

APPENDIX F

TCE Noncancer Dose-Response Analyses

This document is a draft for review purposes only and does not constitute Agency policy.

10/20/09

F-i

DRAFT—DO NOT CITE OR QUOTE

CONTENTS—Appendix F: TCE Noncancer Dose-Response Analyses

LIST OF TABLES	F-iv
LIST OF FIGURES	F-v
APPENDIX F: TCE NONCANCER DOSE-RESPONSE ANALYSES	F-1
F.1. DATA SOURCES	F-1
F.2. DOSIMETRY	F-1
F.2.1. Estimates of Trichlorethylene (TCE) in Air From Urinary Metabolite Data Using Ikeda et al. (1972)	F-1
F.2.1.1. Results for Chia et al. (1996)	F-1
F.2.1.2. Results for Mhiri et al. (2004)	F-4
F.2.2. Dose Adjustments to Applied Doses for Intermittent Exposure	F-4
F.2.3. Physiologically Based Pharmacokinetic (PBPK) Model-Based Internal Dose Metrics	F-5
F.3. DOSE-RESPONSE MODELING PROCEDURES	F-5
F.3.1. Models for Dichotomous Response Data	F-5
F.3.1.1. Quantal Models	F-5
F.3.1.2. Nested Dichotomous Models	F-6
F.3.2. Models for Continuous Response Data	F-6
F.3.3. Model Selection	F-6
F.3.4. Additional Adjustments for Selected Data Sets	F-7
F.4. DOSE-RESPONSE MODELING RESULTS	F-8
F.4.1. Quantal Dichotomous and Continuous Modeling Results	F-8
F.4.2. Nested Dichotomous Modeling Results	F-8
F.4.2.1. Johnson et al. (2003) Fetal Cardiac Defects	F-8
F.4.2.2. Narotsky et al. (1995)	F-12
F.4.3. Model Selections and Results	F-20
F.5. DERIVATION OF POINTS OF DEPARTURE	F-27
F.5.1. Applied Dose Points of Departure	F-27
F.5.2. Physiologically Based Pharmacokinetic (PBPK) Model-Based Human Points of Departure	F-27
F.6. SUMMARY OF POINTS OF DEPARTURE (PODs) FOR CRITICAL STUDIES AND EFFECTS SUPPORTING THE INHALATION REFERENCE CONCENTRATION (RfC) AND ORAL REFERENCE DOSE (RfD)	F-28
F.6.1. National Toxicology Program (NTP, 1988)—Benchmark Dose (BMD) Modeling of Toxic Nephropathy in Rats	F-28
F.6.1.1. Dosimetry and Benchmark Dose (BMD) Modeling	F-29
F.6.1.2. Derivation of HEC ₉₉ and HED ₉₉	F-29
F.6.2. National Cancer Institute (NCI, 1976)—Lowest-Observed-Adverse- Effect Level (LOAEL) for Toxic Nephrosis in Mice	F-29
F.6.2.1. Dosimetry	F-31
F.6.2.2. Derivation of HEC ₉₉ and HED ₉₉	F-31

This document is a draft for review purposes only and does not constitute Agency policy.

CONTENTS (continued)

F.6.3.	Woolhiser et al. (2006)—Benchmark Dose (BMD) Modeling of Increased Kidney Weight in Rats	F-32
	F.6.3.1. Dosimetry and Benchmark Dose (BMD) Modeling.....	F-32
	F.6.3.2. Derivation of HEC ₉₉ and HED ₉₉	F-34
F.6.4.	Keil et al. (2009)—Lowest-Observed-Adverse-Effect Level (LOAEL) for Decreased Thymus Weight and Increased Anti-dsDNA and Anti-ssDNA Antibodies in Mice	F-34
F.6.5.	Keil et al. (2009)—Lowest-Observed-Adverse-Effect Level (LOAEL) for Decreased Thymus Weight and Increased Anti-dsDNA and Anti-ssDNA Antibodies in Mice	F-34
	F.6.5.1. Dosimetry.....	F-34
	F.6.5.2. Derivation of HEC ₉₉ and HED ₉₉	F-35
F.6.6.	Johnson et al. (2003)—Benchmark Dose (BMD) Modeling of Fetal Heart Malformations in Rats.....	F-35
	F.6.6.1. Dosimetry and Benchmark Dose (BMD) Modeling.....	F-36
	F.6.6.2. Derivation of HEC ₉₉ and HED ₉₉	F-36
F.6.7.	Peden-Adams et al. (2006)—Lowest-Observed-Adverse-Effect Level (LOAEL) for Decreased PFC Response and Increased Delayed-Type Hypersensitivity in Mice.....	F-37
F.7.	REFERENCES	F-38

This document is a draft for review purposes only and does not constitute Agency policy.

LIST OF TABLES

F-1. Dose-response data from Chia et al. (1996)..... F-1

F-2. Data on TCE in air (ppm) and urinary metabolite concentrations in workers reported by Ikeda et al. (1972)..... F-2

F-3. Estimated urinary metabolite and TCE air concentrations in dose groups from Chia et al. (1996)..... F-4

F-4. Data on fetuses and litters with abnormal hearts from Johnson et al. (2003)..... F-9

F-5. Comparison of observed and predicted numbers of fetuses with abnormal hearts from Johnson et al. (2003), with and without the high-dose group, using a nested model..... F-9

F-6. Results of nested log-logistic model for fetal cardiac anomalies from Johnson et al. (2003) without the high-dose group, on the basis of applied dose (mg/kg/d in drinking water)..... F-10

F-7. Results of nested log-logistic model for fetal cardiac anomalies from Johnson et al. (2003) without the high-dose group, using the TotOxMetabBW34 dose metric F-13

F-8. Results of nested log-logistic model for fetal cardiac anomalies from Johnson et al. (2003) without the high-dose group, using the AUCCBld dose metric..... F-14

F-9. Analysis of LSCs with respect to dose from Narotsky et al. (1995)..... F-15

F-10. Results of nested log-logistic and Rai-VanRyzin model for fetal eye defects from Narotsky et al. (1995), on the basis of applied dose (mg/kg/d in drinking water) F-16

F-11. Comparison of results of nested log-logistic (without LSC or IC) and quantal log-logistic model for fetal eye defects from Narotsky et al. (1995)..... F-18

F-12. Results of nested log-logistic and Rai-VanRyzin model for prenatal loss from Narotsky et al. (1995), on the basis of applied dose (mg/kg/d in drinking water)..... F-20

F-13. Model selections and results for noncancer dose-response analyses..... F-23

This document is a draft for review purposes only and does not constitute Agency policy.

LIST OF FIGURES

F-1.	Regression of TCE in air (ppm) and TCA in urine (mg/g creatinine) based on data from Ikeda et al. (1972).....	F-3
F-2.	BMD modeling of Johnson et al. (2003) using nested log-logistic model, with applied dose, without LSC, with IC, and without the high-dose group, using a BMR of 0.05 extra risk (top panel) or 0.01 extra risk (bottom panel).....	F-11
F-3.	BMD modeling of Johnson et al. (2003) using nested log-logistic model, with TotOxMetabBW34 dose metric, without LSC, with IC, and without the high-dose group, using a BMR of 0.01 extra risk.....	F-13
F-4.	BMD modeling of Johnson et al. (2003) using nested log-logistic model, with AUCCBld dose metric, without LSC, with IC, and without the high-dose group, using a BMR of 0.01 extra risk.....	F-14
F-5.	BMD modeling of fetal eye defects from Narotsky et al. (1995) using nested log-logistic model, with applied dose, with both LSC and IC, using a BMR of 0.05 extra risk.....	F-17
F-6.	BMD modeling of fetal eye defects from Narotsky et al. (1995) using nested log-logistic model, with applied dose, without either LSC or IC, using a BMR of 0.05 extra risk.....	F-18
F-7.	BMD modeling of fetal eye defects from Narotsky et al. (1995) using nested Rai-VanRyzin model, with applied dose, without either LSC or IC, using a BMR of 0.05 extra risk.....	F-19
F-8.	BMD modeling of prenatal loss reported in Narotsky et al. (1995) using nested log-logistic model, with applied dose, without LSC, with IC, using a BMR of 0.05 extra risk (top panel) or 0.01 extra risk (bottom panel).....	F-21
F-9.	BMD modeling of prenatal loss reported in Narotsky et al. (1995) using nested Rai-VanRyzin model, with applied dose, without LSC, with IC, using a BMR of 0.05 extra risk or 0.01 extra risk.....	F-22
F-10.	BMD modeling of NTP (1988) toxic nephropathy in female Marshall rats.....	F-30
F-11.	Derivation of HEC ₉₉ and HED ₉₉ corresponding to the rodent idPOD from NTP (1988) toxic nephropathy in rats.....	F-31

This document is a draft for review purposes only and does not constitute Agency policy.

LIST OF FIGURES (continued)

F-12. Derivation of HEC₉₉ and HED₉₉ corresponding to the rodent idPOD from NTP (1988) toxic nephrosis in mice..... F-32

F-13. BMD modeling of Woolhiser et al. (2006) for increased kidney weight in female S-D rats. F-33

F-14. Derivation of HEC₉₉ and HED₉₉ corresponding to the rodent idPOD from Woolhiser et al. (2006) for increased kidney weight in rats. F-35

F-15. Derivation of HEC₉₉ and HED₉₉ corresponding to the rodent idPOD from Keil et al. (2009) for decreased thymus weight and increased anti-dsDNA and anti-ssDNA antibodies in mice. F-36

F-16. Derivation of HEC₉₉ and HED₉₉ corresponding to the rodent idPOD from Johnson et al. (2003) for increased fetal cardiac malformations in female S-D rats using the total oxidative metabolism dose metric. F-37

1 **APPENDIX F: TCE NONCANCER DOSE-RESPONSE ANALYSES**

2
3
4 **F.1. DATA SOURCES**

5 Data sources are cited in the body of this report in the section describing dose-response
6 analyses (see Chapter 5).

7
8 **F.2. DOSIMETRY**

9 This section describes some of the more detailed dosimetry calculations and adjustments
10 used in Section 5.1.

11
12 **F.2.1. Estimates of Trichloroethylene (TCE) in Air From Urinary Metabolite Data Using**
13 **Ikeda et al. (1972)**

14 **F.2.1.1. Results for Chia et al. (1996)**

15 Chia et al. (1996) demonstrated a dose-related effect on hyperzoospermia in male
16 workers exposed to trichloroethylene (TCE), lumping subjects into four groups based on range of
17 trichloroacetic acid (TCA) in urine (see Table F-1).

18
19 **Table F-1. Dose-response data from Chia et al. (1996)**

20

TCA, mg per g creatinine	No. of subjects	No. with hyperzoospermia
0.8 to <25	37	6
50 to <75	18	8
75 to <100	8	4
≥100 to 136.4	5	3

21
22 Minimum and maximum TCA levels are reported in the text of Chia et al. (1996), the other data, in their
23 Table 5.

24
25
26 Data from Ikeda et al. (1972) were used to estimate the TCE exposure concentrations
27 corresponding to the urinary TCA levels reported by Chia et al. (1996). Ikeda et al. (1972)
28 studied 10 workshops, in each of which TCE vapor concentration was “relatively constant.”
29 They measured atmospheric concentrations of TCE and concentrations in workers’ urine of total
30 trichloro compounds (TTC), TCA, and creatinine, and demonstrated a linear relation between
31 TTC/creatinine (mg/g) in urine and TCE in the work atmosphere. Their data are tabulated as
32 geometric means (the last column was calculated by us, as described in Table F-2).

This document is a draft for review purposes only and does not constitute Agency policy.

1 **Table F-2. Data on TCE in air (ppm) and urinary metabolite concentrations**
 2 **in workers reported by Ikeda et al. (1972)**
 3

<i>n</i>	TCE (ppm)	TTC (mg/L)	TCA (mg/L)	TTC (mg/g creatinine)	TCA (mg/g creatinine)
9	3	39.4	12.7	40.8	13.15127
5	5	45.6	20.2	42.4	18.78246
6	10	60.5	17.6	47.3	13.76
4	25	164.3	77.2	122.9	57.74729
4	40	324.9	90.6	221.2	61.68273
5	45	399	138.4	337.7	117.137
5	50	418.9	146.6	275.8	96.52012
5	60	468	155.4	359	119.2064
4	120	915.3	230.1	518.9	130.4478
4	175	1210.9	235.8	1040.1	202.5399

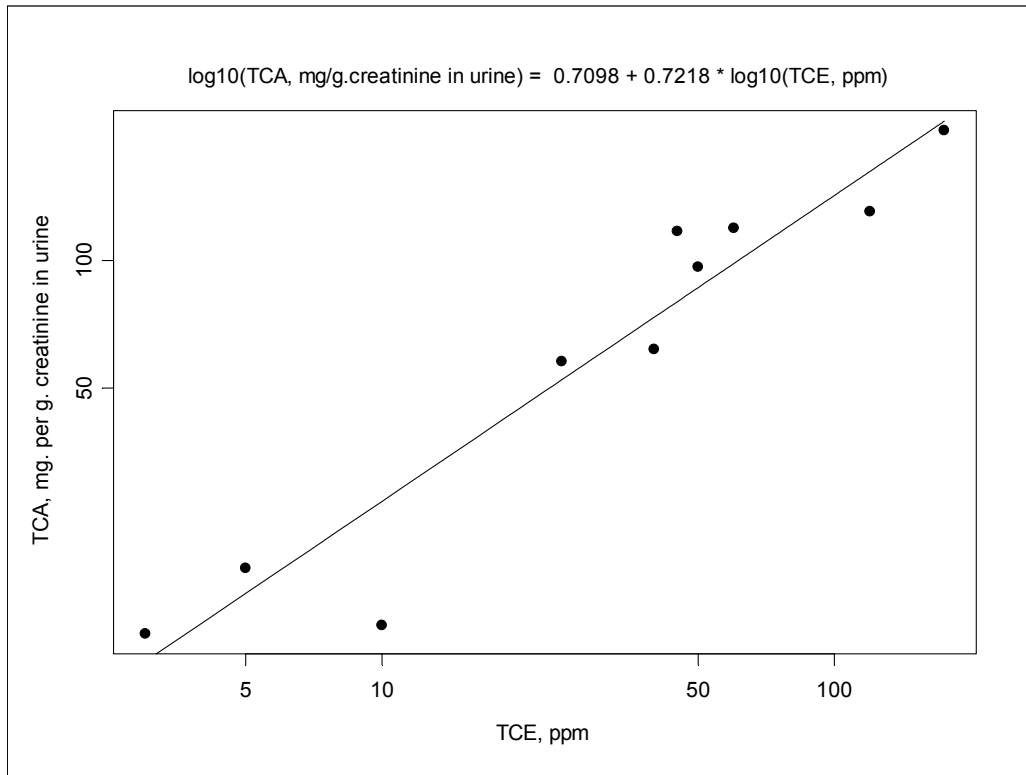
4
 5
 6 These data were used to construct the last column “TCA.cr.mg.g” (mg TCA/g creatinine),
 7 as follows: TCA (mg/g creatinine) = TCA (mg/L) × TTC (mg/g creatinine)/TTC (mg/L). The
 8 regression relation between TCE (ppm) and TCA (mg/g creatinine) was evaluated using these
 9 data. Ikeda et al. (1972) reported that the measured values are lognormally distributed and
 10 exhibit heterogeneity of variance, and that the reported data (above) are geometric means. Thus,
 11 the regression relation between log₁₀(TCA [mg/g creatinine]) and log₁₀(TCE [ppm]) was used,
 12 assuming constant variances and using number of subjects “*n*” as weights. Figure F-1 shows the
 13 results.

14 Next, a Berkson setting for linear calibration was assumed, in which one wants to predict
 15 *X* (TCE, ppm) from means for *Y* (TCA, mg/g creatinine), with substantial error in *Y* (Snedecor
 16 and Cochran, 1980). Thus, the inverse prediction for the data of Chia et al. (1996) was used to
 17 infer their mean TCE exposures. The relation based on data from Ikeda et al. (1972) is

$$\log_{10}(\text{TCA, mg/g creatinine}) = 0.7098 + 0.7218 \cdot \log_{10}(\text{TCE, ppm}) \quad (\text{Eq. F-1})$$

18
 19
 20
 21 and the inverse prediction is

1



2

Coefficients:				
	Value	Std. Error	t value	Pr(> t)
(Intercept)	0.7098	0.1132	6.2688	0.0002
log10(TCE.ppm)	0.7218	0.0771	9.3578	0.0000

Residual standard error: 0.3206 on 8 degrees of freedom
Multiple R-Squared: 0.9163
F-statistic: 87.57 on 1 and 8 degrees of freedom, the p-value is 0.0000139

3

4

5

6

7

8

9

Figure F-1. Regression of TCE in air (ppm) and TCA in urine (mg/g creatinine) based on data from Ikeda et al. (1972).

10

$$\log_{10}(\text{TCE}) = [\log_{10}(\text{TCA}) - 0.7098]/0.7218 \quad (\text{Eq. F-2})$$

11

$$\text{TCE, ppm} = 10^{([\log_{10}(\text{TCA}) - 0.7098]/0.7218)}$$

12

13

14

Because of the lognormality of data reported by Ikeda et al. (1972), the means of the logarithms of the ranges for TCA (mg/g creatinine) in Chia et al. (1996), which are estimates of the median for the group, were used. The results are shown in Table F-3.

This document is a draft for review purposes only and does not constitute Agency policy.

1 **Table F-3. Estimated urinary metabolite and TCE air concentrations in dose**
 2 **groups from Chia et al. (1996)**
 3

TCA, mg per g Creatinine	Estim. TCA median ^a	Log10(TCA median)	Estim. ppm TCE ^b
0.8 to <25	4.47	0.650515	0.827685
50 to <75	61.2	1.787016	31.074370
75 to <100	86.6	1.937531	50.226119
≥100 to 136.4	117	2.067407	76.008668

4
 5 ^a $10^{(\text{mean}[\log_{10}(\text{TCA limits in first column})])}$.

6 ^b $10^{([\log_{10}(\text{TCA median})] - 0.7098)/0.7218}$.

7
 8
 9 Dose-response relations for the data of Chia et al. (1996) were modeled using both the
 10 estimated medians for TCA (mg/g creatinine) in urine and estimated TCE (ppm in air) as doses.
 11 The TCE-TCA-TTC relations are linear up to about 75 ppm TCE (Figure 1 of Ikeda et al. 1972),
 12 and certainly in the range of the benchmark dose (BMD). As noted below (see Section F.2.2),
 13 the occupational exposure levels are further adjusted to equivalent continuous exposure for
 14 deriving the point of departure (POD).

15
 16 **F.2.1.2. Results for Mhiri et al. (2004)**

17 The lowest-observed-adverse-effect level (LOAEL) group for abnormal trigeminal nerve
 18 somatosensory evoked potential reported in Mhiri et al. (2004) had a urinary TCA concentration
 19 of 32.6 mg TCA/mg creatinine. Using Eq. F-2, above gives an occupational exposure level =
 20 $10^{([\log_{10}(32.6) - 0.7098]/0.7218)} = 12.97404$ ppm. As noted below (see Section F.2.2), the
 21 occupational exposure levels are further adjusted to equivalent continuous exposure for deriving
 22 the POD.

23
 24 **F.2.2. Dose Adjustments to Applied Doses for Intermittent Exposure**

25 The nominal applied dose was adjusted for exposure discontinuity (e.g., exposure for
 26 5 days per week and 6 hours per day reduced the dose by the factor $[5/7]*[6/24]$). The
 27 physiologically based pharmacokinetic (PBPK) dose metrics took into account the daily and
 28 weekly discontinuity to produce an equivalent average dose for continuous exposure. No dose
 29 adjustments were made for duration of exposure or a less-than-lifetime study, as is typically done
 30 for cancer risk estimates, though in deriving the candidate reference values, an uncertainty factor
 31 for subchronic-to-chronic exposure was applied where appropriate.

This document is a draft for review purposes only and does not constitute Agency policy.

1 For human occupational studies, inhalation exposures (air concentrations) were adjusted
2 by the number of work (vs. nonwork) days and the amount of air intake during working hours as
3 a fraction of the entire day (10 m³ during work/20 m³ for entire day). For the TCE ppm in air
4 converted from urinary metabolite data using Ikeda et al. (1972), the work week was 6 days, so
5 the adjustment for number of work days is 6/7.

7 **F.2.3. Physiologically Based Pharmacokinetic (PBPK) Model-Based Internal Dose Metrics**

8 PBPK modeling was used to estimate levels of dose metrics corresponding to different
9 exposure scenarios in rodents and humans (see Section 3.5). The selection of dose metrics for
10 specific organs and endpoints is discussed under Section 5.1.

11 The PBPK model requires an average body weight. For most of the studies, averages
12 specific to each species, strain, and sex were used. Where these were not reported in the text of
13 an article, data were obtained by digitizing the body weight graphics (Maltoni et al., 1986) or by
14 finding the median of weekly averages from graphs (National Cancer Institute [NCI], 1976;
15 National Toxicology Program [NTP], 1990, 1988). Where necessary, default adult body weights
16 specific to the strain were used (U.S. EPA, 1994).

17 **F.3. DOSE-RESPONSE MODELING PROCEDURES**

18 Where adequate dose-response data were available, models were fitted with the
19 BenchMark Dose Software (BMDS) (<http://www.epa.gov/ncea/bmbs>) using the applicable
20 applied doses or PBPK model-based dose metrics for each combination of study, species, strain,
21 sex, endpoints, and benchmark response (BMR) under consideration.

22 **F.3.1. Models for Dichotomous Response Data**

23 **F.3.1.1. *Quantal Models***

24 For dichotomous responses, the log-logistic, multistage, and Weibull models were fitted.
25 These models adequately describe the dose-response relationship for the great majority of data
26 sets, specifically in past TCE studies (Filipsson and Victorin, 2003). If the slope parameter of
27 the log-logistic model was less than 1, indicating a supralinear dose-response shape, the model
28 with the slope constrained to 1 was also fitted for comparison. For the multistage model, an
29 order one less than the number of dose groups was used, in addition to the 2nd-order multistage
30 model if it differed from the preceding model, and the first-order ('linear') multistage model
31 (which is identical to a Weibull model with power parameter equal to 1). The Weibull model
32 with the power parameter unconstrained was also fitted t.
33
34
35

This document is a draft for review purposes only and does not constitute Agency policy.

1 **F.3.1.2. *Nested Dichotomous Models***

2 In addition, nested dichotomous models were used for developmental effects in rodent
3 studies to account for possible litter effects, such maternal covariates or intralitter correlation.
4 The available nested models in BMDS are the nested log-logistic model, the Rai-VanRyzin
5 models, and the NCTR model. Candidates for litter-specific covariates (LSC) were identified
6 from the studies and considered legitimate for analysis if they were not significantly dose-related
7 (determined via regression, analysis of variance). The need for a LSC was indicated by a
8 difference of at least 3 in the Akaike Information Criteria (AIC) for models with and without a
9 LSC. The need to estimate intralitter correlations (IC) was determined by presence of a high
10 correlation coefficient for at least one dose group and by AIC. The fits for nested models were
11 also compared with the results from quantal models.

12 13 **F.3.2. *Models for Continuous Response Data***

14 For continuous responses, the distinct models available in BMDS were fitted: power
15 model (power parameter unconstrained and constrained to ≥ 1), polynomial model, and Hill
16 model. Both constant variance and modeled variance models were fit; but constant variance
17 models were used for model parsimony unless the p -value for the test of homogenous variance
18 was < 0.10 , in which case the modeled variance models were considered. For the polynomial
19 model, model order was selected as follows. A model of order 1 was fitted first. The next higher
20 order model (up to order $n-1$) was accepted if AIC decreased more than 3 units and the p -value
21 for the mean did not decrease.

22 23 **F.3.3. *Model Selection***

24 After fitting these models to the data sets, the recommendations for model selection set
25 out in U.S. Environmental Protection Agency (U.S. EPA)'s *Benchmark Dose Technical*
26 *Guidance Document* (Inter-Agency Review Draft, U.S. EPA, 2008b) were applied. First, models
27 were generally rejected if the p -value for goodness of fit was < 0.10 . In a few cases in which
28 none of the models fit the data with $p > 0.10$, linear models were selected on the basis of an
29 adequate visual fit overall. Second, models were rejected if they did not appear to adequately fit
30 the low-dose region of the dose-response relationship, based on an examination of graphical
31 displays of the data and scaled residuals. If the benchmark dose lower bound (BMDL) estimates
32 from the remaining models were "sufficiently close" (a criterion of within 2-fold for "sufficiently
33 close" was used), then the model with the lowest AIC was selected. The AIC is a measure of
34 information loss from a dose-response model that can be used to compare a set of models.
35 Among a specified set of models, the model with the lowest AIC is considered the "best." If two

1 or more models share the lowest AIC, the BMD *Technical Guidance Document* (U.S. EPA,
2 2008b) suggests that an average of the BMDLs could be used, but averaging was not used in this
3 assessment (for the one occasion in which models shared the lowest AIC, a selection was made
4 based on visual fit). If the BMDL estimates from the remaining models are not sufficiently
5 close, some model dependence is assumed. With no clear biological or statistical basis to choose
6 among them, the lowest BMDL was chosen as a reasonable conservative estimate, as suggested
7 in the *Benchmark Dose Technical Guidance Document*, unless the lowest BMDL appeared to be
8 an outlier, in which case further judgments were made.

10 **F.3.4. Additional Adjustments for Selected Data Sets**

11 In a few cases, the dose-response data necessitated further adjustments in order to
12 improve model fits.

13 The behavioral/neurological endpoint “number of rears” from Moser et al. (1995)
14 consisted of counts, measured at five doses and four measurement times (with eight observations
15 each). The high dose for this endpoint was dropped because the mean was zero, and no
16 monotone model could fit that well. Analysis of means and standard deviations for these counts
17 suggested a Box-Cox power transform (Box et al., 1978) of $\frac{1}{2}$ (i.e., square root) to stabilize
18 variances (i.e., the slope of the regression of $\log[\text{standard deviation (SD)}]$ on $\log[\text{mean}]$ was
19 0.46, and the relation was linear and highly significant). This information was helpful in
20 selecting a suitable variance model with high confidence (i.e., variance constant, for square-root
21 transformed data). Thus, the square root was taken of the original individual count data, and the
22 mean and variance of the transformed count data were used in the BMD modeling.

23 The high-dose group was dropped due to supra-linear dose-response shapes in two cases:
24 fetal cardiac malformations from Johnson et al. (2003) and decreased PFC response from
25 Woolhiser et al. (2006). Johnson et al. (2003) is discussed in more detail below (see
26 Section F.4.2.1). For Woolhiser et al. (2006), model fit near the BMD and the lower doses as
27 well as the model fit to the variance were improved by dropping the highest dose (a procedure
28 suggested in U.S. EPA (2008b)).

29 In some cases, the supralinear dose-response shape could not be accommodated by these
30 measures, and a LOAEL or no-observed-adverse-effect level (NOAEL) was used instead. These
31 include NCI (1976) (toxic nephrosis, >90% response at lowest dose), Keil et al. (2009)
32 (autoimmune markers and decreased thymus weight, only two dose groups in addition to
33 controls), and Peden-Adams et al. (2006) (developmental immunotoxicity, only two dose groups
34 in addition to controls).

This document is a draft for review purposes only and does not constitute Agency policy.

1 **F.4. DOSE-RESPONSE MODELING RESULTS**

2 **F.4.1. Quantal Dichotomous and Continuous Modeling Results**

3 The documents Appendix.linked.files\AppF.Non-cancer.Plots.TCE.contin.DRAFT.pdf
4 and Appendix.linked.files\AppF.Non-cancer.Plots.TCE.dichot.DRAFT.pdf show the fitted
5 model curves. The graphics include observations (group means or proportions), the estimated
6 model curve (solid red line) and estimated BMD, with a BMDL. Vertical bars show 95%
7 confidence intervals for the observed means. Printed above each plot are some key statistics
8 (necessarily rounded) for model goodness of fit and estimated parameters. Printed in the plots in
9 the upper left are the BMD and BMDL for the rodent data, in the same units as the rodent dose.

10 More detailed results, including alternative BMRs, alternative dose metrics, quantal
11 analyses for endpoints for which nested analyses were performed, etc. are documented in the
12 several spreadsheets. Input data for the analyses are in the following documents:
13 Appendix.linked.files\AppF.Non-cancer.Input.Data.TCE.contin.DRAFT.pdf and
14 Appendix.linked.files\AppF.Non-cancer.Input.Data.TCE.dichot.DRAFT.pdf. The documents
15 Appendix.linked.files\AppF.Non-cancer.Results.TCE.contin.DRAFT.pdf and
16 Appendix.linked.files\AppF.Non-cancer.Results.TCE.dichot.DRAFT.pdf present the data and
17 model summary statistics, including goodness-of-fit measures (Chi-square goodness-of-fit
18 *p*-value, AIC), parameter estimates, BMD, and BMDL. The group numbers “GRP” are arbitrary
19 and are the same as GRP in the plots. Finally, note that not all plots are shown in the documents
20 above, since these spreadsheets include many “alternative” analyses.

21

22 **F.4.2. Nested Dichotomous Modeling Results**

23 **F.4.2.1. Johnson *et al.* (2003) Fetal Cardiac Defects**

24 **F.4.2.1.1. Results using applied dose.** The biological endpoint was frequency of rat fetuses
25 having cardiac defects, as shown in Table F-4. Individual animal data were kindly provided by
26 Dr. Johnson (personal communication from Paula Johnson, University of Arizona, to Susan
27 Makris, U.S. EPA, 26 August 2009). Cochran-Armitage trend tests using number of fetuses and
28 number of litters indicated significant increases in response with dose (with or without including
29 the highest dose).

30 One suitable candidate for a LSC was available: female weight gain during pregnancy.
31 Based on goodness of fit, this covariate did not contribute to better fit and was not used. Some
32 ICs were significant and these parameters were included in the model.

This document is a draft for review purposes only and does not constitute Agency policy.

1 **Table F-4. Data on fetuses and litters with abnormal hearts from Johnson et**
 2 **al. (2003)**
 3

Dose group (mg/kg/d):	0	0.00045	0.048	0.218	129
Fetuses					
Number of pups:	606	144	110	181	105
Abnormal heart:	13	0	5	9	11
Litters					
Number of litters:	55	12	9	13	9
Abnormal heart:	9	0	4	5	6

4
 5
 6 With the high dose included, the chi-square goodness of fit was acceptable, but some
 7 residuals were large (1.5 to 2) for the control and two lower doses. Therefore, models were also
 8 fitted after dropping the highest dose. For these, goodness of fit was adequate, and scaled
 9 residuals were smaller for the low doses and control. Predicted expected response values were
 10 closer to observed when the high dose was dropped, as shown in Table F-5:
 11

12 **Table F-5. Comparison of observed and predicted numbers of fetuses with**
 13 **abnormal hearts from Johnson et al. (2003), with and without the high-dose**
 14 **group, using a nested model**
 15

Dose group (mg/kg/d):	Abnormal hearts (pups)				
	0	0.00045	0.048	0.218	129
Observed:	13	0	5	9	11
Predicted expected:					
With high dose	19.3	4.5	3.5	5.7	11
Without high dose	13.9	3.3	3.4	10	--

16
 17
 18 Accuracy in the low-dose range is especially important because the BMD is based upon
 19 the predicted responses at the control and the lower doses. Based on the foregoing measures of
 20 goodness of fit, the model based on dropping the high dose was used.

1 The nested log-logistic and Rai-VanRyzin models were fitted; these gave essentially the
 2 same predicted responses and POD. The former model was used as the basis for a POD; results
 3 are in Table F-6 and Figure F-2.

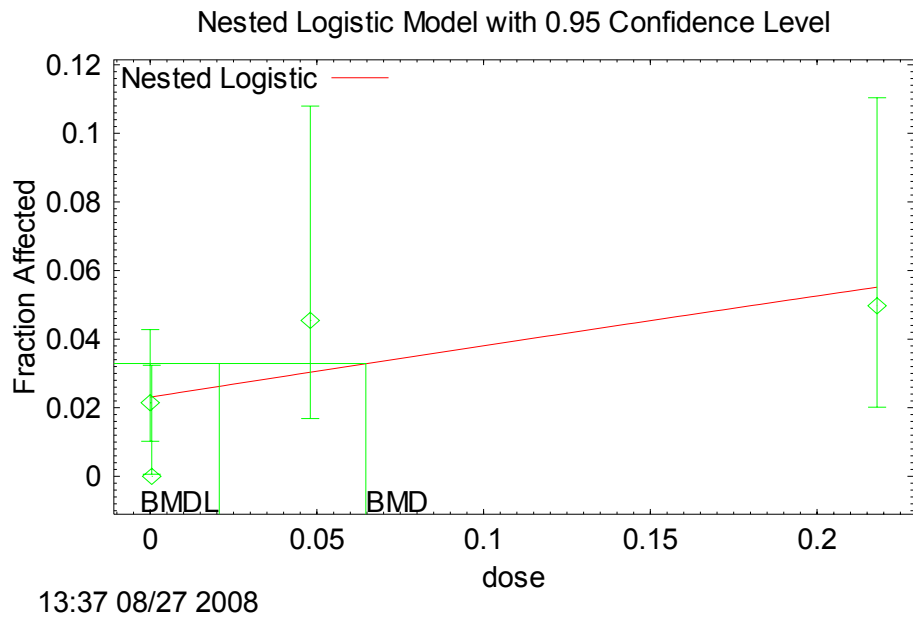
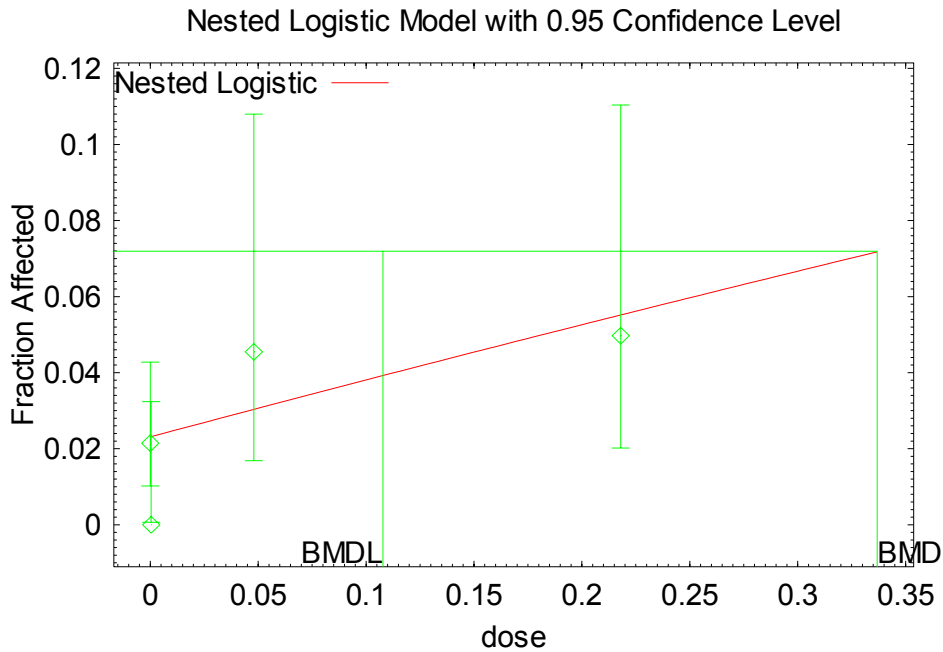
4
 5 **Table F-6. Results of nested log-logistic model for fetal cardiac anomalies**
 6 **from Johnson et al. (2003) without the high-dose group, on the basis of**
 7 **applied dose (mg/kg/d in drinking water)**
 8

Model	LSC?	IC?	AIC	Pval	BMR	BMD	BMDL
NLOG	Y	Y	246.877	NA (df = 0)	0.01	0.252433	0.03776
NLOG	Y	N	251.203	0.0112	0.01	0.238776	0.039285
NLOG	N	N	248.853	0.0098	0.01	0.057807	0.028977
NLOG	N	Y	243.815	0.0128	0.1	0.71114	0.227675
NLOG	N	Y	243.815	0.0128	0.05	0.336856	0.107846
NLOG*	N	Y	243.815	0.0128	0.01	0.064649	0.020698

9
 10 * Indicates model selected (Rai-VanRyzin model fits are essentially the same).

11 NLOG = “nested log-logistic” model.

12 LSC analyzed was female weight gain during pregnancy.
 13



1
2
3
4
5
6
7

Figure F-2. BMD modeling of Johnson et al. (2003) using nested log-logistic model, with applied dose, without LSC, with IC, and without the high-dose group, using a BMR of 0.05 extra risk (top panel) or 0.01 extra risk (bottom panel).

1 **F.4.2.1.2. Chi-square Goodness of Fit Test for nested log-logistic.** The BMDS choice of
2 subgroups did not seem appropriate given the data. The high-dose group of 13 litters was
3 subdivided into three subgroups having sums of expected counts 3, 3, and 2. However, the
4 control group of 55 litters could have been subdivided because expected response rates for
5 controls were relatively high. There was also concern that the goodness of fit might change with
6 alternative choices of subgroupings.

7 An R program was written to read the BMDS output, reading parameters and the table of
8 litter-specific results (dose, covariate, estimated probability of response, litter size, expected
9 response count, observed response count, scaled chi-square residual). The control group of
10 55 litters was subdivided into three subgroups of 18, 18, and 19 litters. Control litters were
11 sampled randomly without replacement 100 times, each time creating 3 subgroups—i.e.,
12 100 random assignments of the 55 control litters to three subgroups were made. For each of
13 these, the goodness-of-fit calculation was made and the *p*-value saved. Within these
14 100 *p*-values, $\geq 75\%$ were ≥ 0.05 , and $\geq 50\%$ had *p*-values ≥ 0.11 , this indicated that the model is
15 acceptable based on goodness-of-fit criteria.

16
17 **F.4.2.1.3. Results using physiologically based pharmacokinetic (PBPK) model-based dose**
18 **metrics.** The nested log-logistic model was also run using the dose metrics in the dams of total
19 oxidative metabolism scaled by body weight to the $3/4$ -power (TotOxMetabBW^{3/4}) and the area-
20 under-the-curve of TCE in blood (AUCCBld). As with the applied dose modeling, LSC
21 (maternal weight gain) was not included, but IC was included, based on the criteria outlined
22 previously (see Section F.3.1.2). The results are summarized in Table F-7 and Figure F-3 for
23 TotOxMetabBW^{3/4} and Table F-8 and Figure F-4 for AUCCBld.

24 25 **F.4.2.2. Narotsky et al. (1995)**

26 Data were combined for the high doses in the single-agent experiment and the lower
27 doses in the ‘five-cube’ experiment. Individual animal data were kindly provided by Dr.
28 Narotsky (personal communications from Michael Narotsky, U.S. EPA, to John Fox, U.S. EPA,
29 19 June 2008, and to Jennifer Jinot, U.S. EPA, 10 June 2008). Two endpoints were examined:
30 frequency of eye defects in rat pups and prenatal loss (number of implantation sites minus
31 number of live pups on postnatal day 1).

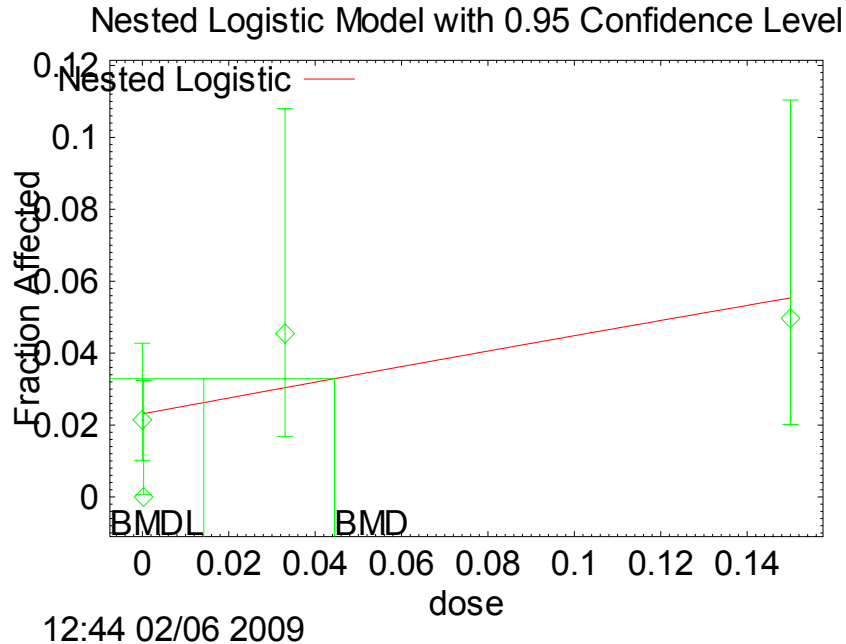
1
2
3
4

Table F-7. Results of nested log-logistic model for fetal cardiac anomalies from Johnson et al. (2003) without the high-dose group, using the TotOxMetabBW34 dose metric

Model	LSC?	IC?	AIC	Pval	BMR	BMD	BMDL
NLOG	Y	Y	246.877	NA (df = 0)	0.01	0.174253	0.0259884
NLOG	Y	N	251.203	0.0112	0.01	0.164902	0.0270378
NLOG	N	Y	243.815	0.0128	0.1	0.489442	0.156698
NLOG*	N	Y	243.815	0.0128	0.01	0.0444948	0.0142453
NLOG	N	N	248.853	0.0098	0.01	0.0397876	0.0199438

5
6
7
8
9
10
11

* Indicates model selected. BMDs failed with the Rai-VanRyzin and NCTR models.
 NLOG = “nested log-logistic” model.
 LSC analyzed was female weight gain during pregnancy.



12
13
14
15

Figure F-3. BMD modeling of Johnson et al. (2003) using nested log-logistic model, with TotOxMetabBW34 dose metric, without LSC, with IC, and without the high-dose group, using a BMR of 0.01 extra risk.

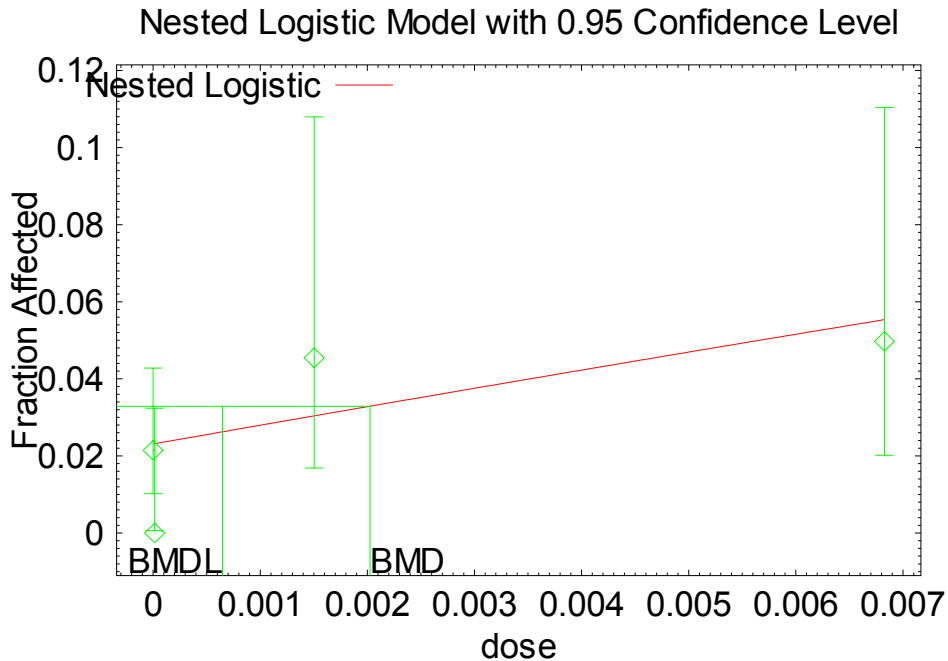
1 **Table F-8. Results of nested log-logistic model for fetal cardiac anomalies**
 2 **from Johnson et al. (2003) without the high-dose group, using the AUCCBld**
 3 **dose metric**
 4

Model	LSC?	IC?	AIC	Pval	BMR	BMD	BMDL
NLOG	Y	Y	246.877	NA (df = 0)	0.01	0.00793783	0.00118286
NLOG	Y	N	251.203	0.0112	0.01	0.00750874	0.00123047
NLOG*	N	Y	243.816	0.0128	0.1	0.0222789	0.00712997
NLOG*	N	Y	243.816	0.0128	0.01	0.00202535	0.000648179
NLOG	N	N	248.853	0.0098	0.01	0.00181058	0.000907513

5 * Indicates model selected. BMDs failed with the Rai-VanRyzin and NCTR models.
 6

7 NLOG = “nested log-logistic” model.
 8

9 LSC analyzed was female weight gain during pregnancy.
 10
 11



12 **Figure F-4. BMD modeling of Johnson et al. (2003) using nested log-logistic**
 13 **model, with AUCCBld dose metric, without LSC, with IC, and without the**
 14 **high-dose group, using a BMR of 0.01 extra risk.**
 15

This document is a draft for review purposes only and does not constitute Agency policy.

1 Two LSCs were considered, with analyses summarized in Table F-9. The number of implants is
 2 unrelated to dose, as inferred from regression and analysis of variance, and was considered as a
 3 LSC for eye defects. As number of implants is part of the definition for the endpoint of prenatal
 4 loss, it is not considered as a LSC for prenatal loss. A second LSC, the dam body weight on
 5 gestation day (GD) 6 (damBW6) was significantly related to dose and is unsuitable as a litter-
 6 specific covariate.

7
 8
 9

Table F-9. Analysis of LSCs with respect to dose from Narotsky et al. (1995)

Relation of litter-specific covariates to dose			
Implants:	none		
damBW6:	significant		
		Mean	Mean
	TCE	Implants	damBW6
	0	9.5	176.0
	10.1	10.1	180.9
	32	9.1	174.9
	101	7.8	170.1
	320	10.4	174.5
	475	9.7	182.4
	633	9.6	185.3
	844	8.9	182.9
	1,125	9.6	184.2
Using expt as covariate, e.g., $\text{damBW6} \sim \text{TCE.mg.kgd} + \text{expt}$			
Linear regression		$p = 0.7486$	$p = 0.0069$
AoV (ordered factor)		$p = 0.1782$	$p = 0.0927$

10
 11
 12
 13
 14
 15
 16
 17

Two LSCs were considered, with analyses summarized in Table F-9. The number of implants is unrelated to dose, as inferred from regression and analysis of variance, and was considered as a LSC for eye defects. As number of implants is part of the definition for the endpoint of prenatal loss, it is not considered as a LSC for prenatal loss. A second LSC, the dam body weight on GD 6 (damBW6) was significantly related to dose and is unsuitable as a litter-specific covariate.

This document is a draft for review purposes only and does not constitute Agency policy.

1 **F.4.2.2.1. Fetal eye defects.** The nested log-logistic and Rai-VanRyzin models were fitted to
 2 the number of pups with eye defects reported by Narotsky et al. (1995), with the results
 3 summarized in Table F-10.

4
 5 **Table F-10. Results of nested log-logistic and Rai-VanRyzin model for fetal**
 6 **eye defects from Narotsky et al. (1995), on the basis of applied dose (mg/kg/d**
 7 **in drinking water)**
 8

Model	LSC?	IC?	AIC	Pval	BMR	BMD	BMDL
NLOG	Y	Y	255.771	0.3489	0.05	875.347	737.328 ^a
NLOG	Y	N	259.024	0.0445	0.05	830.511	661.629
NLOG	N	Y	270.407	0.2281	0.05	622.342	206.460
NLOG	N	N	262.784	0.0529	0.10	691.93	542.101
NLOG	N	N	262.784	0.0529	0.05	427.389	264.386
NLOG	N	N	262.784	0.0529	0.01	147.41	38.7117 ^b
RAI	Y	Y	274.339	0.1047	0.05	619.849	309.925
RAI	Y	N	264.899	0.0577	0.05	404.788	354.961
RAI	N	Y	270.339	0.2309	0.05	619.882	309.941
RAI	N	N	262.481	0.0619	0.10	693.04	346.52
RAI	N	N	262.481	0.0619	0.05	429.686	214.843
RAI	N	N	262.481	0.0619	0.01	145.563	130.938 ^b

9
 10 ^a Graphical fit at the origin exceeds observed control and low dose responses and slope is quite flat (see Figure F-5),
 11 fitted curve does not represent the data well.

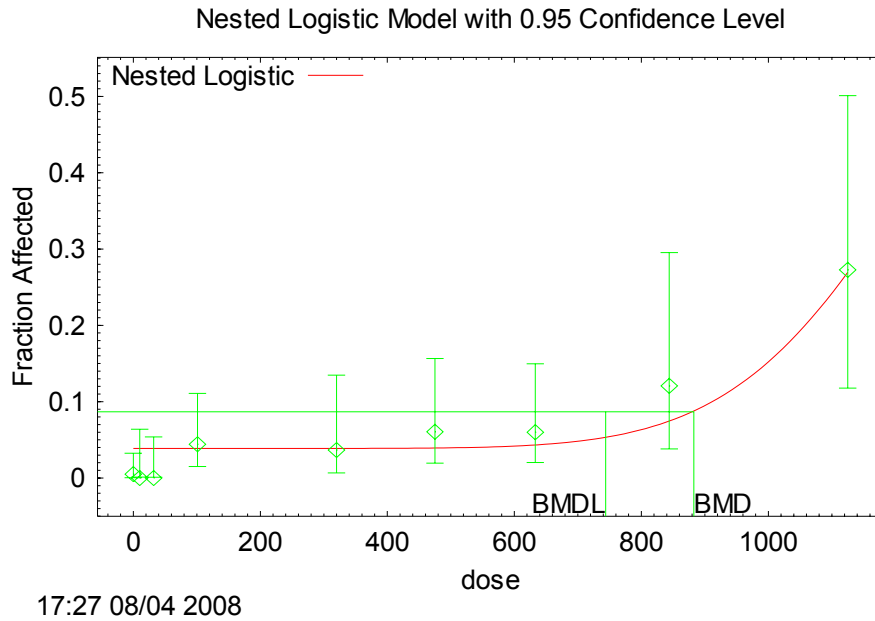
12 ^b Indicates model selected.

13
 14 NLOG = “nested log-logistic” model; RAI = Rai-VanRyzin model.
 15 LSC analyzed was implants.

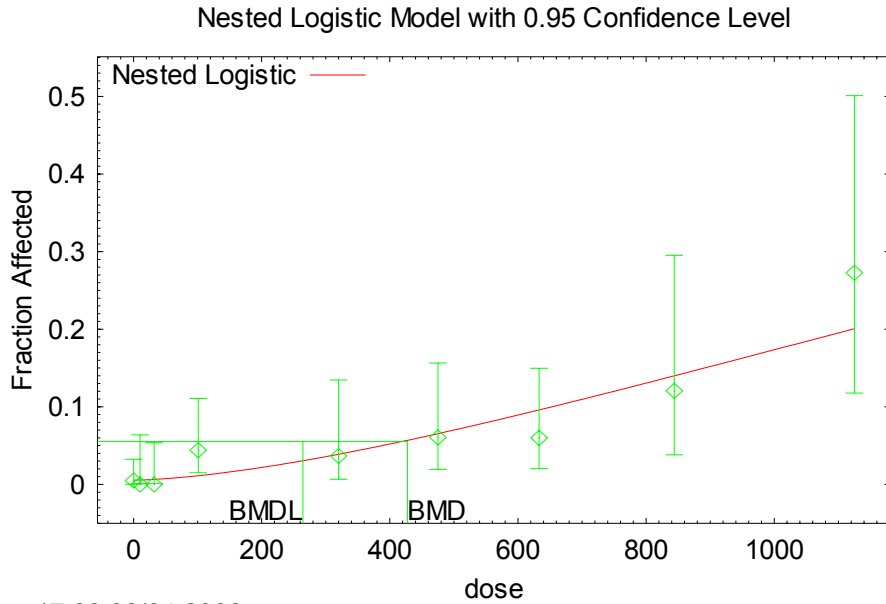
16
 17
 18 Results for the nested log-logistic model suggested a better model fit with the inclusion of
 19 the LSC and IC, based on AIC. However, the graphical fit (see Figure F-5) is strongly sublinear
 20 and high at the origin where the fitted response exceeds the observed low-dose responses for the
 21 control group and two low-dose groups. An alternative nested log-logistic model without either
 22 LSC or IC (see Figure F-6), which fits the low-dose responses better, was selected. Given that
 23 this model had no LSC and no IC, the nested log-logistic model reduces to a quantal log-logistic
 24 model. Parameter estimates and the *p*-values were essentially the same for the two models (see

This document is a draft for review purposes only and does not constitute Agency policy.

1 Table F-11). A similar model selection can be justified for the Rai-Van Ryzin model (see
2 Figure F-7). Because no LSC and no IC were needed, this endpoint was modeled with quantal
3 models, using totals of implants and losses for each dose group, which allowed choice from a
4 wider range of models (those results appear with quantal model results in this appendix).
5



6
7 **Figure F-5. BMD modeling of fetal eye defects from Narotsky et al. (1995)**
8 **using nested log-logistic model, with applied dose, with both LSC and IC,**
9 **using a BMR of 0.05 extra risk.**



1
2 **Figure F-6. BMD modeling of fetal eye defects from Narotsky et al. (1995)**
3 **using nested log-logistic model, with applied dose, without either LSC or IC,**
4 **using a BMR of 0.05 extra risk.**

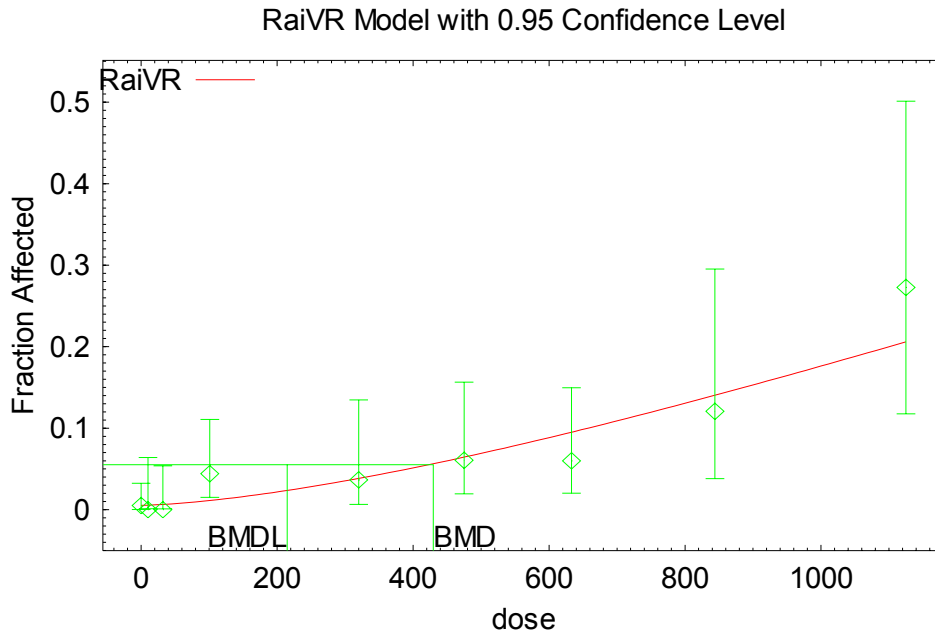
5
6
7 **Table F-11. Comparison of results of nested log-logistic (without LSC or IC)**
8 **and quantal log-logistic model for fetal eye defects from Narotsky et al.**
9 **(1995)**

10

Model	Parameter			BMD ₀₅	BMDL ₀₅
	Alpha	Beta	Rho		
Nested	0.00550062	-12.3392	1.55088	427.4	264.4
Quantal	0.00549976	-12.3386	1.55079	427.4	260.2

11
12

This document is a draft for review purposes only and does not constitute Agency policy.



17:25 08/04 2008

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15

Figure F-7. BMD modeling of fetal eye defects from Narotsky et al. (1995) using nested Rai-VanRyzin model, with applied dose, without either LSC or IC, using a BMR of 0.05 extra risk.

F.4.2.2.2. Narotsky et al. (1995) prenatal loss. The nested log-logistic and Rai-VanRyzin models were fitted to prenatal loss reported by Narotsky et al. (1995), with the results summarized in Table F-12.

The BMDS nested models require a LSC, so dam body weight on GD6 (“damBW6”) was used as the LSC. However, damBW6 is significantly related to dose and, so, is not a reliable LSC. Number of implants could not be used as a LSC because it was identified as number at risk in the BMDS models. These issues were obviated because the model selected did not employ the LSC.

1 **Table F-12. Results of nested log-logistic and Rai-VanRyzin model for**
 2 **prenatal loss from Narotsky et al. (1995), on the basis of applied dose**
 3 **(mg/kg/d in drinking water)**
 4

Model	LSC?	IC?	AIC	Pval	BMR	BMD	BMDL
NLOG	Y	Y	494.489	0.2314	0.10	799.723	539.094
NLOG	Y	N	627.341	0.0000	0.10	790.96	694.673
NLOG	N	N	628.158	0.0000	0.10	812.92	725.928
NLOG	N	Y	490.766	0.2509	0.10	814.781	572.057
NLOG	N	Y	490.766	0.2509	0.05	738.749	447.077
NLOG	N	Y	490.766	0.2509	0.01	594.995	252.437 *
RAI	Y	Y	491.859	0.3044	0.10	802.871	669.059
RAI	Y	N	626.776	0.0000	0.10	819.972	683.31
RAI	N	N	626.456	0.0000	0.10	814.98	424.469
RAI	N	Y	488.856	0.2983	0.10	814.048	678.373
RAI	N	Y	488.856	0.2983	0.05	726.882	605.735
RAI	N	Y	488.856	0.2983	0.01	562.455	468.713 *

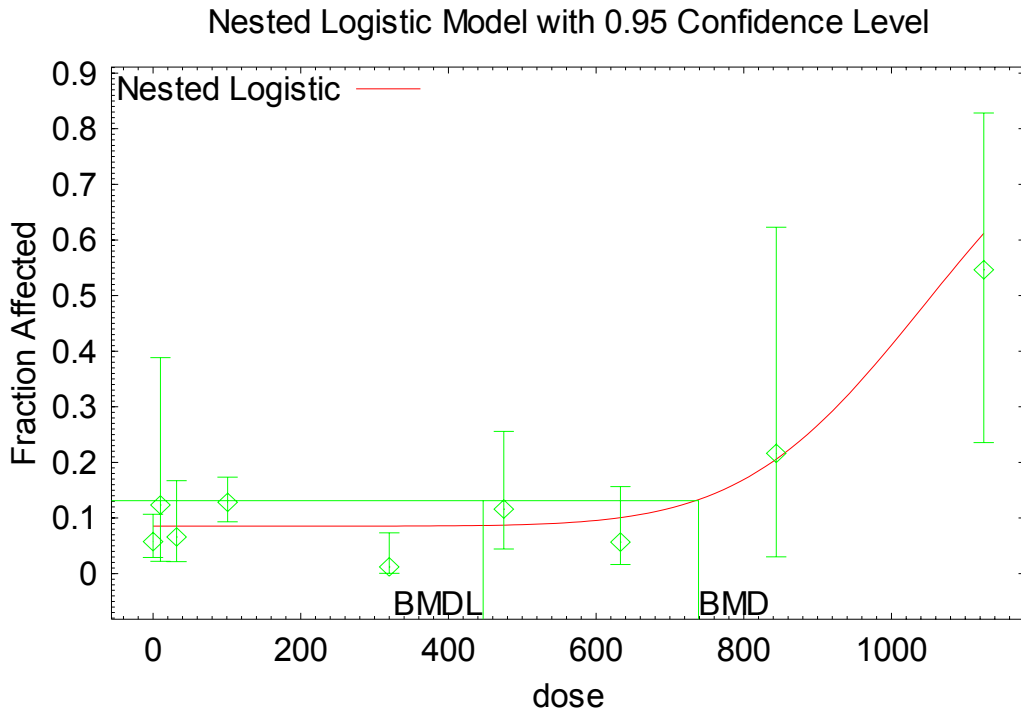
5 * Indicates model selected.

6
 7
 8 NLOG = “nested log-logistic” model; RAI = Rai-VanRyzin model.
 9 LSC analyzed was dam body weight on GD6.

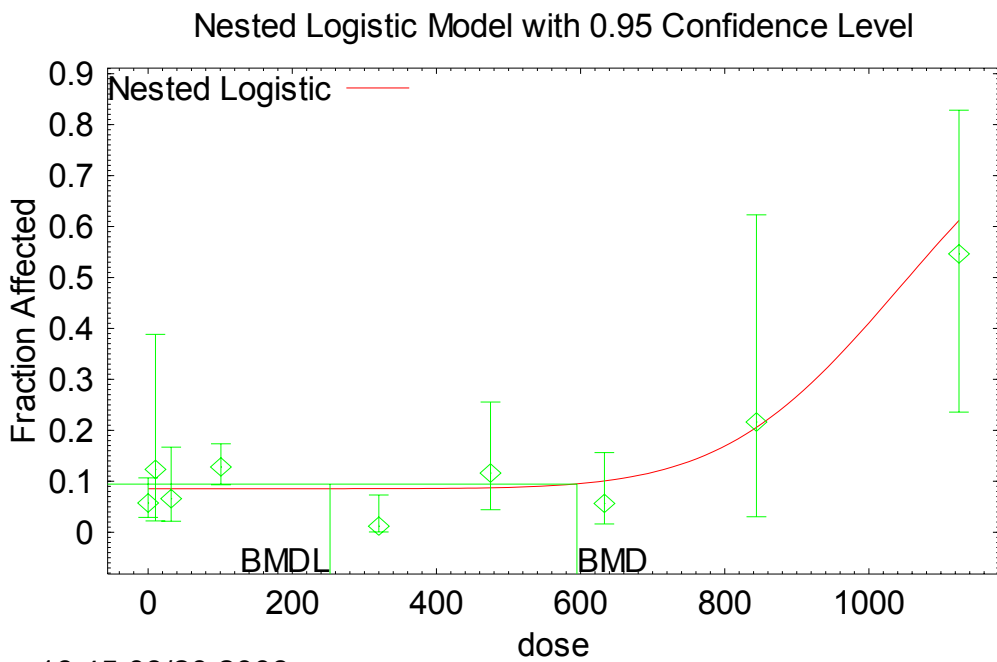
10
 11
 12 For the nested log-logistic models, the AIC is much larger when the IC is dropped, so the
 13 IC is needed in the model. The LSC can be dropped (and is also suspect because it is correlated
 14 with dose). The model with IC and without LSC was selected on the basis of AIC (shown in
 15 Figure F-8). For the Rai-VanRyzin models, the model selection was similar to that for the nested
 16 log-logistic, leading to a model with IC and without LSC, which had the lowest AIC (shown in
 17 Figure F-9).

18
 19 **F.4.3. Model Selections and Results**

20 The final model selections and results for noncancer dose-response modeling are
 21 presented in Table F-13.
 22

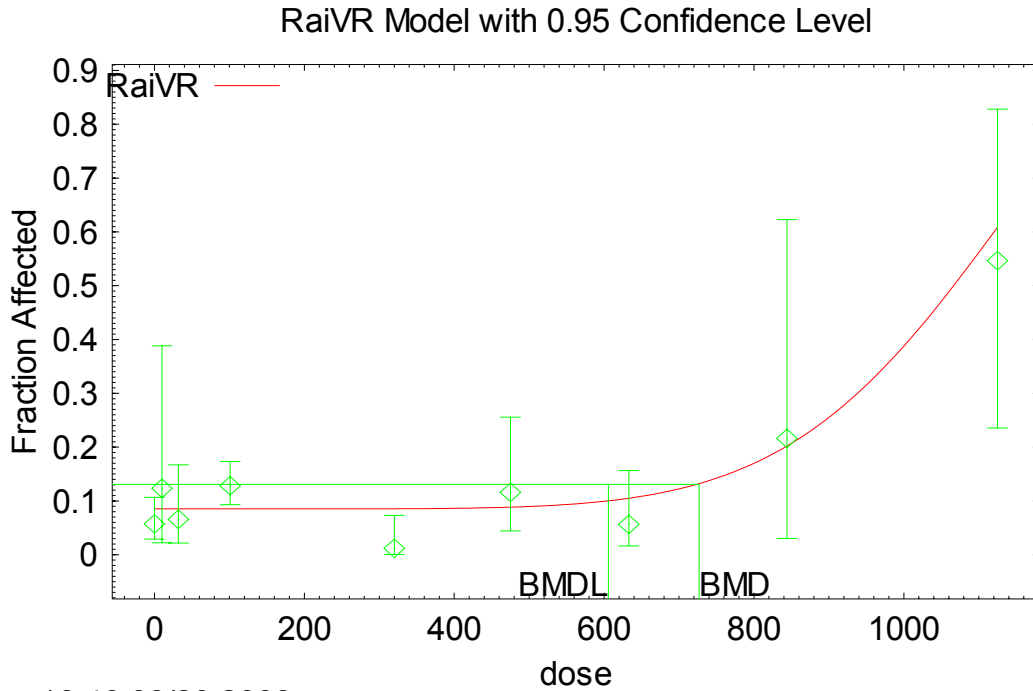


1
2

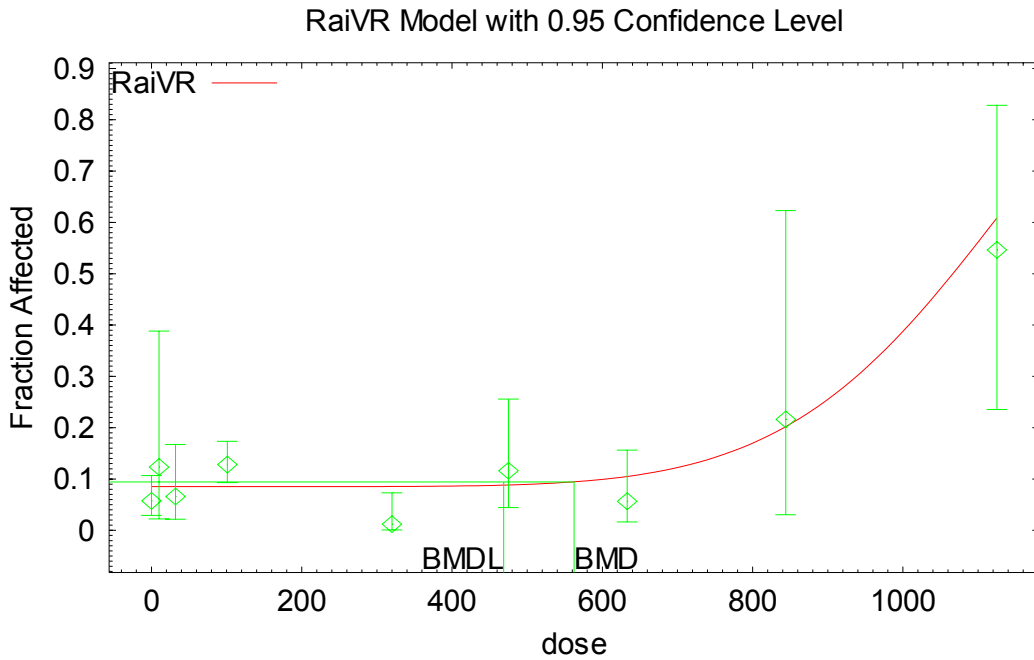


3
4
5
6

Figure F-8. BMD modeling of prenatal loss reported in Narotsky et al. (1995) using nested log-logistic model, with applied dose, without LSC, with IC, using a BMR of 0.05 extra risk (top panel) or 0.01 extra risk (bottom panel).



1



2

3

4

5

6

Figure F-9. BMD modeling of prenatal loss reported in Narotsky et al. (1995) using nested Rai-VanRyzin model, with applied dose, without LSC, with IC, using a BMR of 0.05 extra risk (top panel) or 0.01 extra risk (bottom panel).

Table F-13. Model selections and results for noncancer dose-response analyses

GRP	Study/run abbrev.	Species	Sex	Strain	Exp. route	Endpoint	Dose metric	BMR type	BMR	BMD/BMDL	BMDL	Model	Rep. BMD	Notes
Dichotomous models														
3	Chia et al., 1996	human	M	workers.elec.factory	inhal	N.hyperzoospermia	appl.dose	extra	0.1	2.14	1.43	loglogistic.1	3.06	
7	Narotsky et al., 1995	rat	F	F344	oral.gav	N.pups.eye.defects	appl.dose	extra	0.01	1.46	60.1	multistage	806	a
13	Narotsky et al., 1995.sa	rat	F	F344	oral.gav	N.dams.w.resorbed.litters	appl.dose	extra	0.01	5.47	32.2	multistage.2	570	
13	Narotsky et al., 1995.sa	rat	F	F344	oral.gav	N.dams.w.resorbed.litters	AUCCBld	extra	0.01	5.77	17.5	multistage.2	327	
13	Narotsky et al., 1995.sa	rat	F	F344	oral.gav	N.dams.w.resorbed.litters	TotMetabBW34	extra	0.01	1.77	77.5	weibull	156	
14	Johnson et al., 2003.drophi	rat	F	Sprague.Dawley	oral.dw	N.litters.abnormal.hearts	appl.dose	extra	0.1	2.78	0.0146	loglogistic.1	0.0406	b
36	Griffin et al., 2000	mice	F	MRL++	oral.dw	portal.infiltration	appl.dose	extra	0.1	2.67	13.4	loglogistic.1	35.8	
38	Maltoni et al., 1986	rat	M	Sprague.Dawley	inhal	megalonucleocytosis	appl.dose	extra	0.1	1.22	40.2	multistage	49.2	c
38	Maltoni et al., 1986	rat	M	Sprague.Dawley	inhal	megalonucleocytosis	ABioactDCVCBW34	extra	0.1	1.18	0.0888	loglogistic	0.105	
38	Maltoni et al., 1986	rat	M	Sprague.Dawley	inhal	megalonucleocytosis	AMetGSHBW34	extra	0.1	1.19	0.086	loglogistic	0.102	
38	Maltoni et al., 1986	rat	M	Sprague.Dawley	inhal	megalonucleocytosis	TotMetabBW34	extra	0.1	1.13	53.8	weibull	61	d
39	Maltoni et al., 1986	rat	M	Sprague.Dawley	oral.gav	megalonucleocytosis	appl.dose	extra	0.1	1.53	33.8	multistage.2	51.8	e
49	NTP, 1988	rat	F	Marshall	oral.gav	toxic nephropathy	appl.dose	extra	0.05	1.45	9.45	loglogistic.1	28.9	
49	NTP, 1988	rat	F	Marshall	oral.gav	toxic nephropathy	ABioactDCVCBW34	extra	0.05	1.45	0.0132	loglogistic.1	0.0404	
49	NTP, 1988	rat	F	Marshall	oral.gav	toxic nephropathy	AMetGSHBW34	extra	0.05	1.46	0.0129	loglogistic.1	0.0397	
49	NTP, 1988	rat	F	Marshall	oral.gav	toxic nephropathy	TotMetabBW34	extra	0.05	1.45	2.13	loglogistic.1	6.5	

10/20/09

This document is a draft for review purposes only and does not constitute Agency policy

F-23

DRAFT—DO NOT CITE OR QUOTE

Table F-13. Model selections and results for noncancer dose-response analyses (continued)

GRP	Study/run abbrev.	Species	Sex	Strain	Exp. route	Endpoint	Dose metric	BMR type	BMR	BMD/BMDL	BMDL	Model	Rep. BMD	Notes
Nested dichotomous models														
NA	Johnson et al., 2003.drophi	rat	F	Sprague.Dawley	oral.dw	N.pups.abnormal.hearts	appl.dose	extra	0.01	3.12	0.0207	loglogistic.IC	0.711	b
NA	Johnson et al., 2003.drophi	rat	F	Sprague.Dawley	oral.dw	N.pups.abnormal.hearts	TotOxMetabBW34	extra	0.01	3.12	0.0142	loglogistic.IC		b
NA	Johnson et al., 2003.drophi	rat	F	Sprague.Dawley	oral.dw	N.pups.abnormal.hearts	AUCCBid	extra	0.01	3.12	0.000648	loglogistic.IC		b
NA	Narotsky et al., 1995	rat	F	F344	oral.gav	N.prenatal.loss	appl.dose	extra	0.01	1.2	469	RAI.IC	814	
Continuous models														
2	Land et al., 1981	mouse	M	(C57B1xC3H)F1	inhal	pct.abnormal.sperm	appl.dose	standard	0.5	1.33	46.9	polynomial.constvar	125	
6	Carney et al., 2006	rat	F	Sprague-Dawley (CrI:CD)	inhal	gm.wgt.gain.GD6.9	appl.dose	relative	0.1	2.5	10.5	hill	62.3	
8	Narotsky et al., 1995	rat	F	F344	oral.gav	gm.wgt.gain.GD6.20	appl.dose	relative	0.1	1.11	108	polynomial.constvar	312	
19	Crofton and Zhao, 1997	rat	M	Long-Evans	inhal	dB.auditory.threshold(16kHz)	appl.dose	absolute	10	1.11	274	polynomial.constvar	330	
21	George et al., 1986	rat	F	F344	oral.food	litters	appl.dose	standard	0.5	1.69	179	polynomial.constvar	604	
23	George et al., 1986	rat	F	F344	oral.food	live.pups	appl.dose	standard	0.5	1.55	152	polynomial.constvar	470	
26	George et al., 1986	rat	F	F344	oral.food	Foffspring.BWgm.day21	appl.dose	relative	0.05	1.41	79.7	polynomial.constvar	225	
34sq	Moser et al., 1995+perscom	rat	F	F344	oral.gav	no.rears	appl.dose	standard	1	1.64	248	polynomial.constvar	406	b,f
49	George et al., 1986	rat	F	F344	oral.food	traverse.time.21do	appl.dose	relative	1	1.98	72.6	power	84.9	
51	Buben and O'Flaherty, 1985	mouse	M	SwissCox	oral.gav	Liverwt.pctBW	appl.dose	relative	0.1	1.26	81.5	hill.constvar	92.8	

Table F-13. Model selections and results for noncancer dose-response analyses (continued)

GRP	Study/run abbrev.	Species	Sex	Strain	Exp. route	Endpoint	Dose metric	BMR type	BMR	BMD/BMDL	BMDL	Model	Rep. BMD	Notes
51	Buben and O'Flaherty, 1985	mouse	M	SwissCox	oral.gav	Liverwt.pctBW	AMetLiv1BW34	relative	0.1	1.08	28.6	polynomial.constvar	28.4	
51	Buben and O'Flaherty, 1985	mouse	M	SwissCox	oral.gav	Liverwt.pctBW	TotOxMetabBW34	relative	0.1	1.08	37	polynomial.constvar	36.7	
58	Kjellstrand et al, 1983b	mouse	M	NMRI	inhal	Liverwt.pctBW	appl.dose	relative	0.1	1.36	21.6	hill	30.4	
58	Kjellstrand et al, 1983b	mouse	M	NMRI	inhal	Liverwt.pctBW	AMetLiv1BW34	relative	0.1	1.4	22.7	hill	32.9	
58	Kjellstrand et al, 1983b	mouse	M	NMRI	inhal	Liverwt.pctBW	TotOxMetabBW34	relative	0.1	1.3	73.4	hill	97.7	
60.Rp	Kjellstrand et al, 1983b	mouse	M	NMRI	inhal	Kidneywt.pctBW	appl.dose	relative	0.1	1.17	34.7	polynomial	47.1	
60.Rp	Kjellstrand et al, 1983b	mouse	M	NMRI	inhal	Kidneywt.pctBW	AMetGSHBW34	relative	0.1	1.18	0.17	polynomial	0.236	
60.Rp	Kjellstrand et al, 1983b	mouse	M	NMRI	inhal	Kidneywt.pctBW	TotMetabBW34	relative	0.1	1.17	71	polynomial	95.2	
63	Woolhiser et al, 2006	rat	F	CD (Sprague-Dawley)	inhal	Antibody.Forming Cells	appl.dose	standard	1	1.94	31.2	power.constvar	60.6	b
62	Woolhiser et al, 2006	rat	F	CD (Sprague-Dawley)	inhal	Antibody.Forming Cells	AUCCBid	standard	1	1.44	149	polynomial	214	
62	Woolhiser et al, 2006	rat	F	CD (Sprague-Dawley)	inhal	Antibody.Forming Cells	TotMetabBW34	standard	1	1.5	40.8	polynomial	61.3	
65	Woolhiser et al, 2006	rat	F	CD (Sprague-Dawley)	inhal	kidney.wt.per100gm	appl.dose	relative	0.1	4.29	15.7	hill.constvar	54.3	
65	Woolhiser et al, 2006	rat	F	CD (Sprague-Dawley)	inhal	kidney.wt.per100gm	ABioactDCVCBW34	relative	0.1	4.27	0.0309	hill.constvar	0.103	
65	Woolhiser et al, 2006	rat	F	CD (Sprague-Dawley)	inhal	kidney.wt.per100gm	AMetGSHBW34	relative	0.1	4.28	0.032	hill.constvar	0.107	
65	Woolhiser et al, 2006	rat	F	CD (Sprague-Dawley)	inhal	kidney.wt.per100gm	TotMetabBW34	relative	0.1	1.47	40.8	polynomial.constvar	52.3	
67	Woolhiser et al, 2006	rat	F	CD (Sprague-Dawley)	inhal	liver.wt.per100gm	appl.dose	relative	0.1	4.13	25.2	hill.constvar	70.3	
67	Woolhiser et al, 2006	rat	F	CD (Sprague-Dawley)	inhal	liver.wt.per100gm	AMetLiv1BW34	relative	0.1	1.53	46	polynomial.constvar	56.1	
67	Woolhiser et al, 2006	rat	F	CD (Sprague-Dawley)	inhal	liver.wt.per100gm	TotOxMetabBW34	relative	0.1	1.53	48.9	polynomial.constvar	59.8	

Table F-13. Model selections and results for noncancer dose-response analyses (continued)

^aEight-stage multistage model.

^bDropped highest dose.

^cThree-stage multistage model.

^dWeibull selected over log-logistic with the same AIC on basis of visual fit (less extreme curvature).

^eSecond-order MS selected on basis of visual fit (less extreme curvature).

^fSquare-root transformation of original individual count data.

Applied dose BMDLs are in units of ppm in air for inhalation exposures and mg/kg/d for oral exposures. Internal dose BMDLs are in dose metric units. Reporting BMD is BMD using a BMR of 0.1 extra risk for dichotomous models, and 1 control SD for continuous models.

Log-logistic = unconstrained log-logistic; log-logistic.1 = constrained log-logistic; multistage = multistage with #stages=dose groups-1; multistage.n = n-stage multistage; log-logistic.IC = nested log-logistic with IC, without LSC; RAI.IC = Rai-VanRyzin model with IC, without LSC; zzz.constvar = continuous model zzz with constant variance (otherwise variance is modeled).

Rep. = reporting, Exp. = exposure, Abbrev. = abbreviation.

10/20/09

This document is a draft for review purposes only and does not constitute Agency policy

F-26

DRAFT—DO NOT CITE OR QUOTE

1 **F.5. DERIVATION OF POINTS OF DEPARTURE**

2 **F.5.1. Applied Dose Points of Departure**

3 For oral studies in rodents, the POD on the basis of applied dose in mg/kg/d was taken to
4 be the BMDL, NOAEL, or LOAEL. NOAELs and LOAELs were adjusted for intermittent
5 exposure to their equivalent continuous average daily exposure (for BMDLs, the adjustments
6 were already performed prior to BMD modeling).

7 For inhalation studies in rodents, the POD on the basis of applied dose in ppm was taken
8 to be the BMDL, NOAEL, or LOAEL. NOAELs and LOAELs were adjusted for intermittent
9 exposure to their equivalent continuous average daily exposure (for BMDLs, the adjustments
10 were already performed prior to BMD modeling). These adjusted concentrations are considered
11 human equivalent concentrations, in accordance with U.S. EPA (1994), as TCE is considered a
12 Category 3 gas (systemically acting) and has a blood-air partition coefficient in rodents greater
13 than that in humans (see Section 3.1).

14

15 **F.5.2. Physiologically Based Pharmacokinetic (PBPK) Model-Based Human Points of**
16 **Departure**

17 As discussed in Section 5.1.3, the PBPK model was used for simultaneous interspecies
18 (for endpoints in rodent studies), intraspecies, and route-to-route extrapolation based on the
19 estimates from the PBPK model of the internal dose points of departure (idPOD) for each
20 candidate critical study/endpoints. The following documents contain figures showing the
21 derivation of the human equivalent doses and concentrations (human equivalent doses [HEDs]
22 and human equivalent concentrations [HECs]) for the median (50th percentile) and sensitive (99th
23 percentile) individual from the (rodent or human) study idPOD. In each case, for a specific
24 study/endpoint(s)/sex/species (in the figure main title), and for a particular dose metric (Y-axis
25 label), the horizontal line shows the original study idPOD (a BMDL, NOAEL, or LOAEL as
26 noted) and where it intersects with the human 99th percentile (open square) or median (closed
27 square) exposure-internal-dose relationship:

28 [Appendix.linked.files\AppF.Non-cancer.HECs.Plots.human.inhalation.studies.TCE.DRAFT.pdf](#)

29 [Appendix.linked.files\AppF.Non-cancer.HECs.Plots.rodent.inhalation.studies.TCE.DRAFT.pdf](#)

30 [Appendix.linked.files\AppF.Non-cancer.HECs.Plots.rodent.oral.studies.TCE.DRAFT.pdf](#)

31 [Appendix.linked.files\AppF.Non-cancer.HEDs.Plots.human.inhalation.studies.TCE.DRAFT.pdf](#)

32 [Appendix.linked.files\AppF.Non-cancer.HEDs.Plots.rodent.inhalation.studies.TCE.DRAFT.pdf](#)

33 [Appendix.linked.files\AppF.Non-cancer.HEDs.Plots.rodent.oral.studies.TCE.DRAFT.pdf](#)

34 The original study internal doses are based on the median estimates from about 2,000
35 “study groups” (for rodent studies) or “individuals” (for human studies), and corresponding

This document is a draft for review purposes only and does not constitute Agency policy.

1 exposures for the human median and 99th percentiles were derived from a distribution of 2,000
2 “individuals.” In both cases, the distributions reflect combined uncertainty (in the population
3 means and variances) and population variability.

4 In addition, as part of the uncertainty/variability analysis described in Section 5.1.4.2, the
5 POD for studies/endpoints for which BMD modeling was done was replaced by the LOAEL or
6 NOAEL. This was done to because there was no available tested software for performing BMD
7 modeling in such a context and because of limitations in time and resources to develop such
8 software. However, the relative degree of uncertainty/variability should be adequately captured
9 in the use of the LOAEL or NOAEL. The graphical depiction of the HEC₉₉ or HED₉₉ using
10 these alternative PODs is shown in the following files:

11 Appendix.linked.files\AppF.Non-
12 cancer.HECs.AltPOD.Plots.rodent.inhalation.studies.TCE.DRAFT.pdf

13 Appendix.linked.files\AppF.Non-
14 cancer.HECs.AltPOD.Plots.rodent.oral.studies.TCE.DRAFT.pdf

15 Appendix.linked.files\AppF.Non-
16 cancer.HEDs.AltPOD.Plots.rodent.inhalation.studies.TCE.DRAFT.pdf

17 Appendix.linked.files\AppF.Non-
18 cancer.HEDs.AltPOD.Plots.rodent.oral.studies.TCE.DRAFT.pdf.

20 **F.6. SUMMARY OF POINTS OF DEPARTURE (PODs) FOR CRITICAL STUDIES** 21 **AND EFFECTS SUPPORTING THE INHALATION REFERENCE CONCENTRATION** 22 **(RfC) AND ORAL REFERENCE DOSE (RfD)**

23 This section summarizes the selection and/or derivation of PODs from the critical studies
24 and effects that support the inhalation reference concentration (RfC) and oral reference dose
25 (RfD). In particular, for each endpoint, the following are described the dosimetry (adjustments
26 of continuous exposure, PBPK dose metrics), selection of BMR and BMD model (if BMD
27 modeling was performed), and derivation of the human equivalent concentration or dose for a
28 sensitive individual (if PBPK modeling was used). Section 5.1.3.1 discusses the dose metric
29 selection for different endpoints.

31 **F.6.1. National Toxicology Program (NTP, 1988)—Benchmark Dose (BMD) Modeling of** 32 **Toxic Nephropathy in Rats**

33 The critical endpoint here is toxic nephropathy in female Marshall rats (NTP, 1988),
34 which was the most sensitive sex/strain in this study, although the differences among different
35 sex/strain combinations was not large (BMDLs differed by ≤ 3 -fold).

This document is a draft for review purposes only and does not constitute Agency policy.

1 **F.6.1.1. Dosimetry and Benchmark Dose (BMD) Modeling**

2 Rats were exposed to 500 or 1,000 day, 5 days/week, for 104 weeks. The primary dose
3 metric was selected to be average amount of dichlorovinyl cysteine (DCVC)
4 bioactivated/kg^{3/4}/day, with median estimates from the PBPK model for the female Marshall rats
5 in this study of 0.47 and 1.1.

6 Figure F-10 shows BMD modeling for the dichotomous models used (see Section F.5.1,
7 above). The log-logistic model with slope constrained to ≥ 1 was selected because (1) the log-
8 logistic model with unconstrained slope yielded a slope estimate < 1 and (2) it had the lowest
9 AIC.

10 The idPOD of 0.0132 mg DCVC bioactivated/kg^{3/4}/day was a BMDL for a BMR of 5%
11 extra risk. This BMR was selected because toxic nephropathy is a clear toxic effect. This BMR
12 required substantial extrapolation below the observed responses (about 60%); however, the
13 response level seemed warranted for this type of effect and the ratio of the BMD to the BMDL
14 was not large (1.56 for the selected model).

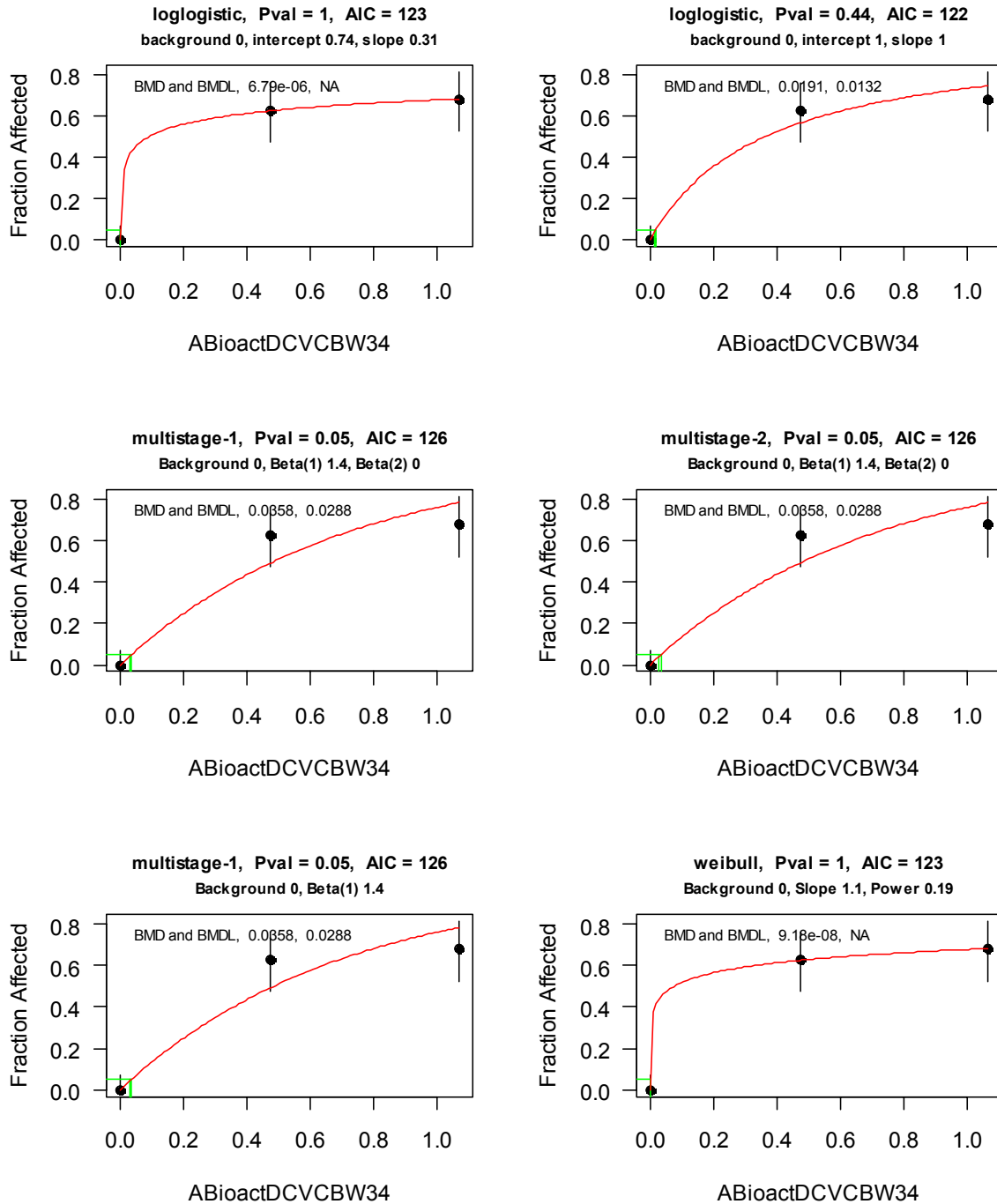
15
16 **F.6.1.2. Derivation of HEC₉₉ and HED₉₉**

17 The HEC₉₉ and HED₉₉ are the lower 99th percentiles for the continuous human exposure
18 concentration and continuous human ingestion dose that lead to a human internal dose equal to
19 the rodent idPOD. The derivation of the HEC₉₉ of 0.0056 ppm and HED₉₉ of 0.00338 mg/kg/d
20 for the 99th percentile for uncertainty and variability are shown in Figure F-11. These values are
21 used as this critical effect's POD to which additional uncertainty factors (UFs) are applied.

22
23 **F.6.2. National Cancer Institute (NCI, 1976)—Lowest-Observed-Adverse-Effect Level**
24 **(LOAEL) for Toxic Nephrosis in Mice**

25 The critical endpoint here is toxic nephrosis in female B6C3F1 mice (NCI, 1976), which
26 was the most sensitive sex in this study, although the LOAEL for males differed by less than
27 50%.

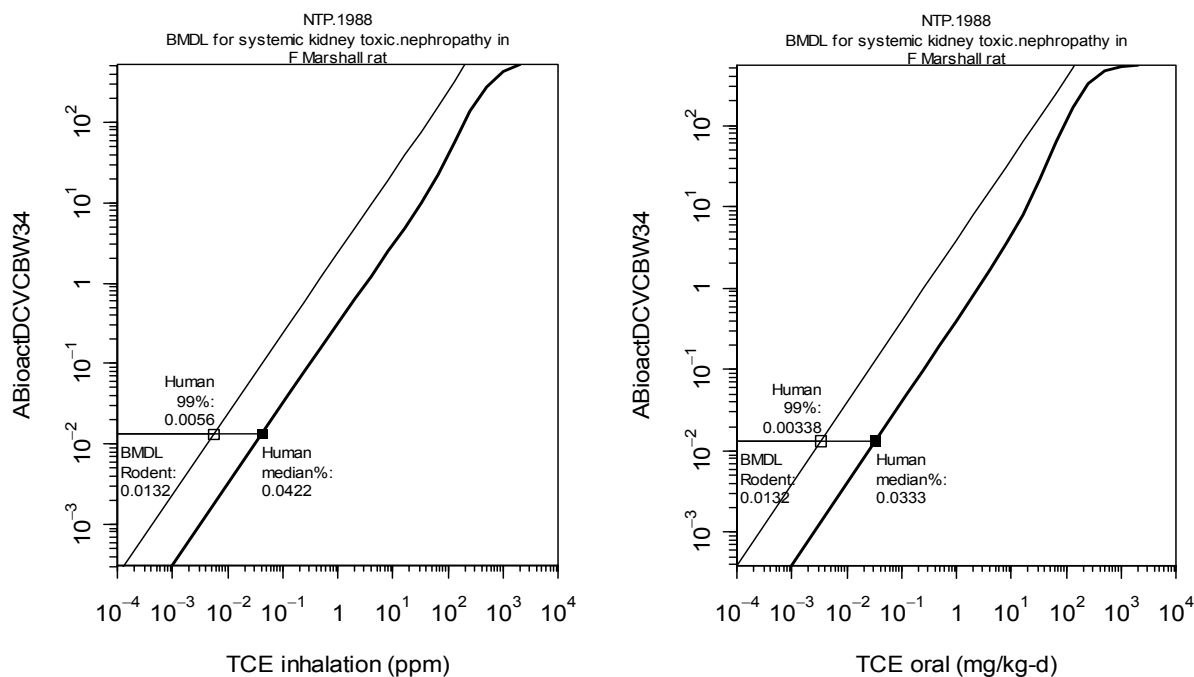
NTP.1988 kidney toxic nephropathy rat Marshall F oral.gav (GRP 49)
BMR: 0.05 extra



1
2
3
4

Figure F-10. BMD modeling of NTP (1988) toxic nephropathy in female Marshall rats.

This document is a draft for review purposes only and does not constitute Agency policy.



1
2
3 **Figure F-11. Derivation of HEC₉₉ and HED₉₉ corresponding to the rodent**
4 **idPOD from NTP (1988) toxic nephropathy in rats.**
5
6

7 **F.6.2.1. Dosimetry**

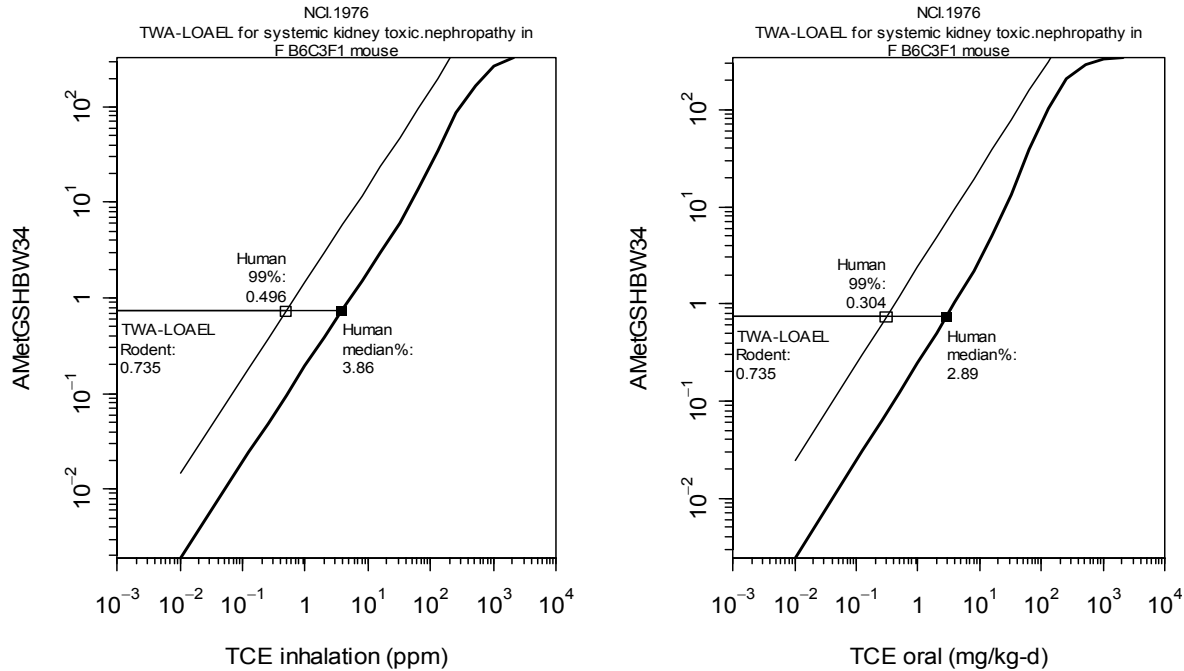
8 Mice were exposed to a time-weighted average of 869 and 1,739 mg/kg/d, 5 days/week,
9 for 78 weeks. BMD modeling was not performed because the response at the LOAEL was
10 >90%. The primary dose metric was selected to be average amount of TCE conjugated with
11 glutathione (GSH)/kg^{3/4}/d. In this study, the lower dose group was exposed to two different dose
12 levels (700 mg/kg/d for 12 weeks and 900 mg/kg/d for 66 weeks). The median estimates from
13 the PBPK model for the two dose levels were 0.583 and 0.762 mg TCE conjugation with
14 GSH/kg^{3/4}/d. Applying the same time-weighted averaging gives an idPOD LOAEL of 0.735 mg
15 TCE conjugation with GSH/kg^{3/4}/d.
16

17 **F.6.2.2. Derivation of HEC₉₉ and HED₉₉**

18 The HEC₉₉ and HED₉₉ are the lower 99th percentiles for the continuous human exposure
19 concentration and continuous human ingestion dose that lead to a human internal dose equal to
20 the rodent idPOD. The derivation of the HEC₉₉ of 0.50 ppm and HED₉₉ of 0.30 mg/kg/d for the

1 99th percentile for uncertainty and variability are shown in Figure F-12. These values are used as
2 this critical effect's POD to which additional UFs are applied.

3



4

5

6 **Figure F-12. Derivation of HEC₉₉ and HED₉₉ corresponding to the rodent**
7 **idPOD from NTP (1988) toxic nephrosis in mice.**

8

9

10 **F.6.3. Woolhiser et al. (2006)—Benchmark Dose (BMD) Modeling of Increased Kidney**
11 **Weight in Rats**

12 The critical endpoint here is increased kidney weights in female Sprague-Dawley (S-D)
13 rats (Woolhiser et al., 2006).

14

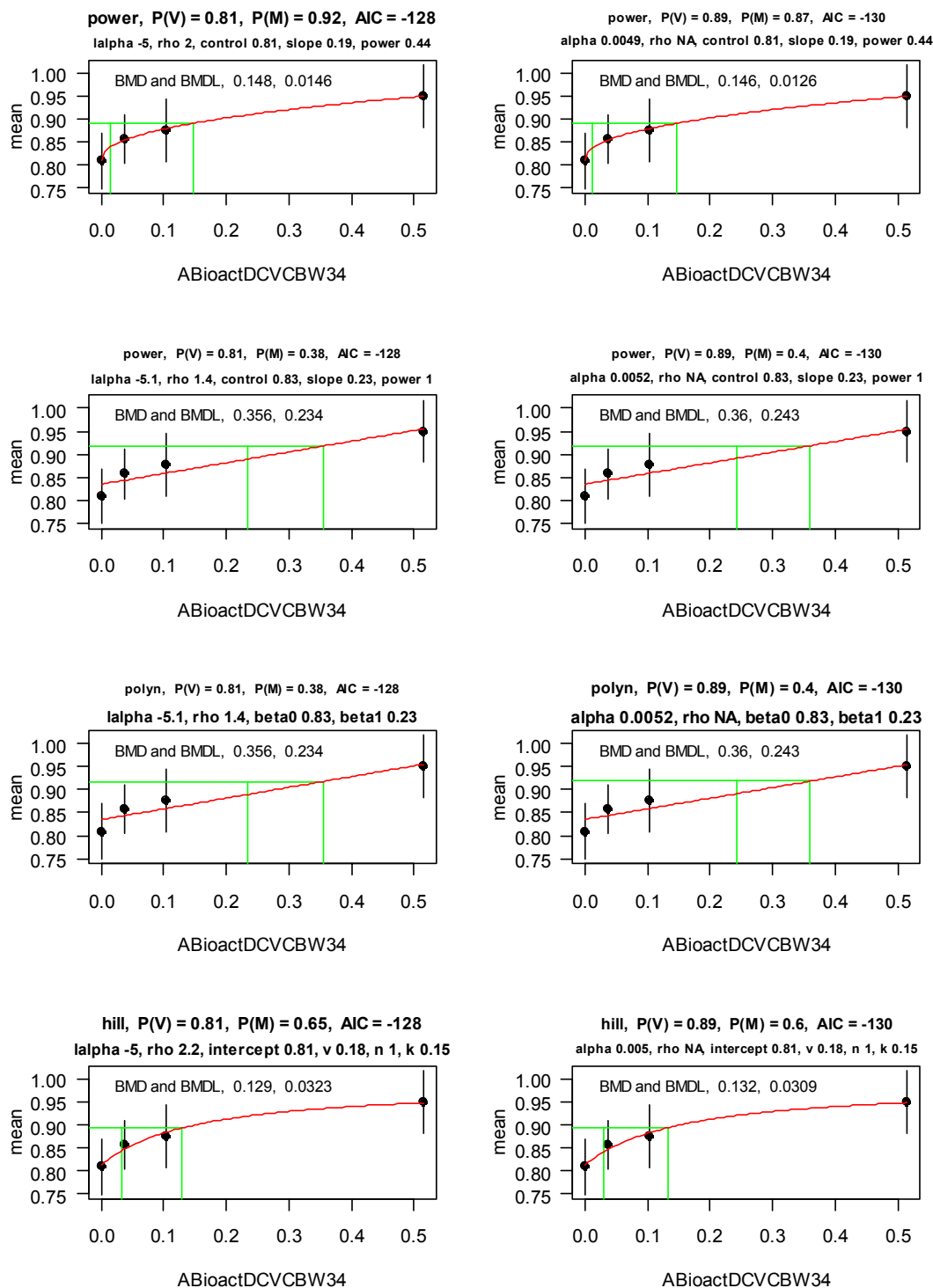
15 **F.6.3.1. Dosimetry and Benchmark Dose (BMD) Modeling**

16 Rats were exposed to 100, 300, and 1000, 6 hours/day, 5 days/week, for 4 weeks. The
17 primary dose metric was selected to be average amount of DCVC bioactivated/kg^{3/4}/day, with
18 median estimates from the PBPK model for this study of 0.038, 0.10, and 0.51.

19 Figure F-13 shows BMD modeling for the continuous models used (see Section F.5.2,
20 above). The Hill model with constant variance was selected because it had the lowest AIC and
21 because other models with the same AIC either were a power model with power parameter <1 or
22 had poor fits to the control data set.

This document is a draft for review purposes only and does not constitute Agency policy.

Woolhiser.etal.2006 Kidney kidney.wt.per100gm rat CD (Sprague-Dawley) F inhal (GRP 65)
 BMR: 0.1 relative



1
 2
 3

Figure F-13. BMD modeling of Woolhiser et al. (2006) for increased kidney weight in female S-D rats.

This document is a draft for review purposes only and does not constitute Agency policy.

1 The idPOD of 0.0309 mg DCVC bioactivated/kg^{3/4}/day was a BMDL for a BMR of 10%
2 weight change, which is the BMR typically used by U.S. EPA for body weight and organ weight
3 changes. The response used in each case was the organ weight as a percentage of body weight,
4 to account for any commensurate decreases in body weight, although the results did not differ
5 much when absolute weights were used instead.
6

7 **F.6.3.2. Derivation of HEC₉₉ and HED₉₉**

8 The HEC₉₉ and HED₉₉ are the lower 99th percentiles for the continuous human exposure
9 concentration and continuous human ingestion dose that lead to a human internal dose equal to
10 the rodent idPOD. The derivation of the HEC₉₉ of 0.0131 ppm and HED₉₉ of 0.00791 mg/kg/d
11 for the 99th percentile for uncertainty and variability are shown in Figure F-14. These values are
12 used as this critical effect's POD to which additional UFs are applied.
13

14 **F.6.4. Keil et al. (2009)—Lowest-Observed-Adverse-Effect Level (LOAEL) for Decreased 15 Thymus Weight and Increased Anti-dsDNA and Anti-ssDNA Antibodies in Mice**

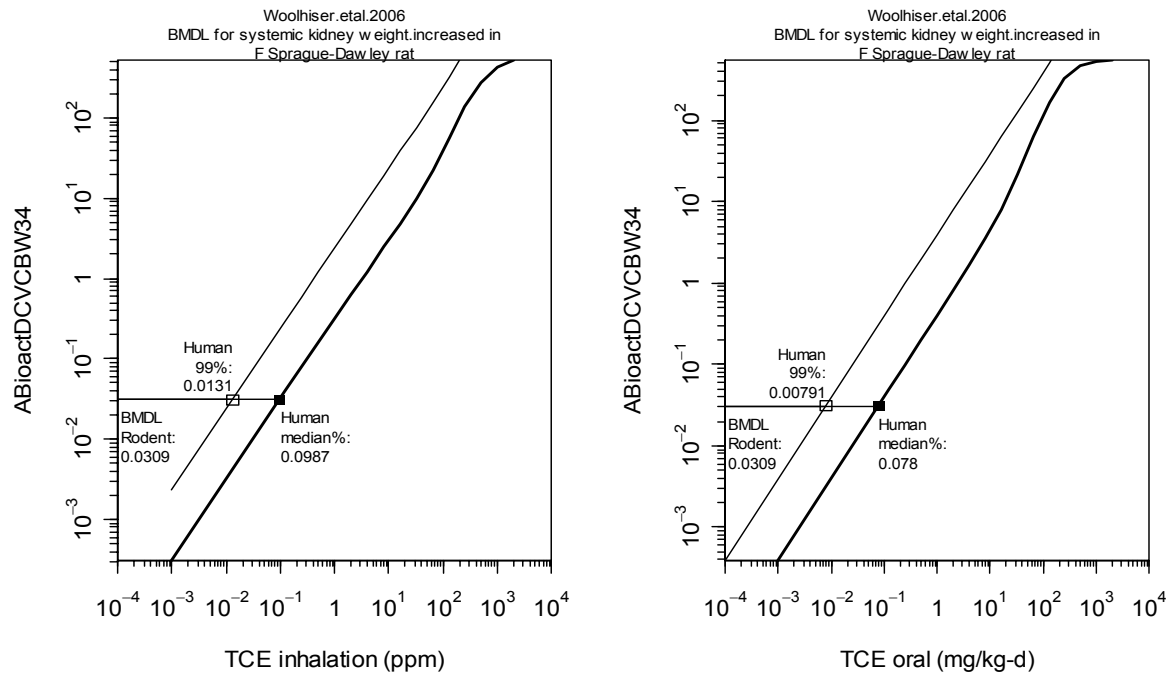
16 The critical endpoints here are decreased thymus weight and increased anti-dsDNA and
17 anti-ssDNA antibodies in female B6C3F1 mice (Keil et al., 2009).
18

19 **F.6.5. Keil et al. (2009)—Lowest-Observed-Adverse-Effect Level (LOAEL) for Decreased 20 Thymus Weight and Increased Anti-dsDNA and Anti-ssDNA Antibodies in Mice**

21 The critical endpoints here are decreased thymus weight and increased anti-dsDNA and
22 anti-ssDNA antibodies in female B6C3F1 mice (Keil et al., 2009).
23

24 **F.6.5.1. Dosimetry**

25 Mice were exposed to 1400 and 14000 ppb of TCE in drinking water, with an average
26 dose estimated by the authors to be 0.35 and 3.5 mg/kg/d, for 30 weeks. The dose-response
27 relationships were sufficiently supralinear that BMD modeling failed to produce an adequate fit.
28 The primary dose metric was selected to be the average amount of TCE metabolized/kg^{3/4}/day.
29 The lower dose group was the LOAEL for both effects, and the median estimate from the PBPK
30 model at that exposure level was 0.139 mg TCE metabolized/kg^{3/4}/day, which is used as the
31 rodent idPOD.
32



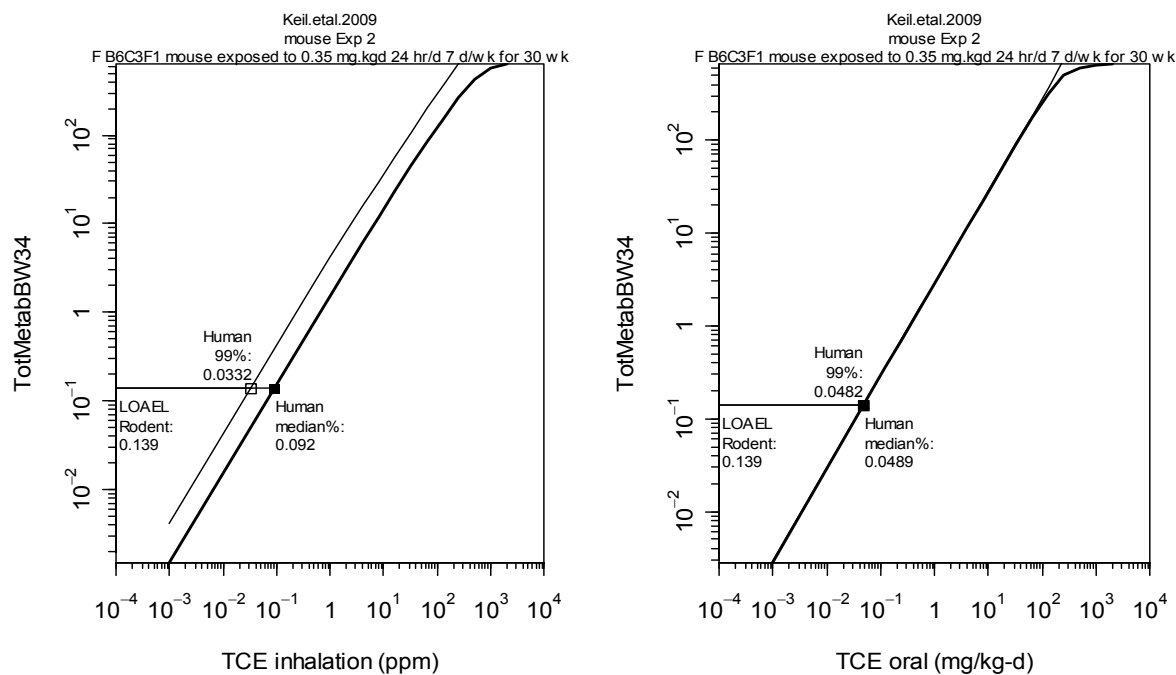
1
2
3 **Figure F-14. Derivation of HEC₉₉ and HED₉₉ corresponding to the rodent**
4 **idPOD from Woolhiser et al. (2006) for increased kidney weight in rats.**
5
6

7 **F.6.5.2. Derivation of HEC₉₉ and HED₉₉**

8 The HEC₉₉ and HED₉₉ are the lower 99th percentiles for the continuous human exposure
9 concentration and continuous human ingestion dose that lead to a human internal dose equal to
10 the rodent idPOD. The derivation of the HEC₉₉ of 0.0332 ppm and HED₉₉ of 0.0482 mg/kg/d for
11 the 99th percentile for uncertainty and variability are shown in Figure F-15. These values are
12 used as this critical effect's POD to which additional UFs are applied.
13

14 **F.6.6. Johnson et al. (2003)—Benchmark Dose (BMD) Modeling of Fetal Heart**
15 **Malformations in Rats**

16 The critical endpoint here is increased fetal heart malformations in female S-D rats
17 (Johnson et al., 2003).
18



1
2
3 **Figure F-15. Derivation of HEC₉₉ and HED₉₉ corresponding to the rodent**
4 **idPOD from Keil et al. (2009) for decreased thymus weight and increased**
5 **anti-dsDNA and anti-ssDNA antibodies in mice.**

6
7
8 **F.6.6.1. Dosimetry and Benchmark Dose (BMD) Modeling**

9 Rats were exposed to 2.5, 250, 1.5, or 1,100 ppm TCE in drinking water for 22 days
10 (GD 1–22). The primary dose metric was selected to be average amount of TCE metabolized by
11 oxidation/kg^{3/4}/day, with median estimates from the PBPK model for this study of 0.00031, 0.033,
12 0.15, and 88.

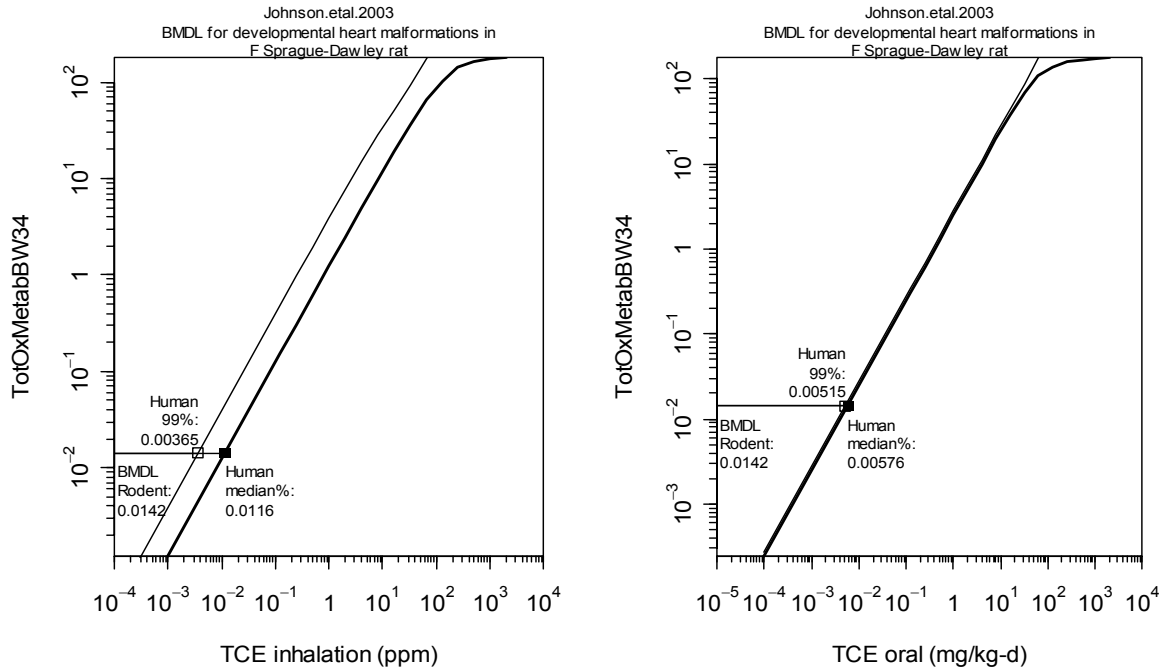
13 As discussed previously in Section F.4.2.1, from results of nested log-logistic modeling
14 of these data, with the highest dose group dropped, the idPOD of 0.0142 mg TCE metabolized
15 by oxidation/kg^{3/4}/day was a BMDL for a BMR of 1% increased in incidence in pups. A 1%
16 extra risk of a pup having a heart malformation was used as the BMR because of the severity of
17 the effect; some of the types of malformations observed could have been fatal.

18
19 **F.6.6.2. Derivation of HEC₉₉ and HED₉₉**

20 The HEC₉₉ and HED₉₉ are the lower 99th percentiles for the continuous human exposure
21 concentration and continuous human ingestion dose that lead to a human internal dose equal to
22 the rodent idPOD. The derivation of the HEC₉₉ of 0.00365 ppm and HED₉₉ of 0.00515 mg/kg/d

1 for the 99th percentile for uncertainty and variability are shown in Figure F-16. These values are
2 used as this critical effect's POD to which additional UFs are applied.

3



4

5

6 **Figure F-16. Derivation of HEC₉₉ and HED₉₉ corresponding to the rodent**
7 **idPOD from Johnson et al. (2003) for increased fetal cardiac malformations**
8 **in female S-D rats using the total oxidative metabolism dose metric.**

9

10

11 **F.6.7. Peden-Adams et al. (2006)—Lowest-Observed-Adverse-Effect Level (LOAEL) for**
12 **Decreased PFC Response and Increased Delayed-Type Hypersensitivity in Mice**

13 The critical endpoints here are decreased PFC response and increased delayed-type
14 hypersensitivity in mice exposed pre- and postnatally (Peden-Adams et al., 2006).

15 Mice were exposed to 1400 and 14,000 ppb in drinking water, with an average dose in
16 the dams estimated by the authors to be 0.37 and 3.7 mg/kg/d, from GD0 to postnatal ages of 3
17 or 8 weeks. The dose-response relationships were sufficiently supralinear that BMD modeling
18 failed to produce an adequate fit. In addition, because of the lack of an appropriate PBPK model
19 and parameters to estimate internal doses given the complex exposure pattern (placental and
20 lactational transfer, and pup ingestion postweaning), no internal dose estimates were made.
21 Therefore, the LOAEL of 0.37 mg/kg/d on the basis of applied dose was used as the critical
22 effect's POD to which additional UFs are applied.

This document is a draft for review purposes only and does not constitute Agency policy.

1 **F.7. REFERENCES**

- 2 Box GEP, Hunter WG, Hunter JS. (1978). Statistics for Experimenters, New York: John Wiley & Sons.
- 3 Buben, JA; O'Flaherty, EJ. (1985) Delineation of the role of metabolism in the hepatotoxicity of trichloroethylene
4 and perchloroethylene: a dose-effect study. Toxicol Appl Pharmacol 78:105–122.
- 5 Carney, EW; Thorsrud, BA; Dugard, PH; Zablony, CL. (2006) Developmental toxicity studies in CrI:Cd (SD) rats
6 following inhalation exposure to trichloroethylene and perchloroethylene. Birth Defects Research (Part B)
7 77:405–412.
- 8 Chia SE, Ong CN, Tsakok MF, Ho A. (1996) Semen parameters in workers exposed to trichloroethylene. Reprod
9 Toxicol 10(4):295–299.
- 10 Crofton, KM; Zhao, X. (1997) The ototoxicity of trichloroethylene: extrapolation and relevance of high-
11 concentration, short-duration animal exposure data. Fundam Appl Toxicol 38(1):101–106.
- 12 Filipsson, A.F., K. Victorin. (2003). Comparison of available benchmark dose softwares and models using
13 trichloroethylene as a model substance. Regulatory Toxicology and Pharmacology 37:343–355.
- 14 George, JD; Reel, JR; Myers, CB; Lawton, AD; Lamb, JC. (1986) Trichloroethylene: reproduction and fertility
15 assessment in F344 rats when administered in the feed. RTI Project No. 310-2344, NTP-86-085. National Institute
16 of Environmental Health Sciences, National Toxicology Program, RTP, NC.
- 17 Griffin JM, Gilbert KM, Lamps LW, Pumford NR. 2000. CD4(+) T-cell activation and induction of autoimmune
18 hepatitis following trichloroethylene treatment in MRL+/+ mice. Toxicol Sci 57:345–352.
- 19 Ikeda M, Otsuji H, Imamura T, Komoike Y. (1972) Urinary excretion of total trichloro-compounds,
20 trichloroethanol, and trichloroacetic acid as a measure of exposure to trichloroethylene and tetrachloroethylene. Brit
21 J Ind Med 29(3):328–33.
- 22 Johnson, PD; Goldberg, SJ; Mays, MZ; Dawson, BV. (2003) Threshold of trichloroethylene contamination in
23 maternal drinking waters affecting fetal heart development in the rat. Environ Health Perspect 111(3):289–292.
- 24 Keil, DE; Peden-Adams, MM; Wallace, S; Ruiz, P; Gilkeson, GS. (2009) Assessment of trichloroethylene (TCE)
25 exposure in murine strains genetically-prone and non-prone to develop autoimmune disease. Journal of
26 Environmental Science and Health, Part A 44: 443–453.
- 27 Kjellstrand P, Holmquist B, Alm P, Kanje M, Romare S, Jonsson I, Månsson L, Bjerkemo M. (1983a)
28 Trichloroethylene: further studies of the effects on body and organ weights and plasma butyrylcholinesterase
29 activity in mice. Acta Pharmacol Toxicol (Copenh) 53(5):375–84.
- 30 Kjellstrand P, Holmquist B, Mandahl N, Bjerkemo M. (1983b) Effects of continuous trichloroethylene inhalation on
31 different strains of mice. Acta Pharmacol Toxicol (Copenh) 53(5):369–74.
- 32 Land, PC; Owen, EL; Linde, HW. (1981) Morphologic changes in mouse spermatozoa after exposure to inhalational
33 anesthetics during early spermatogenesis. Anesthesiology 54:53–56.
- 34 Maltoni, C; Lefemine, G; Cotti, G. (1986) Experimental research on trichloroethylene carcinogenesis. In: Maltoni,
35 C; Mehlman MA., eds. Vol. 5. Archives of research on industrial carcinogenesis. Princeton, NJ: Princeton Scientific
36 Publishing;
- 37 Mhiri, C; Choyakh, F; Ben, HM; et al. (2004) Trigeminal somatosensory evoked potentials in trichloroethylene-
38 exposed workers. Neurosciences 9(2):102–107.

This document is a draft for review purposes only and does not constitute Agency policy.

- 1 Moser, Cheek & MacPhail. A multidisciplinary approach to toxicological screening III. Neurobehavioral toxicity. J
2 Toxicol. Environ. Hlth., 1995, 45, 173–210.
- 3 Narotsky, MG; Weller, EA; Chinchilli, VM; Kavlock, RJ. (1995) Nonadditive developmental toxicity in mixtures of
4 trichloroethylene, di(2-ethylhexyl) phthalate, and heptachlor in a 5 x 5 x 5 design. Fundam Appl Toxicol
5 27:203–216.
- 6 NCI (National Cancer Institute). (1976) Carcinogenesis bioassay of trichloroethylene. Division of Cancer Cause and
7 Prevention, National Cancer Institute, U.S. Department of Health, Education, and Welfare, DHEW Publication No.
8 (NIH) 76-802, Technical Report Series No. 2, 218 pages; NCI-CG-TR-2; NTIS PB254122.
9 http://ntp.niehs.nih.gov/ntp/htdocs/LT_rpts/tr002.pdf.
- 10 NTP (National Toxicology Program). (1988) Toxicology and carcinogenesis studies of trichloroethylene (CAS no.
11 79-01-6) in four strains of rats (ACI, August, Marshall, Osborne-Mendel) (gavage studies). Public Health Service,
12 U.S. Department of Health and Human Services; NTP TR-273; NIH Publication No. 88-2529. Available from the
13 National Institute of Environmental Health Sciences, Research Triangle Park, NC, and the National Technical
14 Information Service, Springfield, VA; PB88-218896. http://ntp.niehs.nih.gov/ntp/htdocs/LT_rpts/tr273.pdf.
- 15 NTP (National Toxicology Program). (1990) Carcinogenesis Studies of Trichloroethylene (Without Epichlorhydrin)
16 (CAS No. 79-01-6) in F344/N Rats and B6C3F1 Mice (Gavage Study). NTP TR 243. Research Triangle Park, NC:
17 U.S. Department of Health and Human Services.
- 18 Peden-Adams MM, Eudaly JG, Heesemann LM, Smythe J, Miller J, Gilkeson GS, et al. (2006). Developmental
19 immunotoxicity of trichloroethylene (TCE): studies in B6C3F1 mice. J Environ Sci Health A Tox Hazard Subst
20 Environ Eng 41:249–271.
- 21 Snedecor GW, Cochran WG. (1980). Statistical Methods (7th ed.), Ch. 9.12 and Ch 9.14 (pp.169–172)
- 22 U.S. EPA (Environmental Protection Agency) (2008b). Benchmark Dose Technical Guidance (Inter-Agency
23 Review Draft).
- 24 U.S. EPA (Environmental Protection Agency). (1994) Methods for derivation of inhalation reference concentrations
25 and application of inhalation dosimetry. Environmental Criteria and Assessment Office, Office of Health and
26 Environmental Assessment, Washington, Washington, DC; EPA/600/8-90/066F. Available from: National
27 Technical Information Service, Springfield, VA; PB2000-500023.
- 28 Woolhiser, MR; Krieger, SM; Thomas, J; Hotchkiss, JA. (2006) Trichloroethylene (TCE): Immunotoxicity potential
29 in CD rats following a 4-week vapor inhalation exposure. Dow Chemical Company, Toxicology & Environmental
30 Research and Consulting, Midland, MI, Study ID 031020, July 5, 2006, unpublished.

This document is a draft for review purposes only and does not constitute Agency policy.