive toxicity. Human studies have reported TCE exposure to be associated (in all but
27 one case statistically-significantly) with increased sperm density and decreased sperm quality,
28 altered sexual drive or function, or altered serum endocrine levels. Measures of male fertility,
29 however, were either not reported or reported to be unchanged with TCE exposure, though the
30 statistical power of the available studies is quite limited. Epidemiologic studies have identified
31 possible associations of TCE exposure with effects on female fertility and with menstrual cycle
32 disturbances, but these data are fewer than those available for male reproductive toxicity. See
33 Section 4.8.1.1 for additional discussion of human data on the reproductive effects of TCE.
34 Evidence of similar effects, particularly for male reproductive toxicity, is provided by several
35 laboratory animal studies that reported effects on sperm, libido/copulatory behavior, and serum

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1 hormone levels, although some studies that assessed sperm measures did not report treatment-
2 related alterations. Additional adverse effects on male reproduction have also been reported,
3 including histopathological lesions in the testes or epididymides and altered *in vitro* sperm-
4 oocyte binding or *in vivo* fertilization due to TCE or metabolites. While reduced fertility in
5 rodents was only observed in one study, this is not surprising given the redundancy and
6 efficiency of rodent reproductive capabilities. In addition, although the reduced fertility
7 observed in the rodent study was originally attributed to systemic toxicity, the database as a
8 whole suggests that TCE does induce reproductive toxicity independent of systemic effects.
9 Fewer data are available in rodents on female reproductive toxicity. While *in vitro* oocyte
10 fertilizability has been reported to be reduced as a result of TCE exposure in rats, a number of
11 other laboratory animal studies did not report adverse effects on female reproductive function.
12 See Section 4.8.1.2 for additional discussion of laboratory animal data on the reproductive
13 effects of TCE. Very limited data are available to elucidate the mode of action (MOA) for these
14 effects, though some aspects of a putative MOA (e.g., perturbations in testosterone biosynthesis)
15 appear to have some commonalities between humans and animals (see Section 4.8.1.3.2).
16 Together, the human and laboratory animal data support the conclusion that TCE exposure poses
17 a potential hazard to the male reproductive system, but are more limited with regard to the
18 potential hazard to the female reproductive system.

20 **6.1.3.7. Developmental Effects (see Sections 4.8.3 and 4.11.1.7)**

21 The relationship between TCE exposure (direct or parental) and developmental toxicity
22 has been investigated in a number of epidemiologic and laboratory animal studies. Postnatal
23 developmental outcomes examined include developmental neurotoxicity (addressed above with
24 neurotoxicity), developmental immunotoxicity (addressed above with immunotoxicity), and
25 childhood cancers. Prenatal effects examined include death (spontaneous abortion, perinatal
26 death, pre- or postimplantation loss, resorptions), decreased growth (low birth weight, small for
27 gestational age, intrauterine growth restriction, decreased postnatal growth), and congenital
28 malformations, in particular cardiac defects. Some epidemiological studies have reported
29 associations between parental exposure to TCE and spontaneous abortion or perinatal death, and
30 decreased birth weight or small for gestational age, although other studies reported mixed or null
31 findings. While comprising both occupational and environmental exposures, these studies are
32 overall not highly informative due to the small numbers of cases and limited exposure
33 characterization or to the fact that exposures were to a mixture of solvents. See Section 4.8.3.1
34 for additional discussion of human data on the developmental effects of TCE. However,
35 multiple well conducted studies in rats and mice show analogous effects of TCE exposure: pre-

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1 or postimplantation losses, increased resorptions, perinatal death, and decreased birth weight.
2 Interestingly, the rat studies reporting these effects used Fischer 344 or Wistar rats, while several
3 other studies, all of which used Sprague-Dawley rats, reported no increased risk in these
4 developmental measures, suggesting a strain difference in susceptibility. See Section 4.8.3.2 for
5 additional discussion of laboratory animal data on the developmental effects of TCE. Therefore,
6 overall, based on weakly suggestive epidemiologic data and fairly consistent laboratory animal
7 data, it can be concluded that TCE exposure poses a potential hazard for prenatal losses and
8 decreased growth or birth weight of offspring.

9 With respect to congenital malformations, epidemiology and experimental animal studies
10 of TCE have reported increases in total birth defects, central nervous system defects, oral cleft
11 defects, eye/ear defects, kidney/urinary tract disorders, musculoskeletal birth anomalies,
12 lung/respiratory tract disorders, skeletal defects, and cardiac defects. Human occupational cohort
13 studies, while not consistently reporting positive results, are generally limited by the small
14 number of observed or expected cases of birth defects. While only one of the epidemiological
15 studies specifically reported observations of eye anomalies, studies in rats have identified
16 increases in the incidence of fetal eye defects following oral exposures during the period of
17 organogenesis with TCE or its oxidative metabolites dichloroacetic acid (DCA) and TCA. The
18 epidemiological studies, while individually limited, as a whole show relatively consistent
19 elevations, some of which were statistically significant, in the incidence of cardiac defects in
20 TCE-exposed populations compared to reference groups. In laboratory animal models, avian
21 studies were the first to identify adverse effects of TCE exposure on cardiac development, and
22 the initial findings have been confirmed multiple times. Additionally, administration of TCE and
23 its metabolites TCA and DCA in maternal drinking water during gestation has been reported to
24 induce cardiac malformations in rat fetuses. It is notable that a number of other studies, several
25 of which were well-conducted, did not report induction of cardiac defects in rats, mice, or rabbits
26 in which TCE was administered by inhalation or gavage. However, many of these studies used a
27 traditional free-hand section technique on fixed fetal specimens, and a fresh dissection technique
28 that can enhance detection of anomalies was used in the positive studies by Dawson et al. (1993)
29 and Johnson et al. (2003, 2005). Nonetheless, two studies that used the same or similar fresh
30 dissection technique did not report cardiac anomalies. Differences in other aspects of
31 experimental design may have been contributing factors to the differences in observed response.
32 In addition, mechanistic studies, such as the treatment-related alterations in endothelial cushion
33 development observed in avian *in ovo* and *in vitro* studies, provide a plausible mechanistic basis
34 for defects in septal and valvular morphogenesis observed in rodents, and consequently support
35 the plausibility of cardiac defects induced by TCE in humans. Therefore, while the studies by

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1 Dawson et al. (1993) and Johnson et al. (2003, 2005) have significant limitations, including the
2 lack of clear dose-response relationship for the incidence of any specific cardiac anomaly and the
3 pooling of data collected over an extended period, there is insufficient reason to dismiss their
4 findings. See Section 4.8.3.3.2 for additional discussion of the conclusions with respect to TCE-
5 induced cardiac malformations. Therefore, overall, based on weakly suggestive, but overall
6 consistent, epidemiologic data, in combination with evidence from experimental animal and
7 mechanistic studies, it can be concluded that TCE exposure poses a potential hazard for
8 congenital malformations, including cardiac defects, in offspring.

9
10 **6.1.4. Carcinogenicity** (*see Sections 4.1, 4.2, 4.4.2, 4.4.5, 4.4.7, 4.5.2, 4.5.5, 4.5.6, 4.5.7,*
11 *4.6.1.2, 4.6.2.4, 4.7.1.2, 4.7.2.2, 4.7.4, 4.8.2, 4.9, and 4.11.2, and Appendices B and C*)

12 In 1995, International Agency for Research on Cancer (IARC) concluded that
13 trichloroethylene is “probably carcinogenic to humans” (IARC, 1995). In 2000, National
14 Toxicology Program (NTP) concluded that trichloroethylene is “reasonably anticipated to be a
15 human carcinogen.” (NTP, 2000). In 2001, the draft U.S. Environmental Protection Agency
16 (U.S. EPA) health risk assessment of TCE concluded that TCE was “highly likely” to be
17 carcinogenic in humans. In 2006, a committee of the National Research Council stated that
18 “findings of experimental, mechanistic, and epidemiologic studies lead to the conclusion that
19 trichloroethylene can be considered a potential human carcinogen” (NRC, 2006).

20 Following U.S. EPA (2005a) *Guidelines for Carcinogen Risk Assessment*, based on the
21 available data as of 2009, TCE is characterized as “*Carcinogenic to Humans*” by all routes of
22 exposure. This conclusion is based on convincing evidence of a causal association between TCE
23 exposure in humans and kidney cancer. The consistency of increased kidney cancer relative risk
24 estimates across a large number of independent studies of different designs and populations from
25 different countries and industries provides compelling evidence given the difficulty, *a priori*, in
26 detecting effects in epidemiologic studies when the relative risks are modest, the cancers are
27 relatively rare, and therefore, individual studies have limited statistical power. This strong
28 consistency of the epidemiologic data on TCE and kidney cancer argues against chance, bias,
29 and confounding as explanations for the elevated kidney cancer risks. In addition, statistically
30 significant exposure-response trends are observed in high-quality studies. These studies were
31 designed to examine kidney cancer in populations with high TCE exposure intensity. These
32 studies addressed important potential confounders and biases, further supporting the observed
33 associations with kidney cancer as causal. See Section 4.4.2 for additional discussion of the
34 human epidemiologic data on TCE exposure and kidney cancer. In a meta-analysis of 14 high-
35 quality studies, a statistically significant pooled relative risk estimate was observed for overall
36 TCE exposure (RRp: 1.25 [95% CI: 1.11, 1.41]). The pooled relative risk estimate was greater

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1 for the highest TCE exposure groups (RRp: 1.53 [95% CI: 1.23, 1.91]; $n = 12$ studies). Meta-
2 analyses investigating the influence of individual studies and the sensitivity of the results to
3 alternate relative risk estimate selections found the pooled relative risk estimates to be highly
4 robust. Furthermore, there was no indication of publication bias or significant heterogeneity. It
5 would require a substantial amount of high-quality negative data to contradict this observed
6 association. See Section 4.4.2.5 and Appendix C for additional discussion of the kidney cancer
7 meta-analysis.

8 The human evidence of carcinogenicity from epidemiologic studies of TCE exposure is
9 compelling for lymphoma but less convincing than for kidney cancer. High quality studies
10 generally reported excess relative risk estimates, with statistically significant increases in three
11 studies, and a statistically significant trend with TCE exposure in one study (see Section 4.6.1.2).
12 The consistency of the association between TCE exposure and lymphoma is further supported by
13 the results of meta-analyses (see Section 4.6.1.2.2 and Appendix C). A statistically significant
14 pooled relative risk estimate was observed for overall TCE exposure (RRp: 1.23 [95% CI: 1.04,
15 1.44]), and, as with kidney cancer, the pooled relative risk estimate was greater for the highest
16 TCE exposure groups (RRp: 1.57 [95% CI: 1.27, 1.94]) than for overall TCE exposure.
17 Sensitivity analyses indicated that this result and its statistical significance were not overly
18 influenced by most individual studies or choice of individual (study-specific) risk estimates, and
19 in only one case was the resulting pooled relative risk estimates not statistically significant
20 (lower confidence bound of 1.00). Some heterogeneity was observed, particularly between
21 cohort and case-control studies, but it was not statistically significant. Notably, no heterogeneity
22 was observed in the meta-analysis of the highest exposure group, providing some evidence of
23 exposure misclassification as a source of heterogeneity in the overall analysis. In addition, there
24 was some evidence of potential publication bias. Thus, while the evidence is strong for
25 lymphoma, issues of study heterogeneity, potential publication bias, and weaker exposure-
26 response results contribute greater uncertainty.

27 The evidence is more limited for liver and biliary tract cancer mainly because only cohort
28 studies are available and most of these studies have small numbers of cases due the comparative
29 rarity of liver and biliary tract cancer. While most high quality studies reported excess relative
30 risk estimates, they were generally based on small numbers of cases or deaths, with the result of
31 wide confidence intervals on the estimates. The low number of liver cancer cases in the
32 available studies made assessing exposure-response relationships difficult. See Section 4.5.2 for
33 additional discussion of the human epidemiologic data on TCE exposure and liver cancer. A
34 consistency of the association between TCE exposure and liver cancer is supported by the results
35 of meta-analyses (see Section 4.5.2 and Appendix C). These meta-analyses found a statistically

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1 significant increased pooled relative risk estimate for liver and biliary tract cancer of 1.33 (95%
2 CI: 1.09, 1.64) with overall TCE exposure; but the meta-analyses using only the highest
3 exposure groups yielded a lower, and nonstatistically significant, pooled estimate for primary
4 liver cancer (1.28 [95% CI: 0.93, 1.77]). Although there was no evidence of heterogeneity or
5 publication bias and the pooled estimates were fairly insensitive to the use of alternative relative
6 risk estimates, the statistical significance of the pooled estimates depends heavily on the one
7 large study by Raaschou-Nielsen et al. (2003). There were fewer adequate, high quality studies
8 available for meta-analysis of liver cancer (9 versus 16 for lymphoma and 14 for kidney), leading
9 to lower statistical power, even with pooling. Thus, while there is epidemiologic evidence of an
10 association between TCE exposure and liver cancer, the much more limited database, both in
11 terms of number of available studies and number of cases upon which the studies are based,
12 contributes to greater uncertainty as compared to the evidence for kidney cancer or lymphoma.

13 There are several other lines of supporting evidence for TCE carcinogenicity in humans
14 by all routes of exposure. First, multiple chronic bioassays in rats and mice have reported
15 increased incidences of tumors with TCE treatment via inhalation and oral gavage, including
16 tumors in the kidney, liver, and lymphoid tissues—target tissues of TCE carcinogenicity also
17 seen in epidemiological studies. Of particular note is the site-concordant finding of low, but
18 biologically and sometimes statistically significant, increases in the incidence of kidney tumors
19 in multiple strains of rats treated with TCE by either inhalation or corn oil gavage (see
20 Section 4.4.5). The increased incidences were only detected at the highest tested doses, and were
21 greater in male than female rats; although, notably, pooled incidences in females from five rat
22 strains tested by NTP (1988, 1990) resulted in a statistically significant trend. Although these
23 studies have shown limited increases in kidney tumors, and several individual studies have a
24 number of limitations, given the rarity of these tumors as assessed by historical controls and the
25 repeatability of this result across studies and strains, these are considered biologically significant.
26 Therefore, while individual studies provide only suggestive evidence of renal carcinogenicity,
27 the database as a whole supports the conclusion that TCE is a kidney carcinogen in rats, with
28 males being more sensitive than females. No other tested laboratory species (i.e., mice and
29 hamsters) have exhibited increased kidney tumors, with no adequate explanation for these
30 species differences (particularly with mice, which have been extensively tested). With respect to
31 the liver, TCE and its oxidative metabolites chloral hydrate (CH), TCA, and DCA are clearly
32 carcinogenic in mice, with strain and sex differences in potency that appear to parallel,
33 qualitatively, differences in background tumor incidence. Data in other laboratory animal
34 species are limited; thus, except for DCA which is carcinogenic in rats, inadequate evidence
35 exists to evaluate the hepatocarcinogenicity of these compounds in rats or hamsters. However, to

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1 the extent that there is hepatocarcinogenic potential in rats, TCE is clearly less potent in the
2 strains tested in this species than in B6C3F1 and Swiss mice. See Section 4.5.5 for additional
3 discussion of laboratory animal data on TCE-induced liver tumors. Additionally, there is more
4 limited evidence for TCE-induced lymphatic cancers in rats and mice, lung tumors in mice, and
5 testicular tumors in rats. With respect to the lymphatic cancers, two studies in mice reported
6 increased incidences of lymphomas in females of two different strains, and two studies in rats
7 reported leukemias in males of one strain and females of another. However, these tumors had
8 relatively modest increases in incidence with treatment, and were not reported to be increased in
9 other studies. See Section 4.6.2.4 for additional discussion of laboratory animal data on TCE-
10 induced lymphatic tumors. With respect to lung tumors, rodent bioassays have demonstrated a
11 statistically significant increase in pulmonary tumors in mice following chronic inhalation
12 exposure to TCE, and nonstatistically significant increases in mice exposed orally; but
13 pulmonary tumors were not reported in other species tested (i.e., rats and hamsters) (see
14 Section 4.7.2.2). Finally, increased testicular (interstitial or Leydig cell) tumors have been
15 observed in multiple studies of rats exposed by inhalation and gavage, although in some cases
16 high (>75%) control rates of testicular tumors in rats limited the ability to detect a treatment
17 effect. See Section 4.8.2.2 for additional discussion of laboratory animal data on TCE-induced
18 tumors of the reproductive system. Overall, TCE is clearly carcinogenic in rats and mice. The
19 apparent lack of site concordance across laboratory animal studies may be due to limitations in
20 design or conduct in a number of rat bioassays and/or genuine interspecies differences in
21 qualitative or quantitative sensitivity (i.e., potency). Nonetheless, these studies have shown
22 carcinogenic effects across different strains, sexes, and routes of exposure, and site-concordance
23 is not necessarily expected for carcinogens.

24 A second line of supporting evidence for TCE carcinogenicity in humans consists of
25 toxicokinetic data indicating that TCE is well absorbed by all routes of exposure, and that TCE
26 absorption, distribution, metabolism, and excretion are qualitatively similar in humans and
27 rodents. As summarized above, there is evidence that TCE is systemically available, distributes
28 to organs and tissues, and undergoes systemic metabolism from all routes of exposure.
29 Therefore, although the strongest evidence from epidemiologic studies largely involves
30 inhalation exposures, the evidence supports TCE carcinogenicity being applicable to all routes of
31 exposure. In addition, there is no evidence of major qualitative differences across species in
32 TCE absorption, distribution, metabolism, and excretion. Extensive *in vivo* and *in vitro* data
33 show that mice, rats, and humans all metabolize TCE via two primary pathways: oxidation by
34 CYPs and conjugation with glutathione via GSTs. Several metabolites and excretion products
35 from both pathways have been detected in blood and urine from exposed humans as well as

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1 from at least one rodent species. In addition, the subsequent distribution, metabolism, and
2 excretion of TCE metabolites are qualitatively similar among species. Therefore, humans
3 possess the metabolic pathways that produce the TCE metabolites thought to be involved in the
4 induction of rat kidney and mouse liver tumors, and internal target tissues of both humans and
5 rodents experience a similar mix of TCE and metabolites. See Sections 3.1–3.4 for additional
6 discussion of TCE toxicokinetics. Quantitative interspecies differences in toxicokinetics do
7 exist, and are addressed through PBPK modeling (see Section 3.5 and Appendix A).
8 Importantly, these quantitative differences affect only interspecies extrapolations of carcinogenic
9 potency, and do not affect inferences as to the carcinogenic hazard for TCE.

10 Finally, available mechanistic data do not suggest a lack of human carcinogenic hazard
11 from TCE exposure. In particular, these data do not suggest qualitative differences between
12 humans and test animals that would preclude any of the hypothesized key events in the
13 carcinogenic MOA in rodents from occurring in humans. For the kidney, the predominance of
14 positive genotoxicity data in the database of available studies of TCE metabolites derived from
15 GSH conjugation (in particular DCVC), together with toxicokinetic data consistent with their
16 systemic delivery to and *in situ* formation in the kidney, supports the conclusion that a mutagenic
17 MOA is operative in TCE-induced kidney tumors. While supporting the biological plausibility
18 of this hypothesized MOA, available data on the von Hippel-Lindau (VHL) gene in humans or
19 transgenic animals do not conclusively elucidate the role of VHL mutation in TCE-induced renal
20 carcinogenesis. Cytotoxicity and compensatory cell proliferation, similarly presumed to be
21 mediated through metabolites formed after GSH-conjugation of TCE, have also been suggested
22 to play a role in the MOA for renal carcinogenesis, as high incidences of nephrotoxicity have
23 been observed in animals at doses that induce kidney tumors. Human studies have reported
24 markers for nephrotoxicity at current occupational exposures, although data are lacking at lower
25 exposures. Nephrotoxicity is observed in both mice and rats, in some cases with nearly 100%
26 incidence in all dose groups, but kidney tumors are only observed at low incidences in rats at the
27 highest tested doses. Therefore, nephrotoxicity alone appears to be insufficient, or at least not
28 rate-limiting, for rodent renal carcinogenesis, since maximal levels of toxicity are reached before
29 the onset of tumors. In addition, nephrotoxicity has not been shown to be necessary for kidney
30 tumor induction by TCE in rodents. In particular, there is a lack of experimental support for
31 causal links, such as compensatory cellular proliferation or clonal expansion of initiated cells,
32 between nephrotoxicity and kidney tumors induced by TCE. Furthermore, it is not clear if
33 nephrotoxicity is one of several key events in a MOA, if it is a marker for an “upstream” key
34 event (such as oxidative stress) that may contribute independently to both nephrotoxicity and
35 renal carcinogenesis, or if it is incidental to kidney tumor induction. Moreover, while

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1 toxicokinetic differences in the GSH conjugation pathway along with their uncertainty are
2 addressed through PBPK modeling, no data suggest that any of the proposed key events for
3 TCE-induced kidney tumors in rats are precluded in humans. See Section 4.4.7 for additional
4 discussion of the MOA for TCE-induced kidney tumors. Therefore, TCE-induced rat kidney
5 tumors provide additional support for the convincing human evidence of TCE-induced kidney
6 cancer, with mechanistic data supportive of a mutagenic MOA.

7 With respect to other tumor sites, data are insufficient to conclude that any of the other
8 hypothesized MOAs are operant. In the liver, a mutagenic MOA mediated by CH, which has
9 evidence for genotoxic effects, or some other oxidative metabolite of TCE cannot be ruled out,
10 but data are insufficient to conclude it is operant. A second MOA hypothesis for TCE-induced
11 liver tumors involves activation of the peroxisome proliferator activated receptor alpha (PPAR α)
12 receptor. Clearly, *in vivo* administration of TCE leads to activation of PPAR α in rodents and
13 likely does so in humans as well. However, the evidence as a whole does not support the view
14 that PPAR α is the sole operant MOA mediating TCE hepatocarcinogenesis. Rather, there is
15 evidential support for multiple TCE metabolites and multiple toxicity pathways contributing to
16 TCE-induced liver tumors. Furthermore, recent experiments have demonstrated that PPAR α
17 activation and the sequence of key events in the hypothesized MOA are not sufficient to induce
18 hepatocarcinogenesis (Yang et al., 2007). Moreover, the demonstration that the PPAR α agonist
19 di(2-ethylhexyl) phthalate induces tumors in PPAR α -null mice supports the view that the events
20 comprising the hypothesized PPAR α activation MOA are not necessary for liver tumor induction
21 in mice by this PPAR α agonist (Ito et al., 2007). See Section 4.5.7 for additional discussion of
22 the MOA for TCE-induced liver tumors. For mouse lung tumors, as with the liver, a mutagenic
23 MOA involving CH has also been hypothesized, but there are insufficient data to conclude that it
24 is operant. A second MOA hypothesis for mouse lung tumors has been posited involving other
25 effects of oxidative metabolites including cytotoxicity and regenerative cell proliferation, but
26 experimental support remains limited, with no data on proposed key events in experiments of
27 duration 2 weeks or longer. See Section 4.7.4 for additional discussion of the MOA for TCE-
28 induced lung tumors. A MOA subsequent to *in situ* oxidative metabolism, whether involving
29 mutagenicity, cytotoxicity, or other key events, may also be relevant to other tissues where TCE
30 would undergo CYP metabolism. For instance, CYP2E1, oxidative metabolites, and protein
31 adducts have been reported in the testes of rats exposed to TCE, and, in some rat bioassays, TCE
32 exposure increased the incidence of rat testicular tumors. However, inadequate data exist to
33 adequately define a MOA hypothesis for this tumor site (see Section 4.8.2.3 for additional
34 discussion of the MOA for TCE-induced testicular tumors).

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1 **6.1.5. Susceptibility (see Sections 4.10 and 4.11.3)**

2 There is some evidence that certain populations may be more susceptible to exposure to
3 TCE. Factors affecting susceptibility examined include lifestage, gender, genetic
4 polymorphisms, race/ethnicity, preexisting health status, and lifestyle factors and nutrition status.
5 Factors that impact early lifestage susceptibility include exposures such as transplacental transfer
6 and breast milk ingestion, early lifestage-specific toxicokinetics, and differential outcomes in
7 early lifestages such as developmental cardiac defects (see Section 4.10.1). Because the weight
8 of evidence supports a mutagenic MOA being operative for TCE carcinogenicity in the kidney
9 (see Section 4.4.7), and there is an absence of chemical-specific data to evaluate differences in
10 carcinogenic susceptibility, early-life susceptibility should be assumed and the age-dependent
11 adjustment factors (ADAFs) should be applied, in accordance with the Supplemental Guidance
12 (see summary below in Section 6.2.2.5). Fewer data are available on later lifestages, although
13 there is suggestive evidence to indicate that older adults may experience increased adverse
14 effects than younger adults due to greater tissue distribution of TCE. In general, more studies
15 specifically designed to evaluate effects in early and later lifestages are needed in order to more
16 fully characterize potential life stage-related TCE toxicity. Gender-specific (see
17 Section 4.10.2.1) differences also exist in toxicokinetics (e.g., cardiac outputs, percent body fat,
18 expression of metabolizing enzymes) and susceptibility to toxic endpoints (e.g., gender-specific
19 effects on the reproductive system, gender differences in baseline risks to endpoints such as
20 scleroderma or liver cancer). Genetic variation (see Section 4.10.2.2) likely has an effect on the
21 toxicokinetics of TCE. Increased CYP2E1 activity and GST polymorphisms may influence
22 susceptibility of TCE due to effects on production of toxic metabolites or may play a role in
23 variability in toxic response. Differences in genetic polymorphisms related to the metabolism of
24 TCE have also been observed among various race/ethnic groups (see Section 4.10.2.3).
25 Preexisting diminished health status (see Section 4.10.2.4) may alter the response to TCE
26 exposure. Individuals with increased body mass may have an altered toxicokinetic response due
27 to the increased uptake of TCE into fat. Other conditions that may alter the response to TCE
28 exposure include diabetes and hypertension, and lifestyle and nutrition factors (see
29 Section 4.10.2.5) such alcohol consumption, tobacco smoking, nutritional status, physical
30 activity, and socioeconomic status. Alcohol intake has been associated with inhibition of TCE
31 metabolism in both humans and experimental animals. In addition, such conditions have been
32 associated with increased baseline risks for health effects also associated with TCE, such as
33 kidney cancer and liver cancer. However, the interaction between TCE and known risk factors
34 for human diseases is not known, and further evaluation of the effects due to these factors is
35 needed.

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1 In sum, there is some evidence that certain populations may be more susceptible to
2 exposure to TCE. Factors affecting susceptibility examined include lifestage, gender, genetic
3 polymorphisms, race/ethnicity, preexisting health status, and lifestyle factors and nutrition status.
4 However, except in the case of toxicokinetic variability characterized using the PBPK model
5 described in Section 3.5, there are inadequate chemical-specific data to quantify the degree of
6 differential susceptibility due to such factors.

8 **6.2. DOSE-RESPONSE ASSESSMENT**

9 This section summarizes the major conclusions of the dose-response analysis for TCE
10 noncancer effects and carcinogenicity, with more detailed discussions in Chapter 5.

12 **6.2.1. Noncancer Effects (see Section 5.1)**

13 **6.2.1.1. Background and Methods**

14 As summarized above, based on the available human epidemiologic data and
15 experimental and mechanistic studies, it is concluded that TCE poses a potential human health
16 hazard for noncancer toxicity to the central nervous system, the kidney, the liver, the immune
17 system, the male reproductive system, and the developing fetus. The evidence is more limited
18 for TCE toxicity to the respiratory tract and female reproductive system.

19 Dose-response analysis for a noncancer endpoint generally involves two steps: (1) the
20 determination of a point of departure (POD) derived from a benchmark dose (BMD)¹, a
21 no-observed-adverse-effect level (NOAEL), or a lowest-observed-adverse-effect level (LOAEL);
22 and (2) adjustment of the POD by endpoint/study-specific “uncertainty factors” (UFs),
23 accounting for adjustments and uncertainties in the extrapolation from the study conditions to
24 conditions of human exposure.

25 Because of the large number of noncancer health effects associated with TCE exposure
26 and the large number of studies reporting on these effects, in contrast to toxicological reviews for
27 chemicals with smaller databases of studies, a formal, quantitative screening process (see
28 Section 5.1) was used to reduce the number of endpoints and studies to those that would best
29 inform the selection of the *critical effects* for the inhalation reference concentration (RfC) and
30 oral reference dose (RfD).² As described in Section 5.1, for all studies described in Chapter 4

¹ more precisely, it is the benchmark dose lower bound (BMDL), i.e., the (one-sided) 95% lower confidence bound on the dose corresponding to the benchmark response (BMR) for the effect, that is used as the POD

² In U.S. EPA noncancer health assessments, the RfC [RfD] is an estimate (with uncertainty spanning perhaps an order of magnitude) of a continuous inhalation [daily oral] exposure to the human population (including sensitive subgroups) that is likely to be without an appreciable risk of deleterious effects during a lifetime. It can be derived from a NOAEL, LOAEL, or benchmark concentration [dose], with uncertainty factors generally applied to reflect limitations of the data used.

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1 which report adverse noncancer health effects and provided quantitative dose-response data,
2 PODs on the basis of applied dose, adjusted by endpoint/study-specific UFs, were used to
3 develop candidate RfCs (cRfCs) and candidate RfDs (cRfDs) intended to be protective for each
4 endpoint individually. Candidate critical effects—those with the lowest cRfCs and cRfDs taking
5 into account the confidence in each estimate—were selected within each of the following health
6 effect domains: (1) neurological, (2) systemic/organ system; (3) immunological; (4)
7 reproductive; and (5) developmental. For each of these candidate critical effects, the PBPK
8 model developed in Section 3.5 was used for interspecies, intraspecies, and route-to-route
9 extrapolation on the basis of internal dose to develop PBPK model-based PODs. Plausible
10 internal dose metrics were selected based on what is understood about the role of different TCE
11 metabolites in toxicity and the MOA for toxicity. These PODs were then adjusted by
12 endpoint/study-specific UFs, taking into account the use of the PBPK model, to develop PBPK
13 model-based candidate RfCs (p-cRfCs) and candidate RfDs (p-cRfDs). The most sensitive
14 cRfCs, p-cRfCs, cRfDs, and p-cRfDs were then evaluated, taking into account the confidence in
15 each estimate, to arrive at overall candidate RfCs and RfDs for each health effect type. Then, the
16 RfC and RfD for TCE were selected so as to be protective of the most sensitive effects. In
17 contrast to the approach used in most assessments, in which the RfC and RfD are each based on
18 a single critical effect, the final RfC and RfD for TCE were based on multiple critical effects that
19 resulted in very similar candidate RfC and RfD values at the low end of the full range of values.
20 This approach was taken here because it provides robust estimates of the RfC and RfD and
21 because it highlights the multiple effects that are all yielding very similar candidate values.
22

23 **6.2.1.2. *Uncertainties and Application of Uncertainty Factors (UFs) (see Section 5.1.1 and*** 24 ***5.1.4)***

25 An underlying assumption in deriving reference values for noncancer effects is that the
26 dose-response relationship for these effects has a threshold. Thus, a fundamental uncertainty is
27 the validity of that assumption. For some effects, in particular effects on very sensitive processes
28 (e.g., developmental processes) or effects for which there is a nontrivial background level and
29 even small exposures may contribute to background disease processes in more susceptible
30 people, a practical threshold (i.e., a threshold within the range of environmental exposure levels
31 of regulatory concern) may not exist.

32 Nonetheless, under the assumption of a threshold, the desired exposure level to have as a
33 reference value is the maximum level at which there is no appreciable risk for an adverse effect
34 in sensitive subgroups (of humans). However, because it is not possible to know what this level
35 is, “uncertainty factors” are used to attempt to address quantitatively various aspects, depending
36 on the data set, of qualitative uncertainty.

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1 First there is uncertainty about the “point of departure” for the application of UFs.
2 Conceptually, the POD should represent the maximum exposure level at which there is no
3 appreciable risk for an adverse effect in the study population under study conditions (i.e., the
4 threshold in the dose-response relationship). Then, the application of the relevant UFs is
5 intended to convey that exposure level to the corresponding exposure level for sensitive human
6 subgroups exposed continuously for a lifetime. In fact, it is again not possible to know that
7 exposure level even for a laboratory study because of experimental limitations (e.g. the power to
8 detect an effect, dose spacing, measurement errors, etc.), and crude approximations like the
9 NOAEL or a BMDL are used. If a LOAEL is used as the POD, the LOAEL-to-NOAEL UF is
10 applied as an adjustment factor to better approximate the desired exposure level (threshold),
11 although the necessary extent of adjustment is unknown. The standard value for the LOAEL-to-
12 NOAEL UF is 10, although sometimes a value of 3 is used if the effect is considered minimally
13 adverse at the response level observed at the LOAEL or even 1 if the effect is an early marker for
14 an adverse effect. For one POD in this assessment, a value of 30 was used for the LOAEL-to-
15 NOAEL UF because the incidence rate for the adverse effect was $\geq 90\%$ at the LOAEL.

16 If a BMDL is used as the POD, there are uncertainties regarding the appropriate dose-
17 response model to apply to the data, but these should be minimal if the modeling is in the
18 observable range of the data. There are also uncertainties about what BMR to use to best
19 approximate the desired exposure level (threshold, see above). For continuous endpoints, in
20 particular, it is often difficult to identify the level of change that constitutes the “cut-point” for an
21 adverse effect. Sometimes, to better approximate the desired exposure level, a BMR somewhat
22 below the observable range of the data is selected. In such cases, the model uncertainty is
23 increased, but this is a trade-off to reduce the uncertainty about the POD not being a good
24 approximation for the desired exposure level.

25 For each of these types of PODs, there are additional uncertainties pertaining to
26 adjustments to the administered exposures (doses). Typically, administered exposures (doses)
27 are converted to equivalent continuous exposures (daily doses) over the study exposure period
28 under the assumption that the effects are related to concentration \times time, independent of the daily
29 (or weekly) exposure regimen (i.e., a daily exposure of 6 hours to 4 ppm is considered equivalent
30 to 24 hours of exposure to 1 ppm). However, the validity of this assumption is generally
31 unknown, and, if there are dose-rate effects, the assumption of $C \times t$ equivalence would tend to
32 bias the POD downwards. Where there is evidence that administered exposure better correlates
33 to the effect than equivalent continuous exposure averaged over the study exposure period (e.g.,
34 visual effects), administered exposure was not adjusted. For the PBPK analyses in this
35 assessment, the actual administered exposures are taken into account in the PBPK modeling, and

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1 equivalent daily values (averaged over the study exposure period) for the dose metrics are
2 obtained (see above, 5.1.3.2). Additional uncertainties about the PBPK based estimates include
3 uncertainties about the appropriate dose metric for each effect, although for some effects there
4 was better information about relevant dose metrics than for others (see Section 5.1.3.1).

5 There is also uncertainty about the other UFs. The human variability UF is to some
6 extent an adjustment factor because for more sensitive people, the dose-response relationship
7 shifts to lower exposures. But there is uncertainty about the extent of the adjustment required,
8 i.e., about the distribution of human susceptibility. Therefore, in the absence of data on a
9 susceptible population(s) or on the distribution of susceptibility in the general population, an UF
10 of 10 is generally used, which breaks down (approximately) to a factor of 3 for pharmacokinetic
11 variability and a factor of 3 for pharmacodynamic variability. This standard value was used for
12 all the PODs based on applied dose in this assessment with the exception of the PODs for a few
13 immunological effects that were based on data from a sensitive (autoimmune-prone) mouse
14 strain. For those PODs, an UF of 3 (reflecting pharmacokinetics only) was used for human
15 variability. The PBPK analyses in this assessment attempt to account for the pharmacokinetic
16 portion of human variability using human data on pharmacokinetic variability. For PBPK
17 model-based candidate reference values, the pharmacokinetic component of this UF was omitted.
18 A quantitative uncertainty analysis of the PBPK derived dose metrics used in the assessment is
19 presented in Section 5.1.4.2 in Chapter 5. There is still uncertainty regarding the susceptible
20 subgroups for TCE exposure and the extent of pharmacodynamic variability.

21 If the data used to determine a particular POD are from laboratory animals, an
22 interspecies extrapolation UF is used. This UF is also to some extent an adjustment factor for the
23 expected scaling for toxicologically-equivalent doses across species (i.e., according to body
24 weight to the $3/4$ power for oral exposures). However, there is also uncertainty about the true
25 extent of interspecies differences for specific noncancer effects from specific chemical
26 exposures. For oral exposures, the standard value for the interspecies UF is 10, which can be
27 viewed as breaking down (approximately) to a factor of 3 for the “adjustment” (nominally
28 pharmacokinetics) and a factor of 3 for the “uncertainty” (nominally pharmacodynamics). For
29 inhalation exposures for systemic toxicants such as TCE, no adjustment across species is
30 generally assumed for fixed air concentrations (ppm equivalence), and the standard value for the
31 interspecies UF is 3 reflects “uncertainty” (nominally pharmacodynamics only). The PBPK
32 analyses in this assessment attempt to account for the “adjustment” portion of interspecies
33 extrapolation using rodent pharmacokinetic data to estimate internal doses for various dose
34 metrics. Equal doses of these dose metrics, appropriately scaled, are then assumed to convey
35 equivalent risk across species. For PBPK model-based candidate reference values, the

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1 “adjustment” component of this UF was omitted. With respect to the “uncertainty” component,
2 quantitative uncertainty analyses of the PBPK derived dose metrics used in the assessment are
3 presented in Section 5.1.4.2 in Chapter 5. However, these only address the pharmacokinetic
4 uncertainties in a particular dose metric, and there is still uncertainty regarding the true dose
5 metrics. Nor do the PBPK analyses address the uncertainty in either cross-species
6 pharmacodynamic differences (i.e., about the assumption that equal doses of the appropriate dose
7 metric convey equivalent risk across species for a particular endpoint from a specific chemical
8 exposure) or in cross-species pharmacokinetic differences not accounted for by the PBPK model
9 dose metrics (e.g., departures from the assumed interspecies scaling of clearance of the active
10 moiety, in the cases where only its production is estimated). A value of 3 is typically used for
11 the “uncertainty” about cross-species differences, and this generally represents true uncertainty
12 because it is usually unknown, even after adjustments have been made to account for the
13 expected interspecies differences, whether humans have more or less susceptibility, and to what
14 degree, than the laboratory species in question.

15 RfCs and RfDs apply to lifetime exposure, but sometimes the best (or only) available
16 data come from less-than-lifetime studies. Lifetime exposure can induce effects that may not be
17 apparent or as large in magnitude in a shorter study; consequently, a dose that elicits a specific
18 level of response from a lifetime exposure may be less than the dose eliciting the same level of
19 response from a shorter exposure period. If the effect becomes more severe with increasing
20 exposure, then chronic exposure would shift the dose-response relationship to lower exposures,
21 although the true extent of the shift is unknown. PODs based on subchronic exposure data are
22 generally divided by a subchronic-to-chronic UF, which has a standard value of 10. If there is
23 evidence suggesting that exposure for longer time periods does not increase the magnitude of an
24 effect, a lower value of 3 or 1 might be used. For some reproductive and developmental effects,
25 chronic exposure is that which covers a specific window of exposure that is relevant for eliciting
26 the effect, and subchronic exposure would correspond to an exposure that is notably less than the
27 full window of exposure.

28 Sometimes a database UF is also applied to address limitations or uncertainties in the
29 database. The overall database for TCE is quite extensive, with studies for many different types
30 of effects, including 2-generation reproductive studies, as well as neurological and
31 immunological studies. In addition, there were sufficient data to develop a reliable PBPK model
32 to estimate route-to-route extrapolated doses for some candidate critical effects for which data
33 were only available for one route of exposure. Thus, there is a high degree of confidence that the
34 TCE database was sufficient to identify some sensitive endpoints, and no database UF was used
35 in this assessment.

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1 **6.2.1.3. Candidate Critical Effects and Reference Values (see Sections 5.1.2 and 5.1.3)**

2 A large number of endpoints and studies were considered within each health effect
3 domain. Chapter 5 contains a comprehensive discussion of all endpoints/studies which were
4 considered for developing candidate reference values (cRfCs, cRfDs, p-cRfCs, and p-cRfDs),
5 their PODs, and the UFs applied. The summary below reviews the selection of candidate critical
6 effects for each health effect domain, the confidence in the reference values, the selection of
7 PBPK model-based dose metrics, and the impact of PBPK modeling on the candidate reference
8 values.

9
10 **6.2.1.3.1. Neurological effects.** Candidate reference values were developed for several
11 neurological domains for which there was evidence of hazard (see Tables 5-1 and 5-8). There is
12 higher confidence in the candidate reference values for trigeminal nerve, auditory, or
13 psychomotor effects, but the available data suggest that the more sensitive indicators of TCE
14 neurotoxicity are changes in wakefulness, regeneration of the sciatic nerve, demyelination in the
15 hippocampus and degeneration of dopaminergic neurons. Therefore, these more sensitive effects
16 are considered the candidate critical effects for neurotoxicity, albeit with more uncertainty in the
17 corresponding candidate reference values. Of these more sensitive effects, there is greater
18 confidence in the changes in wakefulness reported by Arito et al. (1994). In addition, trigeminal
19 nerve effects are considered a candidate critical effect because this is the only type of
20 neurological effect for which human data are available, and the POD for this effect is similar to
21 that from the most sensitive rodent study (Arito et al., 1994, for changes in wakefulness).
22 Between the two human studies of trigeminal nerve effects, Ruitjen et al. (1991) is preferred for
23 deriving noncancer reference values because its exposure characterization is considered more
24 reliable.

25 Because of the lack of specific data as to the metabolites involved and the MOA for the
26 candidate critical neurologic effects, PBPK model predictions of total metabolism (scaled by
27 body weight to the $\frac{3}{4}$ power) were selected as the preferred dose metric based on the general
28 observation that TCE toxicity is associated with metabolism. The area-under-the-curve (AUC)
29 of TCE in blood was used as an alternative dose metric. With these dose metrics, the candidate
30 reference values derived using the PBPK model were only modestly (~3-fold or less) different
31 than those derived on the basis of applied dose.

32
33 **6.2.1.3.2. Kidney effects.** High-confidence candidate reference values were developed for
34 histopathological and weight changes in the kidney (see Tables 5-2 and 5-9), and these are
35 considered to be candidate critical effects for several reasons. First, they appear to be the most

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1 sensitive indicators of toxicity that are available for the kidney. In addition, as discussed in
2 Sections 3.3 and 3.5, both *in vitro* and *in vivo* pharmacokinetic data indicate substantially more
3 production of GSH-conjugates thought to mediate TCE kidney effects in humans relative to rats
4 and mice. Several studies are considered reliable for developing candidate reference values for
5 these endpoints. For histopathological changes, these were the only available inhalation study
6 (Maltoni et al., 1986), the NTP (1988) study in rats, and the National Cancer Institute (NCI,
7 1976) study in mice. For kidney weight changes, both available studies (Kjellstrand et al.,
8 1983b; Woolhiser et al., 2006) were chosen as candidate critical studies.

9 Due to the substantial evidence supporting the role of GSH conjugation metabolites in
10 TCE-induced nephrotoxicity, the preferred PBPK model dose metrics for kidney effects were the
11 amount of DCVC bioactivated in the kidney for rat studies and the amount of GSH conjugation
12 (both scaled by body weight to the $\frac{3}{4}$ power) for mouse studies (inadequate toxicokinetic data are
13 available in mice for predicting the amount of DCVC bioactivation). With these dose metrics,
14 the candidate reference values derived using the PBPK model were 300- to 400-fold lower than
15 those derived on the basis of applied dose. As discussed above and in Chapter 3, this is due to
16 the available *in vivo* and *in vitro* data supporting not only substantially more GSH conjugation in
17 humans than in rodents, but also substantial interindividual toxicokinetic variability.

18
19 **6.2.1.3.3. Liver effects.** Hepatomegaly appears to be the most sensitive indicator of toxicity that
20 is available for the liver and is therefore, considered a candidate critical effect. Several studies
21 are considered reliable for developing high confidence candidate reference values for this
22 endpoint. Since they all indicated similar sensitivity but represented different species and/or
23 routes of exposure, they were all considered candidate critical studies (see Tables 5-2 and 5-9).

24 Due to the substantial evidence supporting the role of oxidative metabolism in TCE-
25 induced hepatomegaly (and evidence against TCA being the sole mediator of TCE-induced
26 hepatomegaly [Evans et al., 2009]), the preferred PBPK model dose metric for liver effects was
27 the amount of hepatic oxidative metabolism (scaled by body weight to the $\frac{3}{4}$ power). Total
28 (hepatic and extrahepatic) oxidative metabolism (scaled by body weight to the $\frac{3}{4}$ power) was
29 used as an alternative dose metric. With these dose metrics, the candidate reference values
30 derived using the PBPK model were only modestly (~3-fold or less) different than those derived
31 on the basis of applied dose.

32
33 **6.2.1.3.4. Immunological effects.** There is high qualitative confidence for TCE immunotoxicity
34 and moderate confidence in the candidate reference values that can be derived from the available
35 studies (see Tables 5-3 and 5-11). Decreased thymus weight reported at relatively low exposures

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1 in nonautoimmune-prone mice is a clear indicator of immunotoxicity (Keil et al., 2009), and is
2 therefore, considered a candidate critical effect. A number of studies have also reported changes
3 in markers of immunotoxicity at relatively low exposures. Among markers for autoimmune
4 effects, the more sensitive measures of autoimmune changes in liver and spleen (Kaneko et al.,
5 2000) and increased anti-dsDNA and anti-ssDNA antibodies (early markers for systemic lupus
6 erythematosus) (Keil et al., 2009) are considered the candidate critical effects. For markers of
7 immunosuppression, the more sensitive measures of decreased PFC response (Woolhiser et al.,
8 2006), decreased stem cell bone marrow recolonization, and decreased cell-mediated response to
9 sRBC (both from Sanders et al., 1982) are considered the candidate critical effects.

10 Developmental immunological effects are discussed below as part of the summary of
11 developmental effects (see Section 6.2.1.3.6).

12 Because of the lack of specific data as to the metabolites involved and the MOA for the
13 candidate critical immunologic effects, PBPK model predictions of total metabolism (scaled by
14 body weight to the $\frac{3}{4}$ power) was selected as the preferred dose metric based on the general
15 observation that TCE toxicity is associated with metabolism. The AUC of TCE in blood was
16 used as an alternative dose metric. With these dose metrics, the candidate reference values
17 derived using the PBPK model were, with one exception, only modestly (~3-fold or less)
18 different than those derived on the basis of applied dose. For the Woolhiser et al. (2006)
19 decreased PFC response, with the alternative dose metric of AUC of TCE in blood, BMD
20 modeling based on internal doses changed the candidate reference value by 17-fold higher than
21 the cRfC based on applied dose. However, the dose-response model fit for this effect using this
22 metric was substantially worse than the fit using the preferred metric of total oxidative
23 metabolism, with which the change in candidate reference value was only 1.3-fold.

24
25 **6.2.1.3.5. Reproductive effects.** While there is high qualitative confidence in the male
26 reproductive hazard posed by TCE, there is lower confidence in the reference values that can be
27 derived from the available studies of these effects (see Tables 5-4 and 5-12). Relatively high
28 PODs are derived from several studies reporting less sensitive endpoints (George et al., 1985,
29 1986; Land et al., 1981), and correspondingly higher cRfCs and cRfDs suggest that they are not
30 likely to be critical effects. The studies reporting more sensitive endpoints also tend to have
31 greater uncertainty. For the human study by Chia et al. (1996), there are uncertainties in the
32 characterization of exposure and the adversity of the effect measured in the study. For the
33 Kumar et al. (2000a, b, 2001), Forkert et al. (2002) and Kan et al. (2007) studies, the severity of
34 the sperm and testes effects appears to be continuing to increase with duration even at the end of
35 the study, so it is plausible that a lower exposure for a longer duration may elicit similar effects.

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1 For the DuTeaux et al. (2004b) study, there is also duration- and low-dose extrapolation
2 uncertainty due to the short duration of the study in comparison to the time period for sperm
3 development as well as the lack of a NOAEL at the tested doses. Overall, even though there are
4 limitations in the quantitative assessment, there remains sufficient evidence to consider these to
5 be candidate critical effects.

6 There is moderate confidence both in the hazard and the candidate reference values for
7 reproductive effects other than male reproductive effects. While there are multiple studies
8 suggesting decreased maternal body weight with TCE exposure, this systemic change may not be
9 indicative of more sensitive reproductive effects. None of the estimates developed from other
10 reproductive effects is particularly uncertain or unreliable. Therefore, delayed parturition
11 (Narotsky et al., 1995) and decreased mating (George et al., 1986), which yielded the lowest
12 cRfDs, were considered candidate critical effects. These effects were also included so that
13 candidate critical reproductive effects from oral studies would not include only that reported by
14 DuTeaux et al. (2004b), from which deriving the cRfD entailed a higher degree of uncertainty.

15 Because of the general lack of specific data as to the metabolites involved and the MOA
16 for the candidate critical reproductive effects, PBPK model predictions of total metabolism
17 (scaled by body weight to the $\frac{3}{4}$ power) was selected as the preferred dose metric based on the
18 general observation that TCE toxicity is associated with metabolism. The AUC of TCE in blood
19 was used as an alternative dose metric. The only exception to this was for the DuTeaux et al.
20 (2004) study, which suggested that local oxidative metabolism of TCE in the male reproductive
21 tract was involved in the effects reported. Therefore, in this case, AUC of TCE in blood was
22 considered the preferred dose metric, while total oxidative metabolism (scaled by body weight to
23 the $\frac{3}{4}$ power) was considered the alternative metric. With these dose metrics, the candidate
24 reference values derived using the PBPK model were only modestly (~ 3.5 -fold or less) different
25 than those derived on the basis of applied dose.

26
27 **6.2.1.3.6. Developmental effects.** There is moderate-to-high confidence both in the hazard and
28 the candidate reference values for developmental effects of TCE (see Tables 5-5 and 5-13). It is
29 also noteworthy that the PODs for the more sensitive developmental effects were similar to or, in
30 most cases, lower than the PODs for the more sensitive reproductive effects, suggesting that
31 developmental effects are not a result of paternal or maternal toxicity. Among inhalation studies,
32 candidate reference values were only developed for effects in rats reported in Healy et al. (1982),
33 of resorptions, decreased fetal weight, and delayed skeletal ossification. These were all
34 considered candidate critical developmental effects. Because resorptions were also reported in
35 oral studies, the most sensitive (rat) oral study for this effect (and most reliable for dose-response

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1 analysis) of Narotsky et al. (1995) was also selected as a candidate critical study. The
2 confidence in the oral studies and candidate reference values developed for more sensitive
3 endpoints is more moderate, but still sufficient for consideration as candidate critical effects.
4 The most sensitive endpoints by far are the increased fetal heart malformations in rats reported
5 by Johnson et al. (2003) and the developmental immunotoxicity in mice reported by Peden-
6 Adams et al. (2006), and these are both considered candidate critical effects.
7 Neurodevelopmental effects are a distinct type among developmental effects. Thus, the next
8 most sensitive endpoints of decreased rearing postexposure in mice (Fredricksson et al., 1993),
9 increased exploration postexposure in rats (Taylor et al., 1985) and decreased myelination in the
10 hippocampus of rats (Isaacson and Taylor, 1989) are also considered candidate critical effects.

11 Because of the general lack of specific data as to the metabolites involved and the MOA
12 for the candidate critical reproductive effects, PBPK model predictions of total metabolism
13 (scaled by body weight to the $3/4$ power) was selected as the preferred dose metric based on the
14 general observation that TCE toxicity is associated with metabolism. The AUC of TCE in blood
15 was used as an alternative dose metric. The only exception to this was for the Johnson et al.
16 (2003) study, which suggested that oxidative metabolites were involved in the effects reported
17 based on similar effects being reported from TCA and DCA exposure. Therefore, in this case,
18 total oxidative metabolism (scaled by body weight to the $3/4$ power) was considered the preferred
19 dose metric, while AUC of TCE in blood was considered the alternative metric. With these dose
20 metrics, the candidate reference values derived using the PBPK model were, with one exception,
21 only modestly (~3-fold or less) different than those derived on the basis of applied dose. For
22 resorptions reported by Narotsky et al. (1995), BMD modeling based on internal doses changed
23 the candidate reference value by 7- to 8-fold larger than the corresponding cRfD based on
24 applied dose. However, there is substantial uncertainty in the low-dose curvature of the dose-
25 response curve for modeling both with applied and internal dose, so the BMD remains somewhat
26 uncertain for this endpoint/study. Finally, for two studies (Isaacson and Taylor, 1989; Peden-
27 Adams et al., 2006), PBPK modeling of internal doses was not performed due to the inability to
28 model the complicated exposure pattern (*in utero*, followed by lactational transfer, followed by
29 drinking water postweaning).

30
31 **6.2.1.3.7. Summary of most sensitive candidate reference values.** As shown in Section 5.1.3
32 and 5.1.5, the most sensitive candidate reference values are for developmental effects of heart
33 malformations in rats (candidate RfC of 0.0004 ppm and candidate RfD of 0.0005 mg/kg/d),
34 developmental immunotoxicity in mice exposed pre- and postnatally (candidate RfD of
35 0.0004 mg/kg/d), immunological effects in mice (lowest candidate RfCs of 0.0003–0.003 ppm

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1 and lowest candidate RfDs of 0.0005–0.005 mg/kg/d), and kidney effects in rats and mice
2 (candidate RfCs of 0.0006–0.002 ppm and candidate RfDs of 0.0003–0.001 mg/kg/d). The most
3 sensitive candidate reference values also generally have low composite uncertainty factors (with
4 the exception of some mouse immunological and kidney effects), so are expected to be reflective
5 of the most sensitive effects as well. Thus, the most sensitive candidate references values for
6 multiple effects span about an order of magnitude for both inhalation (0.0003–0.003 ppm
7 [0.002–0.02 mg/m³]) and oral (0.0004–0.005 mg/kg/d) exposures. The most sensitive candidate
8 references values for neurological and reproductive effects are about an order of magnitude
9 higher (lowest candidate RfCs of 0.007–0.02 ppm [0.04–0.1 mg/m³]) and lowest candidate RfDs
10 of 0.009–0.02 mg/kg/d). Lastly, the liver effects have candidate reference values that are another
11 2 orders of magnitude higher (candidate RfCs of 1–2 ppm [6–10 mg/m³]) and candidate RfDs of
12 0.9–2 mg/kg/d).

13

14 **6.2.1.4. Noncancer Reference Values (see Section 5.1.5)**

15 **6.2.1.4.1. Reference concentration.** The goal is to select an overall RfC that is well supported
16 by the available data (i.e., without excessive uncertainty given the extensive database) and
17 protective for all the candidate critical effects, recognizing that individual candidate RfC values
18 are by nature somewhat imprecise. As discussed in Section 5.1 in Chapter 5, the lowest
19 candidate RfC values within each health effect category span a 3000-fold range from 0.0003–
20 0.9 ppm (see Table 5-21). One approach to selecting a RfC would be to select the lowest
21 calculated value of 0.0003 ppm for decreased thymus weight in mice. However, six candidate
22 RfCs (cRfCs and p-cRfCs) from both oral and inhalation studies are in the relatively narrow
23 range of 0.0003–0.003 ppm at the low end of the overall range (see Table 5-19). Given the
24 somewhat imprecise nature of the individual candidate RfC values, and the fact that multiple
25 effects/studies lead to similar candidate RfC values, the approach taken in this assessment is to
26 select a RfC supported by multiple effects/studies. The advantages of this approach, which is
27 only possible when there is a relatively large database of studies/effects and when multiple
28 candidate values happen to fall within a narrow range at the low end of the overall range, are that
29 it leads to a more robust RfC (less sensitive to limitations of individual studies) and that it
30 provides the important characterization that the RfC exposure level is similar for multiple
31 noncancer effects rather than being based on a sole explicit critical effect.

32 Therefore, six critical studies/effects were chosen to support the RfC for TCE noncancer
33 effects (see Table 5-23). Five of the lowest candidate RfCs, ranging from 0.0003–0.003 ppm for
34 developmental, kidney, and immunologic effects, are values derived from route-to-route
35 extrapolation using the PBPK model. The lowest candidate RfC estimate from an inhalation
36 study is 0.001 ppm for kidney effects. For all six candidate RfCs, the PBPK model was used for

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1 inter- and intraspecies extrapolation, based on the preferred dose metric for each endpoint. There
2 is high confidence in the candidate RfCs for kidney effects for the following reasons: they are
3 based on clearly adverse effects, two of the values are derived from chronic studies, and the
4 extrapolation to humans is based on dose metrics clearly related to toxicity estimated with high
5 confidence with the PBPK model developed in Section 3.5. There is somewhat less confidence
6 in the lowest candidate RfC for developmental effects (heart malformations) (see
7 Section 5.1.2.8), and the lowest candidate RfC estimates for immunological effects (see
8 Section 5.1.2.5). Thus, this assessment does not rely on any single estimate alone; however,
9 each estimate is supported by estimates of similar magnitude from other effects.

10 As a whole, the estimates support a preferred RfC estimate of 0.001 ppm (1 ppb or
11 $5 \mu\text{g}/\text{m}^3$). This estimate is within approximately a factor of 3 of the lowest estimates of
12 0.0003 ppm for decreased thymus weight in mice, 0.0004 ppm for heart malformations in rats,
13 0.0006 ppm for toxic nephropathy in rats, 0.001 ppm for increased kidney weight in rats,
14 0.002 ppm for toxic nephrosis in mice, and 0.003 ppm for increased anti-dsDNA antibodies in
15 mice. Thus, there is robust support for an RfC of 0.001 ppm provided by estimates for multiple
16 effects from multiple studies. The estimates are based on PBPK model-based estimates of
17 internal dose for interspecies, intraspecies, and/or route-to-route extrapolation, and there is
18 sufficient confidence in the PBPK model, as well as support from mechanistic data for some of
19 the dose metrics (specifically total oxidative metabolism for the heart malformations and
20 bioactivation of DCVC and total GSH metabolism for toxic nephropathy) (see Section 5.1.3.1).
21 Note that there is some human evidence of developmental heart defects from TCE exposure in
22 community studies (see Section 4.8.3.1.1) and of kidney toxicity in TCE-exposed workers (see
23 Section 4.4.1).

24 In summary, the preferred RfC estimate is **0.001 ppm** (1 ppb or $5 \mu\text{g}/\text{m}^3$) based on route-
25 to-route extrapolated results from oral studies for the critical effects of heart malformations
26 (rats), immunotoxicity (mice), and toxic nephropathy (rats, mice), and an inhalation study for the
27 critical effect of increased kidney weight (rats).
28

29 **6.2.1.4.2. Reference dose.** As with the RfC determination above, the goal is to select an overall
30 RfD that is well supported by the available data (i.e., without excessive uncertainty given the
31 extensive database) and protective for all the candidate critical effects, recognizing that
32 individual candidate RfD values are by nature somewhat imprecise. As discussed in Section 5.1
33 in Chapter 5, the lowest candidate RfD values (cRfDs and p-cRfDs) within each health effect
34 category span a nearly 3000-fold range from 0.0003–0.8 mg/kg/d (see Table 5-21). However,
35 four candidate RfDs from oral studies are in the relatively narrow range of

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1 0.0003–0.0005 mg/kg/d at the low end of the overall range. Given the somewhat imprecise
2 nature of the individual candidate RfD values, and the fact that multiple effects/studies lead to
3 similar candidate RfD values, the approach taken in this assessment is to select a RfD supported
4 by multiple effects/studies. The advantages of this approach, which is only possible when there
5 is a relatively large database of studies/effects and when multiple candidate values happen to fall
6 within a narrow range at the low end of the overall range, are that it leads to a more robust RfD
7 (less sensitive to limitations of individual studies) and that it provides the important
8 characterization that the RfD exposure level is similar for multiple noncancer effects rather than
9 being based on a sole explicit critical effect.

10 Therefore, four critical studies/effects were chosen to support the RfD for TCE noncancer
11 effects (see Table 5-24). Three of the lowest candidate RfDs—0.0003 mg/kg/d for toxic
12 nephropathy in rats, and 0.0005 mg/kg/d for heart malformations in rats and decreased thymus
13 weights in mice—are derived using the PBPK model for inter- and intraspecies extrapolation,
14 based on the preferred dose metric for each endpoint. The other of these lowest candidate
15 RfDs—0.0004 mg/kg/d for developmental immunotoxicity (decreased PFC response and
16 increased delayed-type hypersensitivity) in mice—is based on applied dose. There is high
17 confidence in the candidate RfD for kidney effects (see Section 5.1.2.2), which is based on
18 clearly adverse effects, derived from a chronic study, and extrapolated to humans based on a
19 dose metric clearly related to toxicity estimated with high confidence with the PBPK model
20 developed in Section 3.5. There is somewhat less confidence in the candidate RfDs for
21 decreased thymus weights (see Section 5.1.2.5) and heart malformations and developmental
22 immunological effects (see Section 5.1.2.8). Thus, this assessment does not rely on any single
23 estimate alone; however, each estimate is supported by estimates of similar magnitude from
24 other effects. As a whole, the estimates support a preferred RfD of 0.0004 mg/kg/d. This
25 estimate is within 25% of the lowest estimates of 0.0003 for toxic nephropathy in rats,
26 0.0004 mg/kg/d for developmental immunotoxicity (decreased PFC and increased delayed-type
27 hypersensitivity) in mice, and 0.0005 mg/kg/d for heart malformations in rats and decreased
28 thymus weights in mice. Thus, there is strong, robust support for an RfD of 0.0004 mg/kg/d
29 provided by the concordance of estimates derived from multiple effects from multiple studies.
30 The estimates for kidney effects, thymus effects, and developmental heart malformations are
31 based on PBPK model-based estimates of internal dose for interspecies and intraspecies
32 extrapolation, and there is sufficient confidence in the PBPK model, as well as support from
33 mechanistic data for some of the dose metrics (specifically total oxidative metabolism for the
34 heart malformations and bioactivation of DCVC for toxic nephropathy) (see Section 5.1.3.1).
35 Note that there is some human evidence of developmental heart defects from TCE exposure in

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1 community studies (see Section 4.8.3.1.1) and of kidney toxicity in TCE-exposed workers (see
2 Section 4.4.1).

3 In summary, the preferred RfD estimate is **0.0004 mg/kg/d** based on the critical effects of
4 heart malformations (rats), adult immunological effects (mice), developmental immunotoxicity
5 (mice), and toxic nephropathy (rats).

6 7 **6.2.2. Cancer (see Section 5.2)**

8 **6.2.2.1. Background and Methods (rodent: see Section 5.2.1.1; human: see Section 5.2.2.1)**

9 As summarized above, following U.S. EPA (2005a) *Guidelines for Carcinogen Risk*
10 *Assessment*, TCE is characterized as “*Carcinogenic to Humans*” by all routes of exposure, based
11 on convincing evidence of a causal association between TCE exposure in humans and kidney
12 cancer, but there is also human evidence of TCE carcinogenicity in the liver and lymphoid
13 tissues. This conclusion is further supported by rodent bioassay data indicating carcinogenicity
14 of TCE in rats and mice at tumor sites that include those identified in human epidemiologic
15 studies. Therefore, both human epidemiologic studies as well as rodent bioassays were
16 considered for deriving PODs for dose-response assessment of cancer endpoints. For PODs
17 derived from rodent bioassays, default dosimetry procedures were applied to convert applied
18 rodent doses to human equivalent doses. Essentially, for inhalation exposures, “ppm
19 equivalence” across species was assumed. For oral doses, ³/₄-power body-weight scaling was
20 used, with a default average human body weight of 70 kg. In addition to applied doses, several
21 internal dose metrics estimated using a PBPK model for TCE and its metabolites were used in
22 the dose-response modeling for each tumor type. In general, an attempt was made to use tissue-
23 specific dose metrics representing particular pathways or metabolites identified from available
24 data as having a likely role in the induction of a tissue-specific cancer. Where insufficient
25 information was available to establish particular metabolites or pathways of likely relevance to a
26 tissue-specific cancer, more general “upstream” metrics had to be used. In addition, the selection
27 of dose metrics was limited to metrics that could be adequately estimated by the PBPK model.

28 Regarding low-dose extrapolation, a key consideration in determining what extrapolation
29 approach to use is the MOA(s). However, MOA data are lacking or limited for each of the
30 cancer responses associated with TCE exposure, with the exception of the kidney tumors. For
31 the kidney tumors, the weight of the available evidence supports the conclusion that a mutagenic
32 MOA is operative; this MOA supports linear low-dose extrapolation. For the other TCE-induced
33 tumors, the MOA(s) is unknown. When the MOA(s) cannot be clearly defined, U.S. EPA
34 generally uses a linear approach to estimate low-dose risk (U.S. EPA, 2005a), based on the
35 following general principles:

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- 1 • A chemical’s carcinogenic effects may act additively to ongoing biological processes,
2 given that diverse human populations are already exposed to other agents and have
3 substantial background incidences of various cancers.
- 4 • A broadening of the dose-response curve (i.e., less rapid fall-off of response with
5 decreasing dose) in diverse human populations and, accordingly, a greater potential for
6 risks from low-dose exposures (Ziese et al., 1987; Lutz et al., 2005) is expected for two
7 reasons: First, even if there is a “threshold” concentration for effects at the cellular level,
8 that threshold is expected to differ across individuals. Second, greater variability in
9 response to exposures would be anticipated in heterogeneous populations than in inbred
10 laboratory species under controlled conditions (due to, e.g., genetic variability, disease
11 status, age, nutrition, and smoking status).
- 12 • The general use of linear extrapolation provides reasonable upper-bound estimates that
13 are believed to be health-protective (U.S. EPA, 2005a) and also provides consistency
14 across assessments.

15 **6.2.2.2. Inhalation Unit Risk Estimate (rodent: see Section 5.2.1.3; human: see**
16 **Section 5.2.2.1 and 5.2.2.2)**

17 The inhalation unit risk for TCE is defined as a plausible upper bound lifetime extra risk
18 of cancer from chronic inhalation of TCE per unit of air concentration. The preferred estimate of
19 the inhalation unit risk for TCE is 2.20×10^{-2} per ppm (**2×10^{-2} per ppm [4×10^{-6} per $\mu\text{g}/\text{m}^3$]**
20 rounded to 1 significant figure), based on human kidney cancer risks reported by Charbotel et al.
21 (2006) and adjusted for potential risk for tumors at multiple sites. This estimate is based on
22 good-quality human data, thus, avoiding the uncertainties inherent in interspecies extrapolation.
23 The Charbotel et al. (2006) case-control study of 86 incident renal cell carcinoma (RCC) cases
24 and 316 age- and sex-matched controls, with individual cumulative exposure estimates for TCE
25 inhalation for each subject, provides a sufficient human data set for deriving quantitative cancer
26 risk estimates for RCC in humans. The study is a high-quality study which used a detailed
27 exposure assessment (Fevotte et al., 2006) and took numerous potential confounding factors,
28 including exposure to other chemicals, into account. A significant dose-response relationship
29 was reported for cumulative TCE exposure and RCC (Charbotel et al., 2006). Human data on
30 TCE exposure and cancer risk sufficient for dose-response modeling are only available for RCC,
31 yet human and rodent data suggest that TCE exposure increases the risk of cancer at other sites
32 as well. In particular, there is evidence from human (and rodent) studies for increased risks of
33 lymphoma and liver cancer. Therefore, the inhalation unit risk estimate derived from human
34 data for RCC incidence was adjusted to account for potential increased risk of those tumor types.
35 To make this adjustment, a factor accounting for the relative contributions to the extra risk for
36 cancer incidence from TCE exposure for these three tumor types combined versus the extra risk
37 for RCC alone was estimated, and this factor was applied to the unit risk estimate for RCC to

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1 obtain a unit risk estimate for the three tumor types combined (i.e., lifetime extra risk for
2 developing *any* of the 3 types of tumor). This estimate is considered a better estimate of total
3 cancer risk from TCE exposure than the estimate for RCC alone. Although only the Charbotel et
4 al. (2006) study was found adequate for direct estimation of inhalation unit risks, the available
5 epidemiologic data provide sufficient information for estimating the *relative* potency of TCE
6 across tumor sites. In particular, the relative contributions to extra risk (for cancer incidence)
7 were calculated from two different data sets to derive the adjustment factor for adjusting the unit
8 risk estimate for RCC to a unit risk estimate for the 3 types of cancers (RCC, lymphoma, and
9 liver) combined. The first calculation is based on the results of the meta-analyses of human
10 epidemiologic data for the 3 tumor types; the second calculation is based on the results of the
11 Raaschou-Nielsen et al. (2003) study, the largest single human epidemiologic study by far with
12 RR estimates for all 3 tumor types. Both calculations support a 4-fold adjustment factor.

13 The preferred estimate of the inhalation unit risk based on human epidemiologic data is
14 supported by inhalation unit risk estimates from multiple rodent bioassays, the most sensitive of
15 which range from 1×10^{-2} to 2×10^{-1} per ppm [2×10^{-6} to 3×10^{-5} per $\mu\text{g}/\text{m}^3$]. From the
16 inhalation bioassays selected for analysis in Section 5.2.1.1, and using the preferred PBPK
17 model-based dose metrics, the inhalation unit risk estimate for the most sensitive sex/species is
18 8×10^{-2} per ppm [2×10^{-5} per $\mu\text{g}/\text{m}^3$], based on kidney adenomas and carcinomas reported by
19 Maltoni et al. (1986) for male Sprague-Dawley rats. Leukemias and Leydig cell tumors were
20 also increased in these rats, and, although a combined analysis for these tumor types which
21 incorporated the different site-specific preferred dose metrics was not performed, the result of
22 such an analysis is expected to be similar, about 9×10^{-2} per ppm [2×10^{-5} per $\mu\text{g}/\text{m}^3$]. The next
23 most sensitive sex/species from the inhalation bioassays is the female mouse, for which
24 lymphomas were reported by Henschler et al. (1980); these data yield a unit risk estimate of
25 1.0×10^{-2} per ppm [2×10^{-6} per $\mu\text{g}/\text{m}^3$]. In addition, the 90% confidence intervals (i.e., 5% to
26 95% bounds) reported in Table 5-34 for male rat kidney tumors from Maltoni et al. (1986) and
27 female mouse lymphomas from Henschler et al. (1980), derived from the quantitative analysis of
28 PBPK model uncertainty, both included the estimate based on human data of 2×10^{-2} per ppm.
29 Furthermore, PBPK model-based route-to-route extrapolation of the results for the most sensitive
30 sex/species from the oral bioassays, kidney tumors in male Osborne-Mendel rats and testicular
31 tumors in Marshall rats (NTP, 1988), leads to inhalation unit risk estimates of 2×10^{-1} per ppm
32 [3×10^{-5} per $\mu\text{g}/\text{m}^3$] and 4×10^{-2} per ppm [8×10^{-6} per $\mu\text{g}/\text{m}^3$], respectively, with the preferred
33 estimate based on human data falling within the route-to-route extrapolation of the 90%
34 confidence intervals reported in Table 5-35. Finally, for all these estimates, the ratios of BMDs

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1 to the BMDLs did not exceed a value of 3, indicating that the uncertainties in the dose-response
2 modeling for determining the POD in the observable range are small.

3 Although there are uncertainties in these various estimates, confidence in the proposed
4 inhalation unit risk estimate of 2×10^{-2} per ppm [4×10^{-6} per $\mu\text{g}/\text{m}^3$], based on human kidney
5 cancer risks reported by Charbotel et al. (2006) and adjusted for potential risk for tumors at
6 multiple sites (as summarized above in Section 6.1.4), is further increased by the similarity of
7 this estimate to estimates based on multiple rodent data sets. Application of the ADAF for
8 kidney cancer risks due to the weight of evidence supporting a mutagenic MOA for this endpoint
9 is summarized below in Section 6.2.2.5.

11 **6.2.2.3. Oral Unit Risk Estimate (rodent: see Section 5.2.1.3; human: see Section 5.2.2.3)**

12 The oral unit risk (or slope factor) for TCE is defined as a plausible upper bound lifetime
13 extra risk of cancer from chronic ingestion of TCE per mg/kg/d oral dose. The preferred
14 estimate of the oral unit risk is 4.63×10^{-2} per mg/kg/d (**5×10^{-2} per mg/kg/d** rounded to
15 1 significant figure), resulting from PBPK model-based route-to-route extrapolation of the
16 inhalation unit risk estimate based on the human kidney cancer risks reported in Charbotel et al.
17 (2006) and adjusted for potential risk for tumors at multiple sites. This estimate is based on
18 good-quality human data, thus, avoiding uncertainties inherent in interspecies extrapolation. In
19 addition, uncertainty in the PBPK model-based route-to-route extrapolation is relatively low
20 (Chiu and White, 2006; Chiu, 2006). In this particular case, extrapolation using different dose
21 metrics yielded expected population mean risks within about a 2-fold range, and, for any
22 particular dose metric, the 95% confidence interval for the extrapolated population mean risks
23 for each site spanned a range of no more than about 3-fold.

24 This value is supported by oral unit risk estimates from multiple rodent bioassays, the
25 most sensitive of which range from **3×10^{-2} to 3×10^{-1} per mg/kg/d**. From the oral bioassays
26 selected for analysis in Section 5.2.1.1, and using the preferred PBPK model-based dose metrics,
27 the oral unit risk estimate for the most sensitive sex/species is 3×10^{-1} per mg/kg/d, based on
28 kidney tumors in male Osborne-Mendel rats (NTP, 1988). The oral unit risk estimate for
29 testicular tumors in male Marshall rats (NTP, 1988) is somewhat lower at 7×10^{-2} per mg/kg/d.
30 The next most sensitive sex/species result from the oral studies is for male mouse liver tumors
31 (NCI, 1976), with an oral unit risk estimate of 3×10^{-2} per mg/kg/d. In addition, the 90%
32 confidence intervals reported in Table 5-35 for male Osborne-Mendel rat kidney tumors (NTP,
33 1988), male F344 rat kidney tumors (NTP, 1990), and male Marshall rat testicular tumors (NTP,
34 1988), derived from the quantitative analysis of PBPK model uncertainty, all included the
35 estimate based on human data of 5×10^{-2} per mg/kg/d, while the upper 95% confidence bound

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1 for male mouse liver tumors from NCI (1976) was slightly below this value at 4×10^{-2} per
2 mg/kg/d. Furthermore, PBPK model-based route-to-route extrapolation of the most sensitive
3 endpoint from the inhalation bioassays, male rat kidney tumors from Maltoni et al. (1986), leads
4 to an oral unit risk estimate of 1×10^{-1} per mg/kg/d, with the preferred estimate based on human
5 data falling within the route-to-route extrapolation of the 90% confidence interval reported in
6 Table 5-34. Finally, for all these estimates, the ratios of BMDs to the BMDLs did not exceed a
7 value of 3, indicating that the uncertainties in the dose-response modeling for determining the
8 POD in the observable range are small.

9 Although there are uncertainties in these various estimates, confidence in the proposed
10 oral unit risk estimate of 5×10^{-2} per mg/kg/d, resulting from PBPK model-based route-to-route
11 extrapolation of the inhalation unit risk estimate based on the human kidney cancer risks reported
12 in Charbotel et al. (2006) and adjusted for potential risk for tumors at multiple sites (as
13 summarized above), is further increased by the similarity of this estimate to estimates based on
14 multiple rodent data sets. Application of the ADAF for kidney cancer risks due to the weight of
15 evidence supporting a mutagenic MOA for this endpoint is summarized below in Section 6.2.2.5.

16 17 **6.2.2.4. Uncertainties in Cancer Dose-Response Assessment**

18 **6.2.2.4.1. Uncertainties in estimates based on human epidemiologic data (see**
19 **Section 5.2.2.1.3).** All risk assessments involve uncertainty, as study data are extrapolated to
20 make inferences about potential effects in humans from environmental exposure. The preferred
21 values for the unit risk estimates are based on good quality human data, which avoids
22 interspecies extrapolation, one of the major sources of uncertainty in quantitative cancer risk
23 estimates.

24 A remaining major uncertainty in the unit risk estimate for RCC incidence derived from
25 the Charbotel et al. (2006) is the extrapolation from occupational exposures to lower
26 environmental exposures. There was some evidence of a contribution to increased RCC risk
27 from peak exposures; however, there remained an apparent dose-response relationship for RCC
28 risk with increasing cumulative exposure without peaks, and the OR for exposure with peaks
29 compared to exposure without peaks was not significantly elevated (Charbotel et al., 2006).
30 Although the actual exposure-response relationship at low exposure levels is unknown, the
31 conclusion that a mutagenic MOA is operative for TCE-induced kidney tumors supports the
32 linear low-dose extrapolation that was used (U.S. EPA, 2005a). Additional support for use of
33 linear extrapolation is discussed above in Section 6.2.2.1.

34 In addition, because a linear model was used in the observable range of the human data
35 and the POD was within the low-dose linear range for extra risk as a function of exposure, linear

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1 extrapolation below the 95% lower confidence limit of the effective concentration for a 1%
2 response (LEC_{01}) is virtually a straight continuation of the 95% upper confidence limit on the
3 linear model used above the LEC_{01} . Thus, the use of linear extrapolation from the POD differed
4 negligibly from extrapolation of the dose-response model itself to low dose.

5 With respect to uncertainties in the dose-response modeling, the two-step approach of
6 modeling only in the observable range, as put forth in U.S. EPA's *Guidelines for Carcinogen*
7 *Risk Assessment* (U.S. EPA, 2005a), is designed in part to minimize model dependence. The
8 ratio of the maximum likelihood estimate of the effective concentration for a 1% response (EC_{01})
9 to the LEC_{01} , which gives some indication of the uncertainties in the dose-response modeling,
10 was about a factor of 2. Thus, overall, modeling uncertainties in the observable range are
11 considered to be negligible.

12 An important source of uncertainty in the underlying Charbotel et al. (2006) study is the
13 retrospective estimation of TCE exposures in the study subjects. This case-control study was
14 conducted in the Arve Valley in France, a region with a high concentration of screw cutting
15 workshops using TCE and other degreasing agents. Since the 1960s, occupational physicians of
16 the region have collected a large quantity of well-documented measurements, including TCE air
17 concentrations and urinary metabolite levels (Fevotte et al., 2006). The study investigators
18 conducted a comprehensive exposure assessment to estimate cumulative TCE exposures for the
19 individual study subjects, using a detailed occupational questionnaire with a customized task-
20 exposure matrix for the screw-cutting workers and a more general occupational questionnaire for
21 workers exposed to TCE in other industries (Fevotte et al., 2006). The exposure assessment also
22 attempted to take dermal exposure from hand-dipping practices into account by equating it with
23 an equivalent airborne concentration based on biological monitoring data. Despite the
24 appreciable effort of the investigators, considerable uncertainty associated with any retrospective
25 exposure assessment is inevitable, and some exposure misclassification is unavoidable. Such
26 exposure misclassification was most likely for the 19 deceased cases and their matched controls,
27 for which proxy respondents were used, and for exposures outside the screw-cutting industry
28 (295 of 1,486 identified job periods involved TCE exposure; 120 of these were not in the screw-
29 cutting industry).

30 Another noteworthy source of uncertainty in the Charbotel et al. (2006) study is the
31 possible influence of potential confounding or modifying factors. This study population, with a
32 high prevalence of metal-working, also had relatively high prevalences of exposure to petroleum
33 oils, cadmium, petroleum solvents, welding fumes, and asbestos (Fevotte et al., 2006). Other
34 exposures assessed included other solvents (including other chlorinated solvents), lead, and
35 ionizing radiation. None of these exposures was found to be significantly associated with RCC

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1 at a $p = 0.05$ significance level. Cutting fluids and other petroleum oils were associated with
2 RCC at a $p = 0.1$ significance level; however, further modeling suggested no association with
3 RCC when other significant factors were taken into account (Charbotel et al., 2006). The
4 medical questionnaire included familial kidney disease and medical history, such as kidney
5 stones, infection, chronic dialysis, hypertension, and use of anti-hypertensive drugs, diuretics,
6 and analgesics. Body mass index (BMI) was also calculated, and lifestyle information such as
7 smoking habits and coffee consumption was collected. Univariate analyses found high levels of
8 smoking and BMI to be associated with increased odds of RCC, and these two variables were
9 included in the conditional logistic regressions. Thus, although impacts of other factors are
10 possible, this study took great pains to attempt to account for potential confounding or modifying
11 factors.

12 Some other sources of uncertainty associated with the epidemiological data are the dose
13 metric and lag period. As discussed above, there was some evidence of a contribution to
14 increased RCC risk from peak TCE exposures; however, there appeared to be an independent
15 effect of cumulative exposure without peaks. Cumulative exposure is considered a good
16 measure of total exposure because it integrates exposure (levels) over time. If there is a
17 contributing effect of peak exposures, not already taken into account in the cumulative exposure
18 metric, the linear slope may be overestimated to some extent. Sometimes cancer data are
19 modeled with the inclusion of a lag period to discount more recent exposures not likely to have
20 contributed to the onset of cancer. In an unpublished report (Charbotel et al., 2005), Charbotel et
21 al. also present the results of a conditional logistic regression with a 10-year lag period, and these
22 results are very similar to the unlagged results reported in their published paper, suggesting that
23 the lag period might not be an important factor in this study.

24 Some additional sources of uncertainty are not so much inherent in the exposure-response
25 modeling or in the epidemiologic data themselves but, rather, arise in the process of obtaining
26 more general Agency risk estimates from the epidemiologic results. U.S. EPA cancer risk
27 estimates are typically derived to represent an upper bound on increased risk of cancer incidence
28 for all sites affected by an agent for the general population. From experimental animal studies,
29 this is accomplished by using tumor incidence data and summing across all the tumor sites that
30 demonstrate significantly increased incidences, customarily for the most sensitive sex and
31 species, to attempt to be protective of the general human population. However, in estimating
32 comparable risks from the Charbotel et al. (2006) epidemiologic data, certain limitations are
33 encountered. For one thing, these epidemiology data represent a geographically limited (Arve
34 Valley, France) and likely not very diverse population of working adults. Thus, there is
35 uncertainty about the applicability of the results to a more diverse general population.

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1 Additionally, the Charbotel et al. (2006) study was a study of RCC only, and so the risk
2 estimate derived from it does not represent all the tumor sites that may be affected by TCE. This
3 uncertainty was addressed by adjusting the RCC estimate to multiple sites, but there are also
4 uncertainties related to the assumptions inherent in the calculations for this adjustment. As
5 discussed in Section 5.2.2.2, adequate quantitative dose-response data were only available for
6 one cancer site in humans, so other human data were used to adjust the estimate derived for RCC
7 to include risk for other cancers with substantial human evidence of hazard (lymphoma and liver
8 cancer). The relative contributions to extra risk (for cancer incidence) were calculated from two
9 different data sets to derive an adjustment factor. The first calculation is based on the results of
10 the meta-analyses for the 3 tumor types; the second calculation is based on the results of the
11 Raaschou-Nielsen et al. (2003) study, the largest single study by far with RR estimates for all 3
12 tumor types. The fact that the calculations based on 2 different data sets yielded comparable
13 values for the adjustment factor provides more robust support for the use of the factor of 4.
14 Additional uncertainties pertain to the weight of evidence supporting the association of TCE
15 exposure with increased risk of cancer for the 3 tumor types. As discussed in Section 4.11.2, it is
16 concluded that the weight of evidence for kidney cancer is sufficient to classify TCE as
17 “Carcinogenic to Humans.” It is also concluded that there is strong evidence that TCE causes
18 lymphoma as well, although the evidence for liver cancer was more limited. In addition, the
19 rodent studies demonstrate clear evidence of multisite carcinogenicity, with tumor types
20 including those for which associations with TCE exposure are observed in human studies, i.e.,
21 liver and kidney cancers and lymphomas. Overall, the evidence is sufficiently persuasive to
22 support the use of the adjustment factor of 4 based on these 3 tumor types. Alternatively, if one
23 were to use the factor based only on the 2 tumor types with the strongest evidence, the cancer
24 inhalation unit risk estimate would be only slightly reduced (25%).

25 Finally, the preferred value for the oral unit risk estimate was based on route-to-route
26 extrapolation of the inhalation unit risk based on human data using predictions from the PBPK
27 model. Because different internal dose metrics are preferred for each target tissue site, a separate
28 route-to-route extrapolation was performed for each site-specific unit risk estimate. As discussed
29 above, uncertainty in the PBPK model-based route-to-route extrapolation is relatively low (Chiu
30 and White, 2006; Chiu, 2006). In this particular case, extrapolation using different dose metrics
31 yielded expected population mean risks within about a 2-fold range, and, for any particular dose
32 metric, the 95% confidence interval for the extrapolated population mean risks for each site
33 spanned a range of no more than about 3-fold.

1 **6.2.2.4.2. Uncertainties in estimates based on rodent bioassays (see Section 5.2.1.4).** With
2 respect to rodent-based cancer risk estimates, the cancer risk is typically estimated from the total
3 cancer burden from all sites that demonstrate an increased tumor incidence for the most sensitive
4 experimental species and sex. It is expected that this approach is protective of the human
5 population, which is more diverse but is exposed to lower exposure levels. In the case of TCE,
6 the impact of selection of the bioassay is limited, since, as discussed in Sections 5.2.1.3 and
7 5.2.3, estimates based on the two or three most sensitive bioassays are within an order of
8 magnitude of each other, and are consistent across routes of exposure when extrapolated using
9 the PBPK model.

10 Another source of uncertainty in the TCE rodent-based cancer risk estimates is
11 interspecies extrapolation. Several plausible PBPK model-based dose metrics were used for
12 extrapolation of toxicokinetics, but the cancer unit risk estimates obtained using the preferred
13 dose metrics were generally similar (within about 3-fold) to those derived using default
14 dosimetry assumptions, with the exception of the bioactivated DCVC dose metric for rat kidney
15 tumors and the metric for the amount of TCE oxidized in the respiratory tract for mouse lung
16 tumors occurring from oral exposure. However, there is greater biological support for these
17 selected dose metrics. The uncertainty in the PBPK model predictions themselves were analyzed
18 quantitatively through an analysis of the impact of parameter uncertainties in the PBPK model.
19 The 95% lower bounds on the BMD including parameter uncertainties in the PBPK model were
20 no more than 4-fold lower than those based on central estimates of the PBPK model predictions.
21 The greatest uncertainty was for unit risks derived from rat kidney tumors, primarily reflecting
22 the substantial uncertainty in the rat internal dose.

23 Regarding low-dose extrapolation, a key consideration in determining what extrapolation
24 approach to use is the MOA(s). However, MOA data are lacking or limited for each of the
25 cancer responses associated with TCE exposure, with the exception of the kidney tumors. For
26 the kidney tumors, the weight of the available evidence supports the conclusion that a mutagenic
27 MOA is operative; this MOA supports linear low-dose extrapolation. For the other TCE-induced
28 tumors, the MOA(s) is unknown. When the MOA(s) cannot be clearly defined, U.S. EPA
29 generally uses a linear approach to estimate low-dose risk (U.S. EPA, 2005a), based on the
30 general principles discussed above.

31 With respect to uncertainties in the dose-response modeling, the two-step approach of
32 modeling only in the observable range, as put forth in U.S. EPA's *Guidelines for Carcinogen
33 Risk Assessment* (U.S. EPA, 2005a), is designed in part to minimize model dependence. The
34 ratios of the BMDs to the BMDLs, which give some indication of the uncertainties in the dose-
35 response modeling, did not exceed a value of 2.5 for all the primary analyses used in this

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1 assessment. Thus, overall, modeling uncertainties in the observable range are considered to be
2 negligible. Some additional uncertainty is conveyed by uncertainties in the survival adjustments
3 made to some of the bioassay data; however, a comparison of the results of two different survival
4 adjustment methods suggest that their impact is minimal relative to the uncertainties already
5 discussed.

6 7 **6.2.2.5. Application of Age-Dependent Adjustment Factors (see Section 5.2.3.3)**

8 When there is sufficient weight of evidence to conclude that a carcinogen operates
9 through a mutagenic MOA, and in the absence of chemical-specific data on age-specific
10 susceptibility, U.S. EPA's *Supplemental Guidance for Assessing Susceptibility from Early-Life*
11 *Exposure to Carcinogens* (U.S. EPA, 2005b) recommends the application of default ADAFs to
12 adjust for potential increased susceptibility from early-life exposure. See the *Supplemental*
13 *Guidance* for detailed information on the general application of these adjustment factors. In
14 brief, the *Supplemental Guidance* establishes ADAFs for three specific age groups. The current
15 ADAFs and their age groupings are 10 for <2 years, 3 for 2 to <16 years, and 1 for 16 years and
16 above (U.S. EPA, 2005b). For risk assessments based on specific exposure assessments, the
17 10-fold and 3-fold adjustments to the unit risk estimates are to be combined with age-specific
18 exposure estimates when estimating cancer risks from early-life (<16 years age) exposure. The
19 ADAFs and their age groups may be revised over time. The most current information on the
20 application of ADAFs for cancer risk assessment can be found at
21 www.epa.gov/cancerguidelines.

22 In the case of TCE, the inhalation and oral unit risk estimates reflect lifetime risk for
23 cancer at multiple sites, and a mutagenic MOA has been established for one of these sites, the
24 kidney. In addition, as discussed in Section 4.10, inadequate TCE-specific data exists to quantify
25 early-life susceptibility to TCE carcinogenicity; therefore, as recommended in the *Supplemental*
26 *Guidance*, the default ADAFs are used. As illustrated in the example calculations in
27 Section 5.2.3.3, application of the default ADAFs to the kidney cancer inhalation and oral unit
28 risk estimates for TCE is likely to have minimal impact on the total cancer risk except when
29 exposure is primarily during early life.

30 In addition to the uncertainties discussed above for the inhalation and oral total cancer
31 unit risk estimates, there are uncertainties in the application of ADAFs to adjust for potential
32 increased early-life susceptibility. The adjustment is made only for the kidney cancer component
33 of total cancer risk because that is the tumor type for which the weight of evidence was sufficient
34 to conclude that TCE-induced carcinogenesis operates through a mutagenic MOA. However, it
35 may be that TCE operates through a mutagenic MOA for other tumor types as well or that it

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1 operates through other MOAs that might also convey increased early-life susceptibility.
2 Additionally, the ADAFs are general default factors, and it is uncertain to what extent they
3 reflect increased early-life susceptibility for exposure to TCE, if increased early-life
4 susceptibility occurs.

6 **6.3. OVERALL CHARACTERIZATION OF TCE HAZARD AND DOSE RESPONSE**

7 There is substantial potential for human exposure to TCE, as it has a widespread presence
8 in ambient air, indoor air, soil, and groundwater. At the same time, humans are likely to be
9 exposed to a variety of compounds that are either metabolites of TCE or which have common
10 metabolites or targets of toxicity. Once exposed, humans, as well as laboratory animal species,
11 rapidly absorb TCE, which is then distributed to tissues via systemic circulation, extensively
12 metabolized, and then excreted primarily in breath as unchanged TCE or CO₂, or in urine as
13 metabolites.

14 Based on the available human epidemiologic data and experimental and mechanistic
15 studies, it is concluded that TCE poses a potential human health hazard for noncancer toxicity to
16 the central nervous system, the kidney, the liver, the immune system, the male reproductive
17 system, and the developing fetus. The evidence is more limited for TCE toxicity to the
18 respiratory tract and female reproductive system. Following U.S. EPA (2005a) *Guidelines for*
19 *Carcinogen Risk Assessment*, TCE is characterized as “*Carcinogenic to Humans*” by all routes
20 of exposure. This conclusion is based on convincing evidence of a causal association between
21 TCE exposure in humans and kidney cancer. The human evidence of carcinogenicity from
22 epidemiologic studies of TCE exposure is compelling for lymphoma, but less convincing than
23 for kidney cancer, and more limited for liver and biliary tract cancer. Further support for the
24 characterization of TCE as “*Carcinogenic to Humans*” by all routes of exposure is derived from
25 positive results in multiple rodent cancer bioassays in rats and mice of both sexes, similar
26 toxicokinetics between rodents and humans, mechanistic data supporting a mutagenic MOA for
27 kidney tumors, and the lack of mechanistic data supporting the conclusion that any of the
28 MOA(s) for TCE-induced rodent tumors are irrelevant to humans.

29 As TCE toxicity and carcinogenicity are generally associated with TCE metabolism,
30 susceptibility to TCE health effects may be modulated by factors affecting toxicokinetics,
31 including lifestage, gender, genetic polymorphisms, race/ethnicity, preexisting health status,
32 lifestyle, and nutrition status. In addition, while some of these factors are known risk factors for
33 effects associated with TCE exposure, it is not known how TCE interacts with known risk factors
34 for human diseases.

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1 For noncancer effects, the most sensitive types of effects, based either on human
2 equivalent concentrations/doses or on candidate RfCs/RfDs, appear to be developmental, kidney,
3 and immunological (adult and developmental) effects. The neurological and reproductive effects
4 appear to be about an order of magnitude less sensitive, with liver effects another two orders of
5 magnitude less sensitive. The preferred RfC estimate of **0.001 ppm** (1 ppb or 5 $\mu\text{g}/\text{m}^3$) is based
6 on route-to-route extrapolated results from oral studies for the critical effects of heart
7 malformations (rats), immunotoxicity (mice), and toxic nephropathy (rats, mice), and an
8 inhalation study for the critical effect of increased kidney weight (rats). Similarly, the preferred
9 RfD estimate for noncancer effects of **0.0004 mg/kg/d** is based on the critical effects of heart
10 malformations (rats), adult immunological effects (mice), developmental immunotoxicity (mice),
11 and toxic nephropathy (rats). There is high confidence in these preferred noncancer reference
12 values, as they are supported by moderate- to high-confidence estimates for multiple effects from
13 multiple studies.

14 For cancer, the preferred estimate of the inhalation unit risk is **2×10^{-2} per ppm**
15 [**4×10^{-6} per $\mu\text{g}/\text{m}^3$**], based on human kidney cancer risks reported by Charbotel et al. (2006)
16 and adjusted, using human epidemiologic data, for potential risk for tumors at multiple sites.
17 The preferred estimate of the oral unit risk for cancer is **5×10^{-2} per mg/kg/d**, resulting from
18 PBPK model-based route-to-route extrapolation of the inhalation unit risk estimate based on the
19 human kidney cancer risks reported in Charbotel et al. (2006) and adjusted, using human
20 epidemiologic data, for potential risk for tumors at multiple sites. There is high confidence in
21 these unit risks for cancer, as they are based on good quality human data, as well as being similar
22 to unit risk estimates based on multiple rodent bioassays. Because there is both sufficient weight
23 of evidence to conclude that TCE operates through a mutagenic MOA for kidney tumors and a
24 lack of TCE-specific quantitative data on early-life susceptibility, the default ADAFs can be
25 applied for the kidney cancer component of the unit risks for cancer; however, the application of
26 ADAFs is likely to have a minimal impact on the total cancer risk except when exposures are
27 primarily during early life.