

ARZ01-13409B

Id  
Date

P. Naphthas  
12.20.2001

**ROBUST SUMMARY  
OF INFORMATION ON**

**Substance Group:**

**PARAFFINIC NAPHTHAS**

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NB. Reliability of data included in this summary has been assessed using the approach described by Klimisch, et al.

Klimisch, H. J., Andreae, M. and Tillman, U, (1997)

A systematic approach for evaluating the quality of experimental toxicological and ecotoxicological data.

Regulatory Toxicology and Pharmacology 25, 1-5.

# 1. General Information

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## 1.1 GENERAL SUBSTANCE INFORMATION

**Substance type** : petroleum product  
**Physical status** : liquid  
**Remark** : Paraffinic naphtha streams are obtained by alkylation (catalytic reaction), isomerization (catalytic conversion) and solvent extraction, and contain saturated hydrocarbons, mainly in the range C5 to C12 and boiling in the range of approximately 35 to 230°C.

Light Alkylate Naphtha (CAS 64741-66-8) is a typical paraffinic naphtha stream.

Several samples have been used in testing for the toxicological properties of this group of substances. Details of three of the materials are given in the following table.

Sample (API 83-19) is a Light Alkylate Naphtha.  
LAN is a Light Alkylate Naphtha.  
LAND is a distillate of LAN.

Parameter	SAMPLE		
	API 83-19	LAN	LAND
Gravity (°API)	71.7		
Sulfur (wt %)	0.001		
RVP (psia)	6.7		
IBP (°F)	103		
FBP (°F)	331		
Olefins (% by MS)	0	0	0.03
Naphthenes (% by MS)	0.4	0.03	0
Aromatics (% by MS)	0.1	0	0
Saturates (% by MS)	99.5		
Paraffins		99.75	99.97
Mon (clear)	93.5		
RON (clear)	94.1		
Carbon No. (vol%)			
4		0.65	3.25
5		8.09	33.3
6		7.66	18.91
7		8.38	9.81
8		56.76	31.14
9		12.44	3.21
10		5.76	0.39

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**Conclusion**

2,2,4 trimethyl pentane; 2,5 dimethyl hexane; 2,3,4 trimethyl pentane, 2,3,3 trimethyl pentane and 1-methyl-1-ethyl cyclopentane, which represent 68% composition of the test substance. Concentrations for these components reached equilibrium in freshwater and saltwater by 24 and 12 hours respectively.

: Individual components of complex petroleum substances have specific and differing solubilities. Calculated and measured water solubilities for LAN components range from <1 to approximately 30 mg/L. At any particular loading rate, the resulting aqueous concentration of each chemical constituent is a function of the relative volume of the two phases (aqueous and the petroleum mixture), the partition coefficient between the phases, the amount of component present and the maximum water solubility of each component. Initially as the petroleum mixture is added in amounts below the solubility limit of the least soluble component the aqueous concentration increases proportionally until the least soluble component reaches a saturation concentration, and only the more soluble components continue to dissolve, resulting in a two phase system. Further addition of the petroleum mixture results in an aqueous concentration that is a non-linear function of the amount added.

**Reliability**  
26.06.2001

: (2) valid with restrictions

(9) (10) (11) (12) (19)

### 3. Environmental Fate and Pathways

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#### 3.1.1 PHOTODEGRADATION

**Type** : calculation  
**Light source** : Sun light  
**Method** : calculated by AOPWIN ver. 1.89. Method based on the work of R. Atkinson  
**Year** : 2000  
**GLP** : no  
**Test substance** : Light Alkylate Naphtha  
**Remark** : AOPWIN ver. 1.89 calculates atmospheric oxidation half lives of hydrocarbons in contact with hydroxyl radicals in the troposphere, under the influence of sunlight and in contact with O<sub>3</sub>. Atmospheric oxidation rates were calculated for the C<sub>5</sub> to C<sub>9</sub> hydrocarbon components found in LAN, CAS No. 64741-66-8. Detailed hydrocarbon analysis performed by Chevron Research was used to identify the components of this specific LAN sample. Based on a 12-hour day, the range for atmospheric half-lives for LAN constituents is: 1.074 days (2,3,5 trimethyl hexane) to 15.985 days (isopentane).

**Result** : Indirect Photolysis Yes  
Sensitizer OH radical  
Conc. of sensitizer 1.50E+06 radicals/cm<sup>3</sup>  
Rate Constant 0.6691E-12 (isopentane) cm<sup>3</sup>/mol-sec  
to 9.956E-12 (2,3,5 trimethyl hexane)  
Half-life 1.074 days to 15.985 days

**Reliability** : (2) valid with restrictions  
26.06.2001 (16)

#### 3.1.2 STABILITY IN WATER

**Test substance** : Light Alkylate Naphtha  
**Conclusion** : Hydrolysis unlikely  
**Reliability** : (1) valid without restriction  
26.06.2001 (13)

#### 3.3.1 TRANSPORT BETWEEN ENVIRONMENTAL COMPARTMENTS

**Type** : Calculated according to Mackay Level 1  
**Media** : Soil, air, water, suspended sediment and sediment  
**Remark** : Model based on chemical fugacity. Multimedia distribution was calculated for the C<sub>5</sub> to C<sub>9</sub> hydrocarbon components found in LAN, CAS No. 64741-66-8. Detailed hydrocarbon analysis performed by Chevron Research was used to identify the components of this specific LAN sample. Mobility in the aquatic and terrestrial environment is low due to low water solubility and high vapor pressure. The naphtha components will partition rapidly to air, where for the majority of these hydrocarbons will be rapidly oxidized by OH radicals.

**Result** : 

Medium	% distribution
Air	99.4 to 100
Soil	0.01 to 0.27
Water	0.001 to 0.02
Sediment	<0.001
Suspended sediment	<0.0

**Conclusion** : This complex petroleum mixture is expected to partition primarily to air

**Reliability** : (2) valid with restrictions  
26.06.2001 (14)

#### 3.5 BIODEGRADATION

**Type** : aerobic  
**Inoculum** : mixed, adapted inoculum of domestic activated sludge and soil  
**Contact time** : 56 day  
**Method** : CONCAWE. Test method for determining the inherent aerobic biodegradability of oil products. 1996/1997, and modification of ISO/DIS 14593

**Year** : 1999  
**GLP** : yes  
**Test substance** : Light Alkylate Naphtha  
**Method** : Test type:

Water quality-Evaluation of ultimate aerobic biodegradability of organic compounds in aqueous medium-Method by analysis of inorganic carbon in sealed vessels (CO<sub>2</sub> headspace test)

**Result** : Test material was inherently biodegradable since it achieved >20% biodegradability based on CO<sub>2</sub> production. By day 21 approximately 40% of the test material was degraded, a slight increase to 48% was observed by day 42, but by day 56 degradation had leveled back down to 40%. The test was considered valid according to CONCAWE criteria, as >60% biodegradation of positive control (63% actual) was observed by day 14, and total blank CO<sub>2</sub> production at termination was less than 15% of the organic carbon added as test substance. Temperature ranged from 18 to 21 °C, which deviated from the protocol value of 22 ±2°C. This deviation was not expected to have affected the outcome of this study.

Test Day	% Degradation (sd)	
	Hexadecane	Test Material
3	13.93 (1.85)	0.12 (0.07)
7	34.40 (4.54)	7.84 (7.80)
14	63.17 (0.94)	26.59 (0.85)
21	77.26 (6.52)	40.24 (5.00)
28	90.35 (7.14)	42.41 (2.54)
35	85.13 (n=1)	41.53 (9.90)
42	85.21 (n=1)	48.12 (1.77)
49	96.93 (8.94)	46.55 (1.04)
56	94.69 (4.10)	40.44 (0.76)

**Test condition** : Mixed inoculum prepared from soil and activated sludge was incubated with test substance or hexadecane (positive control) during a two week adaptation period. Triplicate test systems were incubated for both the test substance and hexadecane fed inoculum. Two additional, similar test substances were concurrently incubated in separate 160 ml test systems using the same inoculum and acclimation procedure. Duplicate blank control test systems were prepared which consisted of the mixed inocula in mineral medium but no test or positive control substance. Test medium consisted of glass distilled water and mineral salts (phosphate buffer, ferric chloride, magnesium sulfate, calcium chloride) prepared as described in ISO method.

Acclimation procedure-Activated sludge from aeration basin of Wareham Wastewater Treatment Plant (Mass., U.S.A.) was sieved through 2 mm and centrifuged at 1000 rpm for 10 minutes. After removal of supernatant the concentrated solids were diluted to 5 mg/ml suspended solids with reagent grade water. Soil was collected from a site located in a mixed hardwood and pine forest (Mass., U.S.A.). Site of sampling was cleared of debris and approximately 500 g of

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soil was obtained at a depth between 5-10cm from the soil surface. Soil was air-dried, sieved through a 2 mm sieve, and analyzed for moisture content (38%).

Test vessels (160 ml serum bottles) were filled with 103 ml of mineral medium containing 50 mg/l of yeast extract and 50 mg/l (dry weight) washed activated sludge, then approximately 0.16g of sieved soil (0.1 g dry wt) was added to each bottle. Test or reference substance were added directly to test systems using a 10 microliter Hamilton gas tight syringe. The volume required to achieve the specified mg carbon/l concentrations were calculated based on %carbon and specific gravity of the respective substance. The test substance %carbon (0.8505) and specific gravity (0.6690 mg/ $\mu$ l) information was supplied by the Sponsor. Hexadecane %carbon (0.8496) was calculated from the empirical formula and specific gravity (0.7749 mg/ $\mu$ l) was obtained from Verschueren (1983). Addition of respective substance was performed on an incremental basis to the appropriate vessels as follows: 4, 8 and 8 mg C/l were added on days 0, 7 and 11, respectively. Test vessels were sealed with butyl rubber septa/aluminum crimp caps and incubated at 22 ( $\pm$  2°C) in the dark.

Biodegradation by CO<sub>2</sub> determination-test initiation and procedure

On day 14 of the acclimation phase, all test system inoculum from blanks, positive control, and each of the three test substances was combined and filtered through glass wool, and aerated prior to use. The aerated mixed inoculum was then added to mineral medium to achieve 10% concentration based on total volume(100 ml inoculum/L). Test vessels (160 ml serum bottles) were filled with 103 ml of inoculated mineral medium. Respective test systems were dosed with either test substance or hexadecane as described for the acclimation procedure to achieve 20 mg carbon/lL concentration.

Duplicate test systems for each test substance, positive control and blank treatments were prepared for sacrifice at weekly sampling intervals for subsequent CO<sub>2</sub> analysis. After test system preparation, all vessels were placed in a walk-in chamber and incubated in the dark at 22°C ( $\pm$  2°).

On days 3, 7, 14, 21, 28, 35, 42, 49 and 56, 1ml of conc H<sub>3</sub>PO<sub>4</sub> was injected through the septum of each sacrificed test vessel. The acidified samples were shaken for 1 hr at 200 ppm, then analyzed for CO<sub>2</sub> using gas chromatography-thermal conductivity detection. Quantitation of inorganic mg C/l evolved was determined by linear regression analysis based on response factors for sodium carbonate standards spanning 1-30 mg carbon/l concentrations.

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: (1) valid without restriction

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## 4.1 ACUTE/PROLONGED TOXICITY TO FISH

<b>Type</b>	: Static with daily renewal
<b>Species</b>	: Pimephales promelas (Fish, fresh water)
<b>Exposure period</b>	: 96 hour(s)
<b>Unit</b>	: mg/l
<b>Analytical monitoring</b>	: yes
<b>Year</b>	: 1994
<b>GLP</b>	: yes
<b>Test substance</b>	: Light Alkylate Naphtha
<b>Method</b>	: No specific guideline was described as being used to conduct the test, however report references 1991 ASTM method E729-88a and 1982 EPA Support Documents for Environmental Testing: EPA 560/6-82-002. LL <sub>50</sub> and LC <sub>50</sub> calculated using binomial probability analysis. ASTM Special Technical Publication 634. 1977, pp 65-84. All NOEC values calculated using Fisher's exact test.
<b>Test condition</b>	: Test solutions were prepared as water accommodated fractions (WAF). Control and dilution water was Mobil Technical Center well water. Nominal loading rates of 0, 1.1, 5.2, 9.7, 19 and 74 mg/l were used to prepare test solutions.

WAFS were prepared for each test concentration by mixing the appropriate mass of substance in 9.4 liters of water for 24 hr in 9 liter glass bottles. The bottles were filled to neck height with water to minimize volatility. A measured amount of test substance was added into each bottle, and the bottles were capped tightly with a positive pressure siphoning apparatus. The siphoning apparatus consisted of a teflon lined neoprene stopper housing two teflon tubes. One tube extended to the bottom of the bottle for removal of the WAF solution, the other tube ended above the WAF surface, and was used to control air pressure during siphoning. During WAF preparation, parafilm was used to seal the external joint between the neoprene stopper and glass bottle. After stirring for 24 hrs using 25% or less vortex, the contents of the WAF solution bottles were allowed to settle for approximately 45 minutes to two hours, then siphoned by the positive pressure apparatus port and used for testing. Samples were also analyzed by Purge & trap/GC-FID for concentrations of the following: 2,3 dimethyl butane; 2,4 dimethyl pentane; 2,2,4 trimethyl pentane; 2,5 dimethyl hexane; 2,3,4 trimethyl pentane, 2,3,3 trimethyl pentane and 1-methyl-1-ethyl cyclopentane, which represent 68% composition of the test substance. Measured test concentrations of the light alkylate naphtha were based on the total combined concentrations of all analytes.

Fish were hatched and raised in-house, and were acclimated prior to experimentation for a minimum of 14 days on a 16/8hr light/dark cycle. Test vessels were 3.8 liter glass containers with teflon lined caps. Two replicates per treatment and 10 organisms per replicate were tested for each treatment and the control. Exposure containers were filled with no headspace and tightly sealed to prevent volatilization. Test solution renewal was performed daily by removal of most of the test solution (leaving adequate volume to avoid stressing test organisms) and replacing it with fresh WAF solution prepared at least 24 hrs prior to use.

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- Result** : Water temperature was 21.2 °C (0.2 °C sd). Test photoperiod was 16 hrs. light and 8 hr dark. Dissolved oxygen measurements were between 7.7 and 8.6, pH values between 7.844 and 8.23.
- Reliability** : Mortality (no. of deaths/treatment) at 96 hrs: 0, 0, 0, 15, 20 and 20, respectively in 0, 1.1, 5.2, 9.7, 19 and 74 mg/L treatments. All surviving organisms exhibited normal behavior.  
96-hr LL<sub>50</sub> = 8.2 mg/l, 5.2-9.7 mg/L w/ 95% C.I. (as nominal loading rate)  
96-hr LC<sub>50</sub> = 305 ppb, 164-384 ppb w/ 95% C.I. (measured concentrations)  
96-hr NOEL = 5.2 mg/l (as nominal loading rate)  
96-hr NOEC = 166 ppb (measured concentrations)
- (2) valid with restrictions  
Measured concentrations may not represent 100% of components, remaining hydrocarbon components in WAFs may be equally toxic and should have been quantitated to determine total measured concentrations.

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### 4.2 ACUTE TOXICITY TO AQUATIC INVERTEBRATES

- Type** : 48 Hr Static Acute Toxicity Test with 24Hr Renewal in Closed Systems  
**Species** : Daphnia magna (Crustacea)  
**Exposure period** : 48 hour(s)  
**Unit** : mg/l  
**Analytical monitoring** : yes  
**Year** : 1994  
**GLP** : yes  
**Test substance** : Light Alkylate Naphtha  
**Method** : No specific guideline was described as being used to conduct the test, however report references 1991 ASTM method E729-88a and 1982 EPA Support Documents for Environmental Testing: EPA 560/6-82-002.  
: Statistical Method: (FT - ME) EL<sub>50</sub> and EC<sub>50</sub> calculated using binomial probability analysis. ASTM Special Technical Publication 634. 1977, pp 65-84.
- Test condition** : Test solutions were prepared as water accommodated fractions (WAF). Control and dilution water was aged well water.

WAFS were prepared for each test concentration by mixing the appropriate mass of substance in 1.2 liters of water for 24 hr in aluminum foil covered 1 liter aspirator bottles fitted w/ a port near the bottle bottom. The bottles were filled to neck height with water to minimize volatility. A measured amount of test substance was pipetted below the water surface. After stirring for 24 hrs using 25% or less vortex, the contents of the aspirator bottles were allowed to settle for approximately 45 minutes, then drained from the port and used for testing. Samples were also analyzed by Purge & trap/GC-FID for concentrations of the following: 2,3 dimethyl butane; 2,4 dimethyl pentane; 2,2,4 trimethyl pentane; 2,5 dimethyl hexane; 2,3,4 trimethyl pentane, 2,3,3 trimethyl pentane and 1-methyl-1-ethyl cyclopentane, which represent 68% composition of the test substance. Measured test concentrations of the light alkylate naphtha were based on the total combined concentrations of all analytes.

Range finding toxicity studies were conducted at 1.2, 9.9

and 99 mg/l loading, using WAFS which were divided into duplicate aliquots and tested. Definitive toxicity studies were conducted at 9, 18, 35, 70, & 140 mg/l loading, using WAFS which were divided into duplicate aliquots and tested.

Test vessels were teflon cap-sealed 237 ml glass jars with 10 daphnids per jar and were completely filled with test solution.

During the study test system solutions: dissolved oxygen concentration range: 8.0 to 8.5; pH ranged from 8.00 to 8.2; temperature was 19.1 to 21.0 °C; hardness (mg/l) ranged from 180 - 204; alkalinity (mg/l) was 140-156 and conductivity (umhos) values were 385 - 390.

Daphnia magna, were supplied by testing laboratory; age < 24 hours old; obtained from culture maintained in-house since January 1994. The primary culture was obtained from Aquatic Research organisms, Hampton, NH, which was derived from EPA laboratory culture, in Cincinnati, Ohio.

**Result**

: Immobility (no. of organisms) at 48 hrs: 0, 0, 0, 12, 13 and 20 for 0, 9, 18, 35, 70 and 140 mg/l treatments. At the 35 and 70 mg/ nominal treatments, 8 and 7 organisms were observed to show lethargic movement, respectively.

48-hr  $EL_{50}$  = 32 mg/l (95% C.I. 18 to 140 mg/l) based upon nominal loading rate.

48 hr  $EC_{50}$  was 556 ug/l (95% C.I. 339 to 1140 µg/l) based on total measured alkyl concentrations.

48-hr NOEL = 18 mg/l based upon nominal loading rate.

48 hr NOEC was 339 ppb based on total measured alkyl concentrations.

**Reliability**

: (2) valid with restrictions  
Measured concentrations may not represent 100% of components, remaining hydrocarbon components in WAF may be equally toxic and should have been quantitated to determine total measured concentrations

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## 4.3 TOXICITY TO AQUATIC PLANTS E.G. ALGAE

<b>Species</b>	: Selenastrum capricornutum (Algae)
<b>Exposure period</b>	: 96 hour(s)
<b>Unit</b>	: Mg/L
<b>Analytical monitoring Method</b>	: yes : other: EPA. 1982. Guidelines and Support Documents for Environmental Effects Testing. EPA 560/6-82-002. Sections EG-8, ES-5.
<b>Year</b>	: 1995
<b>GLP</b>	: yes
<b>Test substance</b>	: Light Alkylate Naphtha
<b>Method</b>	: Statistical Method: EL <sub>50</sub> and EC <sub>50</sub> calculated using binomial probability analysis. ASTM Special Technical Publication 634. 1977, pp 65-84. All NOEL/NOEC values calculated using Fisher's exact test.
<b>Test condition</b>	: Individual test treatment solutions were prepared as Water Accommodated Fractions (WAFs). Test material was added to 2.3 liters of sterilized AAP test media (enriched with 515 mg/l of sodium bicarbonate, pH adjusted to 7.5 ± 0.1 with 0.1N HCl and sterilized by 0.22 micron filtration) in 2.0 liter aspirator bottles, covered with aluminum foil. The mixing vessels were sealed with foil covered stoppers and mixed on magnetic stir plates with teflon coated stir bars for approximately 24 hours at room temperature. After mixing the solutions were allowed to settle for one hour and the WAF was removed and used for testing. Test vessels were 125ml glass Erlenmeyer flasks that were completely filled (135 ml) with treatment solution and inoculated with algae. Algal cells obtained from testing laboratory cultures grown in sterile, nutrient enriched AAP media, and transferred every 5-9 days to fresh media. Original algal cultures obtained from American Type Culture Collection (ATCC Strain 22662), Rockville, MD, June 1994. Cell density of the algal stock culture inoculate was determined prior to study initiation with a hemacytometer cell and compound microscope. Twelve replicates were prepared for each treatment level. Nominal treatment levels were 0, 18, 70, 146, 292 and 1157 mg/l The initial algal concentration was 1.0 x 10 <sup>3</sup> cells/ml. All test replicates were placed on a shaker table at 100 oscillations per minute during the study and exposed to continuous fluorescent light, illumination at 400 + 50-ft candles. Triplicate samples were taken daily for cell counts and analytical testing. Cell densities were determined by direct microscopic examination. Samples at 0, 24, 48, 72 and 96 hrs were also analyzed by Purge & trap/GC-FID for concentrations of the following: 2,3 dimethyl butane; 2,4 dimethyl pentane; 2,2,4 trimethyl pentane; 2,5 dimethyl hexane; 2,3,4 trimethyl pentane, 2,3,3 trimethyl pentane and 1-methyl-1-ethyl cyclopentane, which represent 68% composition of the test substance. Measured test concentrations of the light alkylate naphtha were based on the total combined concentrations of all analytes.  Test temperature was 24 ± 2°C. The pH was 7.5 at test initiation, pH value at test termination not included in report.

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**Result** : Percent inhibition on growth determined by cell density (cells/mL):  
96 hour EL<sub>50</sub>=45mg/l (18-70 mg/l CI @95% )  
96 hour EC<sub>50</sub>=741ug/l (353-1060 ug/l CI @95% )  
96 hour NOEL=18 mg/l  
96 hour NOEC=741ug/l

Subcultures placed in fresh media (no test substance) after acute testing for nine days indicated that growth inhibition was algistatic in all treatments. No excursions from the protocol were noted. However, range finding and two previous definitive tests were performed and considered inconclusive due to inconsistencies in control and treatment cell densities, which presumably were resolved by modification of the AAP media. Additionally, control growth showed a lag during the first 48 hours of the study.

Nominal (mg/l) Conc (meas µg/l).	96hr cell density (cells/ml)	(% Inhibition)
Control	5.7x10 <sup>4</sup>	na
18 (0.112)	5.53x10 <sup>4</sup>	3.1
70 (0.305)	1.27x10 <sup>4</sup>	77.7
146 (0.498)	3.46x10 <sup>3</sup>	93.9
292 (0.610)	1.36x10 <sup>3</sup>	97.6
1157 (0.612)	1.60x10 <sup>3</sup>	97.2

**Reliability** : (2) valid with restrictions  
Measured concentrations may not represent 100% of components, remaining hydrocarbon components in WAFs may be equally toxic and should have been quantitated to determine total measured concentrations.

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## 5.1.1 ACUTE ORAL TOXICITY

<b>Type</b>	: LD <sub>50</sub>
<b>Species</b>	: rat
<b>Strain</b>	: Sprague-Dawley
<b>Sex</b>	: male/female
<b>Number of animals</b>	: 10
<b>Vehicle</b>	: Undiluted
<b>Value</b>	: > 7000 mg/kg bw
<b>Year</b>	: 1986
<b>GLP</b>	: yes
<b>Test substance</b>	: API 83-19 (Light Alkylate Naphtha)
<b>Method</b>	: Groups of five male and five female fasted rats were given API 83-19 at doses of 5 and 7 g/kg as a single oral dose. The animals were then allowed food and water ad libitum and were observed hourly for clinical signs for the first 6 hours after dosing. Observation was twice daily thereafter for 14 days. Body weights were recorded at 7 and 14 days after administration of test material. At the end of the study, the animals were killed and subjected to a gross necropsy and any abnormalities were recorded.
<b>Result</b>	: Clinical signs seen during the study included: hypoactivity, diarrhea, yellow-stained anal area, red discharge from nose, blood-like discharge on or around penile area, pale appearance and one female in the 5 g/kg group died within one hour of dosing. All except two animals had returned to normal by day 3 of the study. The oral LD <sub>50</sub> was found to be greater than 7 g/kg
<b>Reliability</b>	: (1) valid without restriction
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## 5.1.2 ACUTE INHALATION TOXICITY

<b>Type</b>	: LC <sub>50</sub>
<b>Species</b>	: rat
<b>Strain</b>	: Sprague-Dawley
<b>Sex</b>	: male/female
<b>Number of animals</b>	: 10
<b>Vehicle</b>	: Air
<b>Exposure time</b>	: 4 hour(s)
<b>Value</b>	: > 5 mg/l
<b>Year</b>	: 1987
<b>GLP</b>	: yes
<b>Test substance</b>	: API 83-19 (Light Alkylate Naphtha)
<b>Method</b>	: A group of 5 male and 5 female rats were exposed by whole body inhalation to API 83-19 at a nominal concentration of 5mg/l for 4 hours. After the 4 hour exposure the rats were observed twice daily for mortality. The animals were weighed prior to exposure and again on days 7 and 14 post exposure. On day 14 all surviving animals were killed by exsanguination following sodium pentobarbital anesthesia. For all animals, including those found dead during the study the lungs were removed, fixed and examined histologically.
<b>Result</b>	: The mean analytical and nominal exposure concentrations were 5.04 ± 0.74 and 6.31 mg/l respectively. All animals survived the study but exhibited languid behavior and a hunched appearance during the exposure.

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Female body weights were decreased at day 15 but this was attributed to pre-necropsy fasting. At necropsy there were no remarkable findings and histopathology of the lungs was normal.

**Reliability** : (1) valid without restriction  
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### 5.1.3 ACUTE DERMAL TOXICITY

**Type** : LD<sub>50</sub>  
**Species** : rabbit  
**Strain** : New Zealand white  
**Sex** : male/female  
**Number of animals** : 4  
**Vehicle** : Undiluted  
**Value** : > 2000 mg/kg bw  
**Year** : 1986  
**GLP** : yes  
**Test substance** : API 83-19 (Light Alkylate Naphtha)  
**Method** : The skin of the patched area of two rabbits of each sex had been abraded whilst the other two had intact skin. A weighed quantity of undiluted test material (equivalent to a dose of 2 g/kg) was applied to the dorsal skin of each of 4 male and 4 female rabbits. The applied dose was covered with an occlusive dressing (gauze and an impermeable covering). 24 hours after dosing, the patches were removed, the skin wiped and collars fitted to the rabbits to prevent oral intake of any residual test material. The collars were removed 24 hours later and the animals were observed for a total of 14 days post-dosing. At study termination the animals were killed with carbon dioxide and a gross necropsy was performed. Any abnormalities were recorded.

**Result** : A pain response (vocalization) was elicited from all the animals following application of the test material. Apart from skin irritation there were no other clinical signs of toxicity. Skin irritation ranged from slight to severe for erythema and edema, slight to moderate for atonia and coriaceousness and from slight to moderate for desquamation and fissuring. Subcutaneous hemorrhage, blanching and eschar was also observed.

**Reliability** : (1) valid without restriction  
24.10.2001 (4)

### 5.1.4 ACUTE TOXICITY, OTHER ROUTES

Not relevant

### 5.2.1 SKIN IRRITATION

**Species** : rabbit  
**Concentration** : undiluted  
**Exposure** : Occlusive  
**Exposure time** : 24 hour(s)  
**Number of animals** : 6  
**PDII** : 3.9  
**Result** : moderately irritating  
**Method** : Draize Test

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**Year** : 1986  
**GLP** : yes  
**Test substance** : API 83-19 (Light Alkylate Naphtha)  
**Method** : 0.5 ml of undiluted test material was applied to the shorn skin in two areas on each rabbit. One area was intact and the other abraded skin. The treated area was then covered with an occlusive dressing.  
After 24 hours the dressing was removed and the treated skin was wiped to remove any residue of test material. The degree of erythema and edema was recorded according to the Draize scale. A second reading of skin responses was made at 72 hours and again at 96 hours, 7 and 14 days. Results of the 24 and 72 hour readings were used to determine the Primary Irritation Index.

**Result** : The scores for erythema and edema were marginally greater for intact skin than abraded skin, but the difference was not biologically significant.  
Scores for intact skin at each of the observation intervals were:

Time	Erythema	Edema	Irritation score*
24 hours	2.0	1.7	3.5
72 hours	2.5	2.2	4.2
96 hours	2.7	2.8	4.9
7 days	2.5	2.3	4.5
14 days	0.8	1.0	1.2

\* Irritation score calculated as the sum of irritation scores for each test site divided by the number of animals at each observation period.  
PII is the sum of the 24- and 72- hour total irritation scores divided by 2

**Reliability** : (1) valid without restriction  
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(4)

### 5.2.2 EYE IRRITATION

**Species** : rabbit  
**Concentration** : undiluted  
**Dose** : 0.1 ml  
**Exposure Time** : See method  
**Comment** : Eyes rinsed 20-30 seconds after exposure (3 rabbits), Eyes not rinsed in remaining animals

**Number of animals** : 9  
**Result** : not irritating  
**Year** : 1986  
**GLP** : yes  
**Test substance** : API 83-19 (Light Alkylate Naphtha)  
**Method** : 0.1 ml of undiluted test material was applied to the corneal surface of one eye of each of 9 rabbits, the other eye was untreated and served as control.  
After 20 to 30 seconds the treated eyes of 3 rabbits were washed with lukewarm water for 1 minute. Eyes of the other 6 rabbits were not washed.  
Readings of ocular lesions for all animals were made at 1, 24, 48, 72 hours and 7 days after treatment. Sodium fluorescein was used to aid in revealing possible corneal injury.

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**Result** : No pain response was elicited on instillation of test material.  
No corneal or iridial irritation was seen during the study

**Reliability** : (1) valid without restriction  
24.10.2001

(4)

### 5.3 SENSITIZATION

**Type** : Buehler Test  
**Species** : guinea pig  
**Concentration** : Induction: 50 % occlusive, epicutaneous  
Challenge: 25 % occlusive, epicutaneous  
**Number of animals** : 10  
**Vehicle** : paraffin oil  
**Result** : not sensitizing  
**Year** : 1986  
**GLP** : yes  
**Test substance** : API 83-19 (Light Alkylate Naphtha)  
**Method** : 0.4 ml of a 50% mixture of test material and paraffin oil was applied under an occlusive dressing to the shorn skin of 10 male and 10 female animals. 6 hours after application, the dressings were removed and the skin wiped to remove residues of test material. The animals received one application each week for 3 weeks. The same application site was used each time. 2 weeks following the third application, a challenge dose (0.4 ml of a 25% mixture in paraffin oil) was applied in the same manner as the sensitizing doses. A previously untreated site was used for the challenge application. The application sites for sensitizing and challenge doses were read for erythema and edema 24 and 48 hours after patch removal. To assist in the reading of the response to the final challenge dose the test site was depilated 3 hours prior to reading by using a commercially available depilatory cream.

**Result** : Positive control (2,4-dinitrochlorobenzene), vehicle control and naive control groups were included in this study and the procedure for these was the same as for the test groups.  
At challenge, a very slight erythema was exhibited by one animal. The other 9 animals had no response. In contrast, all 20 of the positive controls responded with reactions ranging from slight to severe irritation. Only one naive control exhibited a very slight erythema upon challenge.

**Reliability** : (1) valid without restriction  
24.10.2001

(4)

## 5.4 REPEATED DOSE TOXICITY

<b>Species</b>	:	rat
<b>Sex</b>	:	male/female
<b>Strain</b>	:	Sprague-Dawley
<b>Route of admin.</b>	:	inhalation
<b>Exposure period</b>	:	13 weeks
<b>Frequency of treatment</b>	:	6 hours/day, 5 days/week
<b>Doses</b>	:	668, 2220, 6646 ppm
<b>Control group</b>	:	Yes
<b>NOAEL</b>	:	= 2220 ppm
<b>Method</b>	:	OECD Guide-line 413 "Subchronic Inhalation Toxicity: 90-day Study"
<b>Year</b>	:	1998
<b>GLP</b>	:	Yes
<b>Test substance</b>	:	The test material (LAND) was a distillate of a Light Alkylate Naphtha.

The compositions of the LAN and the distillate derived from it are shown in section 1.1. above.

The composition and uniformity chamber gas chromatographic results (%weight) were:

Component	Liquid	Vapor	
		At Start	At termination
n-butane	2.442	3.217	3.210
iso-pentane	29.854	33.517	34.343
2,3-dimethylbutane	12.437	11.963	12.977
2-methylpentane	4.064	4.775	4.096
2,4-dimethylpentane	5.923	5.663	5.663
2,3-dimethylpentane	2.904	2.794	2.680
2,2,4-trimethylpentane	18.35	16.897	16.885
2,3,4-trimethylpentane	4.343	3.772	3.578
2,3,3-trimethylpentane	5.258	4.614	4.505
2,2,5-trimethylhexane	3.096	2.641	2.499

Chamber concentrations were monitored throughout the study. Actual chamber concentrations were close to target concentrations.

Particle mass distribution measurements confirmed that no measurable test material was present as aerosol.

<b>Method</b>	:	Groups of 12 male and 12 female rats underwent whole body exposures to 668, 2220 and 6646 ppm LAND. Exposures were for 6 hours each day, 5 days per week for 13 weeks. Extra groups of 12 rats of each sex were exposed to the high dose level and for a recovery control group. These animals were maintained untreated for 28 days following cessation of the 13 weeks exposure.
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Neurobehavioral evaluations of motor activity and functional activity were performed pretest and during weeks 5, 9, 14 and 18 recovery groups. Animals were not exposed to LAND during these tests.

Following 13 weeks of exposure, 12 animals/sex/group were necropsied and microscopic examination was performed on selected tissues. Nervous tissue from 6 rats/sex/group was also examined microscopically.

At the end of the 4 week recovery period, 12 animals of each sex from the high and control groups were necropsied and selected tissues were examined microscopically.

During the study, clinical observations were made twice daily. Ophthalmoscopic evaluations were performed pretest and just prior to the scheduled sacrifices at 14 weeks and 18 weeks (recovery groups). Body weights and food consumption was measured throughout the study. Blood samples were taken from 12 fasted rats/sex/group at 14 and 18 weeks for hematological and clinical chemical measurements.

At termination (after 13 weeks exposure for the main study and after 18 weeks for the recovery animals) all animals were killed and subjected to a complete macroscopic examination. The following organs were weighed: adrenals, brain, heart, kidneys, liver, lung, ovaries, prostate, spleen, testes (with epididymes), thymus and uterus. Brain lengths and widths were measured for each rat.

A wide range of tissues (39) was removed from the control and high dose animals, were fixed and examined histopathologically. Additionally, kidneys from selected animals were stained with Mallory-Heidenhain and examined. Tissues were also removed from the nervous system (central and peripheral) of all animals for subsequent special staining and histopathological examination. Nervous system tissues were selected randomly from 6 rats/sex/group in the high dose and controls at the end of 13 weeks for microscopic examination. Specific brain regions examined were forebrain, cerebral cortex, hippocampus, basal ganglia, midbrain cerebellum and pons and medulla.

Neurobehavioral studies were undertaken as follows:

### Motor activity

Locomotor activity was monitored as the number of beam breaks in an activity box. Monitoring sessions were for 60 minutes, divided into twelve 5-minute intervals. Evaluation was made pretest and during weeks 5, 9, 14 and at the end of the 4 week recovery period. [A detailed description of the evaluation and analysis is provided in the publication but is not included here.]

### Functional Operational Battery

An assessment of the following was made:

- Home cage evaluations for Posture, vocalization, palpebral closure.
- Handling evaluations for reactivity to general stimuli, signs of autonomic function.
- open field behavior: arousal level, gait, urination and defecation

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frequency, convulsions, tremor, abnormal behavior, piloerection and exophthalmos.  
Reflex assessments for: response to visual and auditory stimuli, tail pinch, pupillary function.

### Result

Animals were also evaluated for fore limb and hind limb grip strength, landing foot splay and air righting ability.  
: There were no mortalities during the study and there were no treatment related signs of toxicity. A possible treatment related sign was an increased incidence of red facial staining in rats of both sexes in the high dose group. Mean body weights, body weight gains and food consumption was unaffected by treatment.

Hematological and clinical chemical measurements were unaffected except for a 5% decrease in hemoglobin, a 5% decrease in hematocrit and a 7% decrease in erythrocytes. The hemoglobin was still decreased (4%) after the 4 week recovery period. However, it was considered that these differences were toxicologically unimportant because they were small and within the historical range for the test laboratory.

Although there were some changes in AST and ALT in high dose females they were not considered to be toxicologically significant because several control animals also had elevated levels for these enzymes in the control groups and also relative to historical controls.

The organ weight changes were few. Absolute and relative kidney weights were increased in the males at all dose levels and they were also elevated in the high dose recovery animals. These increases correlated with the finding of hyaline droplets in the proximal convoluted tubules at microscopy.

Absolute and relative liver weights were observed in the high dose males and females at 13 weeks but the differences had disappeared after the recovery period. There were no pathological findings associated with this increase. The magnitude of the organ weight increases is shown below.

	Dose level (ppm)			
	668	2220	6646	Recovery
Males				
Abs. Kidney wt.	13.2	19.8	27	23
Rel. Kidney wt.		18	30	11
Abs. Liver wt.			21	
Rel. Liver wt.			25	
Females				
Abs. Liver wt.			17	12
Rel. Liver wt.			12	

### Conclusion

In the neurobehavioral studies no treatment-related effects were observed in the functional operational battery. In the study of motor activity, there were some statistically significant differences, but overall they did not occur in a dose related manner and furthermore were smaller than some of the differences seen during the pre dosing period.  
: LAND was not a neurotoxicant in the neurobehavioral studies that were conducted.  
LAND did induce a light hydrocarbon nephropathy in the male

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rats at all exposure levels, but this is regarded as species and sex specific and not relevant for human health risk assessment.  
Excluding the nephropathy, the NOEL for subchronic toxicity was 2220 ppm and for neurotoxicity was 6646 ppm.  
: (1) valid without restriction

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**Species** : rabbit  
**Sex** : male/female  
**Strain** : New Zealand white  
**Route of admin.** : dermal  
**Exposure period** : 28 Days  
**Frequency of treatment** : Once per day, three times per week for 4 weeks  
**Post obs. period** :  
**Doses** : 200, 1000 & 2000 mg/kg  
**Control group** : yes  
**Year** : 1986  
**GLP** : yes  
**Test substance** : API 83-19 (Light Alkylate Naphtha)  
**Method** : Prior to the study, 5-day range finding study was conducted. The method and results of the range-finding study are not included in this summary.

Undiluted API 83-19 was applied at doses of 200, 1000 and 2000 mg/kg/day to the shorn dorsal skin of groups of five male and five female rabbits. The test material was applied to the skin 3 times each week for 4 weeks (12 applications total). The applied material was covered with an occlusive dressing for 6 hours, which was then removed, and the skin was wiped with a dry gauze to remove any residual material. A group of five rabbits of each sex served as sham controls. The test skin site of each animal was examined and scored for irritation prior to each application of test material. Mortality and moribundity checks were performed twice daily and body weights were recorded weekly. At termination, blood samples were taken for a range of hematological and clinical chemical measurements. Urine samples were also collected and frozen for possible future examination. A complete gross necropsy was performed on all animals. Major organs were weighed and tissues were processed for subsequent histopathological examination.

**Result** : No deaths occurred during the study. During the latter half of the study, all but one high dose female looked thin. This was considered to be a treatment-related effect. Apart from skin irritation, there were no other treatment-related clinical signs. Weight gains of treated animals over the duration of the study was similar to controls except for the females at 2000 mg/kg. which were significantly reduced. The mean weight for these rabbits was the same at the end of the study as it was on day 1. A mean irritation score was calculated for each day and overall means were also calculated. (The mean irritation score, MIS, was the sum of irritation scores for both erythema and edema for all animals of a given dose group and sex.)

The overall MIS for each dose group was:

<u>Group/sex</u>	<u>MIS</u>	<u>Classification</u>
2000 mg/kg M	3.5	Moderate irritant
2000 mg/kg F	3.6	Moderate irritant
1000 mg/kg M	2.8	Moderate irritant
1000 mg/kg F	2.7	Moderate irritant
200 mg/kg M	0.5	Minimal irritant
200 mg/kg F	0.5	Minimal irritant
Control M	0	Non irritant
Control F	0	Non irritant

There were no remarkable findings in the hematological data from any of the male or female groups compared to controls. The only significant clinical chemical finding was an approximately 40% reduction in ALP of the 2000 mg/kg females. All other clinical chemical measurements were unremarkable.

There were few differences in organ weight between the control and treated animals, these were:

- 18% increase in R adrenal weight in 1000 mg/kg males
- 28% increase in L adrenal weight in 1000 mg/kg males
- 37% decrease in R ovary weight of 2000 mg/kg females.

In none of the above was there an associated change in the relative organ weights and the differences were not considered to be treatment-related.

At gross necropsy, treatment - related skin findings consisted of: dry, scaly, rough, fissured, reddened, crusted, and/or thickened skin. There were no other treatment-related findings at necropsy.

Although there were some findings at histopathology, they were not treatment-related except those in the skin. The skin changes consisted of a slight to moderate proliferative and minimal to moderately severe inflammatory changes in the skin of all animals in the 2000 mg/kg groups. These skin changes were accompanied by an increased granulopoiesis of the bone marrow. This was considered to be possibly related to stress or other factors resulting from skin irritation.

**Reliability** : (1) valid without restriction  
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(3)

### 5.5 GENETIC TOXICITY 'IN VITRO'

**Type** : Mouse lymphoma assay  
**System of testing** : Forward mutation assay using cell line L5178Y TK+/-  
**Metabolic activation** : with and without  
**Result** : Negative  
**Year** : 1985  
**GLP** : Yes  
**Test substance** : API 83-19 (Light Alkylate Naphha)  
**Method** : The test material was dissolved in acetone for this assay. Two positive control substances were used viz Ethyl methane sulphonate (EMS) at concentrations of 1.0 & 0.5 µl/ml and 7, 12-DMBA at concentrations of 7.5 & 5.0 µg/ml.

A cytotoxicity study was carried out prior to the mutagenicity assay. The results were difficult to interpret and as a consequence a second study was carried out and the results from this were used to determine the concentrations to be used in the subsequent lymphoma assay. It was established that complete toxicity occurred at 0.05 µl/ml for the non-activated cultures and at 0.5 µl/ml for S-9 activated cultures.

For the mutation assay the lymphoma cells were exposed for 4 hours to test material at concentrations ranging from 0.005 to 0.08 µl/ml without activation and 0.00004 to 0.8 µl/ml with S-9 activation. After exposure to the test material, the cells were allowed to recover for 2 days and then cultures were selected for cloning and mutant selection; TFT was used as the restrictive agent.

Eight non-activated and nine activated cultures were selected for cloning based on their degree of toxicity. The non-activated cultures that were cloned were treated with 0.005, 0.01, 0.015, 0.02, 0.025, 0.03, 0.035 or 0.04 µl/ml and resulted in a range of growth of 6 to 97%. The activated cultures that were cloned were treated with 0.0002, 0.0009, 0.0028, 0.008, 0.02, 0.045, 0.09, 0.7 or 0.75 µl/ml and produced a range of growth from 24 to 109%. Plates were prepared from TFT and from the VC cultures and after 10 to 12 days incubation these plates were scored for total number of colonies per plate. A mutation frequency was then determined.

The following criteria were used in judging the significance of the activity of the test article.

Positive - if there is a positive dose response and one or more of the 3 highest doses exhibit a mutant frequency which is two-fold greater than background level.

Equivocal - if there is no dose response but any one or more doses exhibit a two-fold increase in mutant frequency over background.

Negative - if there is no dose response and none of the test cultures exhibit mutant frequencies which are two-fold greater than background.

**Remark**

: Six mouse lymphoma assays were conducted but for technical reasons four of the assays were invalid. In the fifth assay none of the cultures that were cloned, whether in the presence or absence of S-9 activation exhibited mutant frequencies that were greater than those for the solvent control. However, the toxic response in the S-9 activation portion of the assay was erratic and this portion of the assay was repeated.

**Result**

This summary includes information from the fifth and sixth assays only, since they are the only ones considered to be valid.  
: The results of the fifth assay are as follows:  
After the 2 day recovery period, eight non-activated cultures and nine S-9 activated cultures were cloned based on their degree of toxicity. The mutant frequencies and the percentage total growth at each of the test concentrations is summarised in the following table.

Concentration (µl/ml)	Mutant frequency	% Total growth
<u>Non-Activated</u>		
0.04	0	34
0.035	0.5	3
0.03	0.2	30
0.025	0	46
0.02	0	93
0.015	0.2	102
0.01	0	79
0.005	0	93
Solvent 1	0.5	
Solvent 2	0.6	
DMBA 7.5 µl/ml	3.6	27
DMBA 5 µl/ml	1.9	57

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### S-9 Activated

0.75	0.2	101
0.7	0.2	16
0.09	0	88
0.045	-0.1	107
0.02	0	107
0.008	0.1	104
0.0028	0	100
0.0009	0	113
0.0002	-0.1	111
Solvent 1	0.6	
Solvent 2	0.6	
EMS 1µl/ml	8.7	3
EMS 0.5 µl/ml	6.8	29

The sixth assay was with S-9 activation only and the results were as follows:

### S-9 Activated

0.8	0.2	50
0.75	0	84
0.7	-0.1	90
0.65	-0.4	143
0.6	-0.1	99
0.5	-0.1	18
0.45	0.1	89
0.4	-0.1	72
0.35	0.1	76
0.25	-0.3	31
Solvent 1	0.8	
Solvent 2	0.8	
DMBA 7.5 µl/ml	1.4	62
DMBA 5 µl/ml	1.1	86

The authors concluded that according to the criteria used to judge the activity of the test material, the sample produced a negative response in the presence and absence of S-9 activation.

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: (2) valid with restrictions

(2)

## 5.6 GENETIC TOXICITY 'IN VIVO'

**Type** : Cytogenetic assay  
**Species** : rat  
**Sex** : male/female  
**Strain** : Sprague-Dawley  
**Route of admin.** : i.p.  
**Exposure period** : Up to 48 hours after single dose  
**Doses** : 0.3, 1.0 & 3.0 g/kg  
**Result** : negative  
**Year** : 1985  
**GLP** : yes  
**Test substance** : API 83-19 (Light Alkylate Naphtha)  
**Method** : The study design was as follows:

Treatment	Animals/sex/sacrifice time		
	6 hrs.	24 hrs.	48 hrs
Corn oil (vehicle)	5	5	5
API 83-19, 3 g/kg	5	5	5
API 83-19, 1 g/kg	5	5	5
API 83-19, 0.3 g/kg	5	5	5
Triethylenemelamine (Positive control)		5	

Test material in vehicle was given intraperitoneally at a dose of 5 ml/kg to groups of rats as shown above. Corn oil was used as vehicle control and TEM (0.5 mg/kg) as the positive control

Two to four hours prior to sacrifice the rats were given a single intraperitoneal dose of colchicine (1 mg/kg). 2 Males and one female in the high dose group died, these were replaced by substitute animals that were killed approximately 50 hours after administration of the test material.

Immediately after sacrifice bone marrow was obtained from the femurs of the animals. The marrow was washed and the cells were fixed before being spread on slides (at least 3 from each animal) for examination.

Slides were scored for chromosomal aberrations.

Where possible, a minimum of 50 metaphase cells from each animal were examined and scored for chromatid and chromosome gaps and breaks, fragments, structural rearrangements and ploidy (1-3).

A mitotic index (= No. of cells in mitosis/500 counted X 100) was calculated and recorded.

The data were evaluated according to the following criteria:

For the test to be considered to be valid, the % of cells in the negative control group demonstrating aberrations of any type, other than gaps, must not exceed 4%.

The % of cells with aberrations in the positive control group must be statistically increased ( $p < 0.05$ ) relative to the vehicle control using Chi-square statistics.

The test material is considered positive when the % of cells with aberrations in any treatment group is significantly increased ( $p < 0.05$ ) relative to the vehicle control using Chi-square analysis and the number of aberrations per

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### Result

cell is also significantly increased ( $p < \text{or} = 0.05$ ) relative to the vehicle control using t-test statistics.

: The dose levels used in the assay were selected on the basis of a preliminary screen in which only one male rat died within 24 hours following the administration of API 83-19 as a single i.p. dose to 4 rats of each sex. In the cytogenetics assay, 5 of 18 males and 4 of 18 females receiving 3 g/kg API 83-19 died within 3 days. At this dose level, there was a weight loss of 10% and 9% in males and females respectively within 48 hours of administration. Other signs of toxicity included piloerection, crusty eyes and noses and excess lacrimation. No sex-related differences were noted in the study and therefore the data for males and females were combined for the cytogenetics evaluation. The results are summarized in the following table.

	0.3 g/kg	1 g/kg	3 g/kg	Positive	Vehicle
<u>Cells with aberrations</u>					
6 hrs	0	2	0		0
24 hrs	1	0	1	171	0
48 hrs	1	0	1		0
<u>Incidence of aberrations (%)</u>					
6 hrs	0	0.4	0		0
24 hrs	0.2	0	0.2	34.2	0
48 hrs	0.2	0	0.3		0
<u>No. Gaps</u>					
6 hrs	0	2	0		0
24 hrs	0	0	0	15	1
48 hrs	0	0	4		1
<u>No. Breaks</u>					
6 hrs	0	2	0		0
24 hrs	1	0	1	197	0
48 hrs	2	0	1		0
<u>Aberrations per cell</u>					
6 hrs	0	0.004	0		0
24 hrs	0.002	0	0.002	2.336	0
48 hrs	0.004	0	0.003		0

NB.1. 500 cells were evaluated for each time point at each dose level.

NB.2. In the API 83-19 and vehicle control groups no rearrangements were observed and no aberrations from severely damaged cells were seen. In contrast, 51 rearrangements and 920 aberrations from severely damaged cells were seen in the positive control group.

**Reliability**  
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: (1) valid without restriction

(1)

## 5.8 TOXICITY TO REPRODUCTION

Type	: One generation study
Species	: rat
Sex	: male/female
Strain	: Sprague-Dawley
Route of admin.	: inhalation
Exposure period	: 6 hours per day
Frequency of treatment	: Daily
Premating exposure period	
Male	: 14 days
Female	: 14 days
Duration of test	: females 7 weeks, males 8 weeks
Doses	: 5, 12.5 and 25 g/m <sup>3</sup>
Control group	: yes
NOAEL Parental	: > 24700 mg/m <sup>3</sup>
NOAEL F1 Offspr.	: > 24700 mg/m <sup>3</sup>
Method	: Adaptation of OECD No. 421
Year	: 1998
GLP	: yes
Test substance	: Distillate of light alkylate naphtha

The test material was prepared to be representative of the fraction of light alkylate naphtha to which man would normally be exposed during normal handling and use.

The test material (LAND) was obtained by the distillation of light alkylate naphtha (LAN) and collecting that fraction that boiled over the temperature range 78 to 145°F. The sample was analyzed and its composition compared to the light alkylate naphtha from which it was derived. The compositions of the distillate and starting material were as follows:

Compound	Weight %	
	LAND	LAN
n-butane	3.42	0.84
isopentane	63.59	12.61
n-pentane	1.33	0.23
2,3-dimethylbutane	22.51	4.74
2-methylpentane	6.44	1.57
3-methylpentane	2.26	0.74
2,4-dimethylpentane	0.29	4.09
2,2,4-trimethylpentane	0.06	23.92
2,3,3-trimethylpentane	0	8.99
2,3,4-trimethylpentane	0	11.56

**Method** : The test material was totally vaporized and diluted with air to achieve the desired concentrations for the study. Exposures were conducted in one cubic meter whole-body chambers. Chamber concentrations were monitored three times daily by GC/FID. All animals were housed individually in suspended mesh cages.

10 animals of each sex were exposed 6 hours each day to test material at target concentrations of 5, 12.5 and 25 g/m<sup>3</sup>.

The animals were exposed for 6 hours each day. Parental females were exposed for 14 days prior to mating, throughout mating and gestation days 0-19 (7 consecutive weeks). Dams and their litters were sacrificed on postpartum day 14. Parental males were also exposed for 14 days prior to mating, during mating, throughout the female gestation and post partum period and throughout the female necropsy period (8 consecutive weeks). Rats were mated in a 1:1 ratio and females were monitored for evidence of mating by the examination of a vaginal lavage sample for sperm or vaginal plug. If sperm or a vaginal plug were observed, the female was considered to be at day 0 of gestation and the male was removed from the female at this stage. If there was no evidence that mating had occurred the pairs were allowed to remain together up to a period of 2 weeks after which time the female was assumed to be pregnant. All animals were observed for clinical signs at least twice daily throughout the study. Body weights and food consumption were recorded throughout the study. Each litter was examined as soon as possible after delivery to establish number and sex of pups, stillbirths, live births and presence of gross abnormalities. Neonatal survival was monitored and all pups were killed postpartum days 4 or 5. Parental females were killed on gestation day 25 if they had not delivered, otherwise they were killed on postpartum days 4 or 5.

**Result**

At necropsy each parental animal was examined macroscopically for structural abnormalities and pathological changes with emphasis on reproductive organs. Additionally the number of implantation sites and corpora lutea of each female were recorded. Lungs, trachea and larynx were removed in their entirety. The right middle lobe of the lung was weighed, the remaining lobes were fixed for subsequent histopathological examination. The testes and epididymes of the males were weighed and then fixed for histological examination as were the ovaries of the females.

: The chamber concentrations of test material were found to be between 96 and 104% of nominal, the mean highest dose concentration being 24.7 mg/m<sup>3</sup>. The vapor compositions were also found to be similar to that of the parent test material. No parent animals died or were killed during the study and there were no clinical signs. Body weights and food consumption were unaffected by exposure to test material. Results on reproductive capacity and fertility are summarized in the following table.

Parameter	Treatment group 9g/m <sup>3</sup>			
	0	5	12.5	25
Pregnancy (%)	80	80	100	80
Litters with live pups	8	8	9	8
Implantation sites	14.9	16.8	13.9	17.3
Pups delivered	14.4	15.6	14.3	15.6
Live pups/litter	14.4	14.8	13.8	15.5
No. liveborn	115	118	124	124
Live birth index (%)	100	94	96	99
Pups surviving 4 days	113	114	122	123
Viability index (%)	98	97	98	99
Litter wt day 1	7.2	7.3	7.1	7.1
Litter wt day 4	10.8	11.1	11.2	10.5

## 5. Toxicity

**Id** P. Naphthas  
**Date** 10.12.2001

**Reliability**  
24.10.2001

There were no treatment-related findings observed at necropsy .  
Organ weights were unaffected by treatment and there were no  
treatment-related histological findings.  
: (1) valid without restriction

(8)

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Id P. Naphthas  
Date 10.12.2001

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Date 10.12.2001

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## 6. References

**Id** P. Naphthas  
**Date** 10.12.2001

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**ROBUST SUMMARY  
OF INFORMATION ON**

**Substance Group:** **GASOLINE**

**Summary prepared by:** American Petroleum Institute

**Creation date:** 13 MARCH 2001

**Printing date:** 11 DECEMBER 2001

**Date of last Update:** 11 DECEMBER 2001

**Number of Pages:** 49

NB. Reliability of data included in this summary has been assessed using the approach described by Klimisch, et al.

Klimisch, H. J., Andreae, M. and Tillman, U, (1997)

A systematic approach for evaluating the quality of experimental toxicological and ecotoxicological data.  
Regulatory Toxicology and Pharmacology 25, 1-5.

# 1. General Information

**Id** Gasoline  
**Date** 11.12.2001

## 1.1 GENERAL SUBSTANCE INFORMATION

**Substance type** : petroleum product  
**Physical status** : liquid  
**Remark** : Gasoline is a complex combination of hydrocarbons consisting primarily of paraffins, cycloparaffins, aromatic and olefinic hydrocarbons having carbon numbers predominantly greater than C3 and boiling in the range of 30°C to 260°C.

To achieve acceptable physical and combustion properties, gasoline is prepared by blending naphtha streams, each of which will consist predominantly of one or other chemical type.

The streams normally used are:

Paraffinic streams derived from alkylation, isomerization and solvent extraction

Olefinic streams derived from catalytic cracking

Naphthenic streams derived from crude oil distillation

Aromatic streams derived from catalytic reforming

The American Petroleum Institute prepared a blend of naphtha streams which was considered to be typical of US gasoline in the middle 1970s. The blend was designated PS-6 gasoline.

The benzene

content of PS-6 was adjusted upwards to 2.0% by volume, although this would never happen in normal refinery practice. The proportions of the various naphtha streams used to prepare the PS-6 gasoline blend were:

<u>Naphtha stream/Component</u>	<u>% vol.</u>
Paraffinic naphtha streams	22.0
Olefinic naphtha streams	52.1
Aromatic naphtha streams	21.3
Benzene	0.8
n-butane	3.8

A more comprehensive characterization of PS-6 gasoline was reported by MacFarland et al. as follows:

Research Octane No.	92.0
Motor Octane No.	84.1
Reid vapor pressure (Lbs)	9.5
Distillation	
Initial boiling point (°C)	93
Final boiling point (°C)	428
API Gravity	60.6
Gum, ASTM D381 (mg/gal)	1
Sulfur (ppm)	97
Phosphorus (g/gal)	<0.005
Lead (g/gal)	<0.05
Stability (hrs)	>24
Hydrocarbon analysis, ASTM D1319	
Aromatics (vol%)	26.1
Olefins (vol%)	8.4

# 1. General Information

**Id** Gasoline  
**Date** 11.12.2001

Saturate (vol%)	65.5
Benzene (%)	2.0

An unleaded gasoline, tested in an inhalation teratology study had the following physical chemical characteristics:

API Gravity	57.4
Reid vapor pressure (Lbs)	8.8
Initial boiling point (°C)	88
Final boiling point (°C)	378
Sulfur (ppm)	157
Vapor density	3.4
Paraffins (vol%)	47
Olefins (vol%)	4
Naphthenes (vol%)	10
Aromatics (vol%)	39
C6 Aromatics (%)	1.2
C7 Aromatics (%)	10.2
C8 Aromatics (%)	16.3
C9-plus Aromatics (%)	11.3

19.11.2001

(22)

## 1.8 OCCUPATIONAL EXPOSURE LIMIT VALUES

<b>Type of limit</b>	: TLV (US)
<b>Limit value</b>	: 300 ppm
<b>Short term exposure</b>	
<b>Limit value</b>	: 500 ppm
<b>Schedule</b>	: 8 hour(s)

04.07.2001

(1)

## 2. Physico-Chemical Data

**Id** Gasoline  
**Date** 11.12.2001

### 2.1 MELTING POINT

Not relevant

### 2.2 BOILING POINT

See section 1.1 (General substance information)

### 2.3 DENSITY

**Type** : relative density  
**Value** : ca. 50  
**Year** : 1984  
**Test substance** : API PS-6 gasoline  
**Reliability** : (1) valid without restriction  
18.11.2001 (24)

### 2.5 PARTITION COEFFICIENT

**Log pow** : = 2.13 - 4.5 at 25° C  
**Method** : Calculated by LOGKOWWIN ver. 1.65.  
**Year** : 2000  
**GLP** : no  
**Test substance** : Gasoline CONCAWE sample, CWE5, Blend (match to API PS-6), CAS No. 86290-81-5  
**Remark** : Log P values represent the spread of calculated and/or measured values for C5 to C8 hydrocarbon components found in gasoline, CAS No 86290-81-5. Detailed hydrocarbon analysis was used to identify the components of this specific gasoline sample. Calculation based on an atom/fragment contribution method of W. Meylan and P. Howard. Calculated SAR result for surrogate structure contained in program database (smilecas.dat).  
**Reliability** : (2) valid with restrictions  
19.11.2001 (13) (26)

**Log pow** : = 2.73 - 4.85 at 25° C  
**Method** : Calculated by LOGKOWWIN ver. 1.65.  
**Year** : 2000  
**GLP** : No  
**Test substance** : Individual test treatment solutions were prepared as Water Accommodated Fractions (WAFs). Test material was added to 2.4 liters of sterilized AAP test media (enriched with 515 mg/l of sodium bicarbonate, 300 µg/l EDTA  
**Remark** : Log P values represent the spread of calculated and/or measured values for C5 to C9 hydrocarbon components found in LSRN, CAS No 64741-46-4. Detailed hydrocarbon analysis was used to identify the components of this specific high naphthenic LSRN sample. Calculation based on an atom/fragment contribution method of W. Meylan and P. Howard. Calculated SAR result for surrogate structure contained in program database (smilecas.dat).  
**Reliability** : (2) valid with restrictions  
21.11.2001 (13) (26)

### 2.6.1 WATER SOLUBILITY

<b>Method</b>	: Preparation of Water Soluble Fraction
<b>Year</b>	: 1995
<b>GLP</b>	: yes
<b>Test substance</b>	: Gasoline CONCAWE sample, CWE5, Blend (match to API PS-6), CAS No. 86290-81-5
<b>Method</b>	: Water Accommodated Fractions (WAFs) of LSRN were prepared at 100 mg/l loading in freshwater and equilibrated for 48 hours in tightly closed systems with minimal headspace.
<b>Result</b>	: Gas chromatographic analysis of BTEX components indicated freshwater solubility at 24 hours of 3.1, 3.1, <6.9E-3, and 0.92 ppm (as BTEX, respectively).
<b>Conclusion</b>	: Individual components of complex petroleum substances have specific and differing solubilities. Calculated and measured water solubilities for LSRN components range from approximately 3 to 2000 mg/l. At any particular loading rate, the resulting aqueous concentration of each chemical constituent is a function of the relative volume of the two phases (aqueous and the petroleum mixture), the partition coefficient between the phases, the amount of component present and the maximum water solubility of each component. Initially as the petroleum mixture is added in amounts below the solubility limit of the least soluble component the aqueous concentration increases proportionally until the least soluble component reaches a saturation concentration, and only the more soluble components continue to dissolve, resulting in a two phase system. Further addition of the petroleum mixture results in an aqueous concentration that is a non-linear function of the amount added.
<b>Reliability</b> 19.11.2001	: (2) valid with restrictions

(7) (14) (15) (16) (30)

### 3. Environmental Fate and Pathways

Id Gasoline  
Date 11.12.2001

#### 3.1.1 PHOTODEGRADATION

<b>Type</b>	: Calculated
<b>Light source</b>	: Sun light
<b>Indirect photolysis</b>	
<b>Sensitizer</b>	: OH
<b>Rate constant</b>	: $\text{cm}^3/(\text{molecule}\cdot\text{sec})$
<b>Degradation</b>	: % after
<b>Method</b>	: Calculated by AOPWIN ver. 1.89. Method based on the work of R. Atkinson
<b>Year</b>	: 2000
<b>GLP</b>	: No
<b>Test substance</b>	: Gasoline CONCAWE sample, CWE5, Blend (match to API PS-6), CAS No. 86290-81-5
<b>Remark</b>	: AOPWIN ver. 1.89 calculates atmospheric oxidation half lives of hydrocarbons in contact with hydroxyl radicals in the troposphere, under the influence of sunlight and in contact with $\text{O}_3$ . Atmospheric oxidation rates were calculated for the C5 to C8 hydrocarbon components found in gasoline. Detailed hydrocarbon analysis was used to identify the components of this specific gasoline sample. Based on a 12-hour day, the range for atmospheric half-lives for gasoline constituents is: 0.789 days (m-xylene) to 15.985 days (isopentane).
<b>Result</b>	: Indirect Photolysis Sensitizer: OH radical Conc. of sensitizer: $1.50\text{E}+06$ OH radicals/cm <sup>3</sup> Rate Constant: $0.6991\text{E}-12$ (isopentane) to $13.5606\text{E}-12$ (m-xylene) $\text{cm}^3/\text{molecule}\cdot\text{sec}$ Half-life: 0.789 to 15.985 days
<b>Reliability</b> 19.11.2001	: (2) valid with restrictions <span style="float: right;">(13) (27)</span>

#### 3.1.2 STABILITY IN WATER

<b>Test substance</b>	: Gasoline CONCAWE sample, CWE5, Blend (match to API PS-6), CAS No. 86290-81-5
<b>Conclusion</b>	: Hydrolysis unlikely
<b>Reliability</b> 19.11.2001	: (1) valid without restriction <span style="float: right;">(17)</span>

#### 3.3.1 TRANSPORT BETWEEN ENVIRONMENTAL COMPARTMENTS

<b>Type</b>	: Calculated
<b>Media</b>	: Soil, air, water, suspended sediment, sediment
<b>Method</b>	: Calculated according to Mackay Level I
<b>Year</b>	: 2000
<b>Remark</b>	: Model based on chemical fugacity. Physical properties input are those calculated by the EPIWIN Estimation 3.04 program and included in this summary. Values represent the spread of calculated values for C5 to C8 hydrocarbon components found in gasoline. Detailed hydrocarbon analysis was used to identify the components of this specific gasoline sample.

The majority of components in gasoline will partition rapidly to air, where these hydrocarbons will be rapidly oxidized by OH radicals. With the exception of toluene, partitioning to air is > 97% for all components.

### 3. Environmental Fate and Pathways

Id Gasoline  
Date 11.12.2001

**Result** : Medium % distribution  
Air: 97 to 99.99  
Soil: 0.00 to 1.2  
Water: 0.003 to 2.7  
Sediment <0.001 to 0.02  
Suspended sediment <0.001 to 0.02

**Test substance Conclusion** : Gasoline CONCAWE sample, CWE5, Blend (match to API PS-6)  
: The constituents of this complex petroleum mixture are expected to partition primarily to air. Moderate partitioning to water and soil is predicted for the aromatic components of this mixture

**Reliability** : (2) valid with restrictions  
19.11.2001 (13) (23)

#### 3.5 BIODEGRADATION

**Type** : Aerobic  
**Inoculum** : Activated aerobic sludge obtained from an urban wastewater treatment plant.  
**Deg. Product Method** : Non-guideline research method using a closed-system shake flask apparatus.  
**Year** : 1999  
**GLP** : No data  
**Test substance** : CAS No. 86290-81-5; a commercial unleaded gasoline topped at 76°C by distillation

**Method** : Aerobic Biodegradability - Evaluation of biodegradability of gasoline in aqueous medium. Method by analysis of disappearance of carbon compounds (gas chromatography with flame ionization detector), kinetics of O<sub>2</sub> consumption (respirometry), and CO<sub>2</sub> production (gas chromatography with thermal conductivity detector).

Exposure period was 16 or 25 days.  
See test conditions for more details.

**Result** : Biodegradation and Mineralization of Gasoline  
Gasoline was degraded up to 94% under non-limiting conditions after 25 d incubation (500 ml substrate/l medium). The carbon balance of gasoline degradation showed that 61.7% of gasoline was mineralized to CO<sub>2</sub> and that microbial cell production accounted for the remaining carbon of gasoline degraded. Biomass formation and mineralization occurred mainly during the initial fast degradation phase whereas essentially mineralization occurred during the second slow degradation phase. Individual classes of hydrocarbons degraded and carbon balance were shown to be:

Hydrocarbon Class	Amount in Gasoline mg/g	Amount in	
		After 2 days	After 25 days
Aromatics	789	88%	99%
Branched alkanes	165	14	74
Linear alkanes	23	17	92
Cyclic alkanes	17	10	99
Alkenes	6	71	99

### 3. Environmental Fate and Pathways

Id Gasoline  
Date 11.12.2001

#### Carbon balance

Substrate or Products	Initial amount (mg C/l)	Final amount (mg C/l)
Gasoline	357	18
Biomass	39	165
CO <sub>2</sub>	0	204
Total Carbon	396	387

#### Kinetic Experiments with Gasoline

Two main degradation phases were found, one fast degradation phase (FDP), which started after an 18 h lag period and lasted until the 40th hour. The maximum rate of oxygen consumption during the FDP was 44 mg/l/h and the average rate was 24.5 mg/l/h. The FDP was followed by a slow degradation phase (SDP) where the rate of oxygen consumption slowed steadily from the 40th hour until the 25th day. The average rate was 15 mg/l/d, which was approximately 40 times slower than during the FDP.

#### Test condition

: Activated sludge containing approximately 3 g/l dry weight was centrifuged at 15000 g for 20 min and re-suspending the biomass in the same volume of nutrient solution. The microbial suspension was used to inoculate nutrient solution at a final concentration of 100 mg dry weight/l. Gasoline (400 mg/l) or individual hydrocarbons (150 mg/l) were added to the medium as the sole carbon source. The nutrient solution was a vitamin-enriched mineral salt medium described by Bouchez et al. Appl. Microbiol. Biotechnol. 43:156-164 (1995).

#### Biodegradation of Gasoline

The biodegradation tests were performed in 500-ml flasks with sidearms equipped with Mininert® valves. 25 ml of gasoline were added to 50 ml of inoculated nutrient medium (i.e., 500 ml substrate/l medium) through the valve with a syringe. The flasks were incubated for 25 days at 30°C with alternate shaking (70 strokes per min). After the incubation period, 5 ml of CH<sub>2</sub>Cl<sub>2</sub> containing 600 mg/ml dodecane as internal standard was introduced to the flasks through the valve, and the remaining hydrocarbon compounds were extracted for 1 h under shaking. The flasks were refrigerated overnight at 4°C before opening. The suspensions were centrifuged at 35000 g for 30 min at 4°C. The CH<sub>2</sub>Cl<sub>2</sub> phase of each flask was then analyzed by gas chromatography for carbon compounds. Experiments were performed in duplicate and abiotic controls were prepared similarly to the other treatments with the exception that 1 g/l HgCl<sub>2</sub> were added to the flasks before incubation.

#### Mineralization of Gasoline

Measurements of CO<sub>2</sub> evolved during the biodegradation of gasoline were conducted in 240-ml flasks closed by Viton® stoppers. 18 ml of inoculated culture medium were added to each flask along with 5 ml of gasoline (i.e., 500 ml substrate/l medium). Flasks were incubated at 30°C for 25 days under alternate shaking. At the end of the incubation period the contents of each flask was acidified with 0.5 ml HNO<sub>3</sub> (68%) and CO<sub>2</sub> was measured by gas chromatography. Endogenous respiration of inoculated medium was measured in flasks without gasoline added.

### 3. Environmental Fate and Pathways

Id Gasoline  
Date 11.12.2001

Kinetic Experiments with Gasoline  
Kinetics of O<sub>2</sub> consumption during gasoline biodegradation were determined in duplicate at 30°C over 25 d by respirometry. 500 ml stirred culture flasks contained 250 ml of inoculated nutrient medium and 125 ml of gasoline (i.e., 500 ml substrate/l medium). Control experiments without gasoline were also done. Kinetics of hydrocarbon degradation also was monitored by respirometry. Incubation was stopped at selected times and the remaining hydrocarbons were extracted as described above and analyzed by gas chromatography.

Kinetic Experiments with Individual Hydrocarbons  
Kinetics of CO<sub>2</sub> production during the degradation of individual hydrocarbons was carried out at 30°C over 16 days. Treatments were prepared in 125 ml shaken flasks with 25 ml of nutrient solution containing 70 mg/l of inoculum biomass and 5 ml of hydrocarbon (i.e., 200 ml substrate/l medium). Flasks were closed with Teflon-coated stoppers and sealed. CO<sub>2</sub> was measured at various times by gas chromatography. Endogenous respiration was determined in flasks without hydrocarbon added.

**Test substance** : CAS No. 86290-81-5; a commercial unleaded gasoline topped at 76°C by distillation. It was free of hydrocarbons having less than six carbon atoms and contained no oxygenated compounds.

**Conclusion** : Activated sludge microorganisms were found to biodegrade unleaded commercial gasoline up to 94% within 25 days. For each hydrocarbon class, degradation occurred at different rates. Aromatic compounds were found to be the most readily consumed, although compounds bearing neighboring substituents and those containing longer alkyl groups were consumed at a slower rate than those with no or only one alkyl chain. Likewise, linear alkanes (exception for undecane), alkenes with five to nine carbons, cyclohexane and substituted cyclopentanes were biodegraded. Residual components of gasoline most recalcitrant to biodegradation were found to be branched alkanes, particularly those containing a quaternary carbon and/or alkyl chains on consecutive carbon atoms.

**Reliability** : (2) valid with restrictions  
21.11.2001

(29)

**Type** :  
**Inoculum** : Native soil suspension (NSS) from a spruce forest and microbially-reinforced native soil suspension (MRSS).

**Method** : Non-guideline research method using a closed-system shake flask apparatus

**Year** : 1998

**Test substance** : CAS No. 86290-81-5; artificially-prepared produced gasoline-model mixture (GM23)

**Method** : Soil biodegradation - Evaluation of biodegradability of GM23 by soil microflora; mineralization of GM23; and mineralization of 23 principal components of GM23. Method of analysis of parent mixture, individual components, and CO<sub>2</sub> production made by gas chromatography with flame ionization detector.

Exposure period: 14, 28 & 34 days

**Result** : See test condition for full description.  
The gasoline model mixture GM23 was degraded about 89% by a

### 3. Environmental Fate and Pathways

Id Gasoline  
Date 11.12.2001

native soil suspension, based on GC/FID analysis of the initial and residual individual hydrocarbon concentrations. Cyclohexane, 2,2,4-trimethylpentane and 2,3,4-trimethylpentane were only slightly biodegraded and 3-methylhexane, 2,4-dimethylhexane, and 1,3,5-trimethylbenzene were incompletely biodegraded. All other compounds were consumed by NSS. Supplementation of NSS with cyclohexane-degrading and 2,2,4-trimethylpentane-degrading microflora led to complete degradation of GM23.

Biodegradation and mineralization of GM23 at 28 days

Mineralization yields were:

MRSS inoculum: 0.58 (58%)  
NSS inoculum: 0.45 (45%) .

Kinetic Experiments with GM23

Consumption of each component by NSS did not occur at the same rate. n-alkanes and most of the mono-aromatic compounds were completely biodegraded within the first 14 days, whereas cyclohexane and di- and trimethyl alkanes were not. Methyl alkanes and dimethyl hexanes were slightly consumed over this period of time, but they were nearly totally biodegraded after 28 days. Cyclohexane and trimethyl pentanes showed little biodegradation after 28 days.

Mineralization Experiments with Individual Hydrocarbons

The mineralization yields of individual components of GM23 were high for the aromatics and alkanes with no or few methyl groups. Furthermore, the mineralization yield of GM23 determined with NSS or MRSS agreed with the mineralization data for individual components. The ability of the NSS to mineralize individual hydrocarbons indicated:

Substrate Mineralization Yield at 34 days

heptane	0.74
octane	0.49
cyclohexane	0
2-methylhexane	0.47
3-methylhexane	0.71
3-methylheptane	0.69
2,4-dimethylhexane	0.56
2,5-dimethylhexane	0.44
2,2,4-trimethylpentane	0.02
2,3,4-trimethylpentane	0
benzene	0.56
toluene	0.63
o-xylene	0.49
m-xylene	0.61
p-xylene	0.48
ethylbenzene	0.48
n-propylbenzene	0.34
2-ethyltoluene	0
3-ethyltoluene	0.38
4-ethyltoluene	0.34
1,2,3-trimethylbenzene	0
1,2,4-trimethylbenzene	0.18
1,3,5-trimethylbenzene	0

**Test condition**

: The soil sample originated from a spruce forest (Foulain, France) and contained mineral and superficial organic matter. The NSS soil suspension was prepared with 20 g of homogenized soil per liter of nutrient solution. The MRSS soil suspension contained a cyclohexane-degrading microflora

and a 2,2,4-trimethylpentane-degrading microflora obtained from gasoline-polluted sites and added at 5% v/v each to the NSS. The nutrient solution contained minerals and vitamins prepared as described in Bouchez et al. Appl. Microbiol. Biotechnol. 43:156-164 (1995).

#### Biodegradation Experiments with GM23

The biodegradation tests were performed with NSS and MRSS suspensions (50 ml) in 500-ml flasks with sidearms equipped with Mininert<sup>®</sup> valves. 25 ml of GM23 were added to 50 ml of soil suspension through the Mininert<sup>®</sup> valve with a syringe (500 ml substrate/l medium). After an incubation period of 14 or 28 days at 30°C, 5 ml of CH<sub>2</sub>Cl<sub>2</sub>, containing dodecane at 600 mg/ml as internal standard, were introduced in the flasks through the valve, and the remaining hydrocarbons were extracted for 1 h with shaking. The flasks were refrigerated overnight at 4°C before opening. The suspension was centrifuged at 4°C and 35000 g for 30 min. The CH<sub>2</sub>Cl<sub>2</sub> phase of each flask was then analyzed by gas chromatography. Mineralization yields were estimated at the end of the biodegradation experiment by recovery of CO<sub>2</sub> after acidification of the flask contents. Abiotic treatments were prepared similarly to the other treatments with the exception that 1 g/l HgCl<sub>2</sub> were added to the flasks before incubation.

#### Kinetic Experiments with GM23

Kinetics of CO<sub>2</sub> production during the degradation of GM23 were studied at 30°C for 28 days. 18 ml of NSS or MRSS were introduced into 240-ml flasks that were sealed by stoppers covered with Teflon film. 8 ml of GM23 were dispensed into the sealed flasks by syringe (444 ml substrate/l medium). A 250-ml gas-tight syringe was used to sample head space gas. Endogenous respiration of NSS and MRSS were determined under the same conditions in flasks incubated without test substance.

#### Mineralization Experiments with Individual Hydrocarbons

Mineralization yields of the individual hydrocarbon compounds were determined using 125-ml flasks containing 20 ml of soil suspension and 5 ml of each hydrocarbon (250 ml substrate/l medium). Flasks were incubated for 34 days. Two flasks for each compound and six control flasks were used.

**Test substance** : CAS No. 86290-81-5; artificially-prepared produced gasoline-model mixture (GM23) of main compounds making up a topped gasoline-cut obtained by distillation at 76°C of a French commercial unleaded gasoline. The model mixed gasoline was free of compounds lighter than C<sub>6</sub> and contained no oxygenated compounds. Biodegradation also was measured on 23 individual components of the model gasoline.

**Conclusion** : The use of optimized degradative inoculum (soil microbes pre-exposed to cyclohexane and 2,2,4 TMP) in conjunction with non-acclimated soil organisms enhanced both rate and extent of the more structurally complex hydrocarbons which showed little to minimal degradation in non-acclimated soil systems.

The results of this study indicated that the pattern of gasoline degradation was represented as the sum of the degradation of the individual compounds. No marked occurrence of co-metabolism was observed. Inhibitory effects were observed for 1,3,5 trimethylbenzene, 2-ethyltoluene and 1,2,3 trimethylbenzene at 200 mg/l, but were totally

### 3. Environmental Fate and Pathways

**Id** Gasoline  
**Date** 11.12.2001

**Reliability**

16.11.2001

: degraded at 35 mg/l by non-acclimated soil suspensions.  
(2) valid with restrictions acceptable, well-documented  
publication which meets basic scientific principles

(28)

## 4. Ecotoxicity

Id Gasoline  
Date 11.12.2001

### 4.1 ACUTE/PROLONGED TOXICITY TO FISH

**Type** : static  
**Species** : Oncorhynchus mykiss (Fish, fresh water)  
**Exposure period** : 96 hour(s)  
**Unit** : mg/l  
**Analytical monitoring** : yes  
**Method** : OECD Guide-line 203 "Fish, Acute Toxicity Test"  
**Year** : 1995  
**GLP** : yes  
**Test substance** : Gasoline CAS No. 86290-81-5  
**Method** : LL<sub>50</sub> at 96 hr calculated using binomial probability analysis. ASTM Special Technical Publication 634. 1977, pp 65-84  
**Result** : Mortality (no. of deaths/treatment) at 96 hrs:

Treatment (mg/l)	No. of deaths
0	0
0.1	0
1.0	0
5.0	0
10	0
25	15

96-hr LL<sub>50</sub> = 16 mg/l, 99% C.I.: 10-25 mg/l (as nominal loading rate)  
96-hour No Observed Effect Loading (NOEL) was 10 mg/l, based on mortality, both calculated (Dunnett's Procedure) and observed.

Results are quoted in terms of 50% Lethal Loading (LL<sub>50</sub>), the loading rate of test substance resulting in 50% mortality of the test species exposed to the WAF. At termination, loss of equilibrium was observed in all surviving fish at the 10 mg/l treatment.

#### Analytical results

Losses of the soluble BETXN components from the WAF over each 24 hour period ranged from 0 to 8% for the 1.0, 5, 10 and 25 mg/l loadings. Up to 100% loss was observed in the 0.1 mg/l treatment in 24 hrs samples.

#### Analytical results

Day	Control	Measured BTEXN (mg/l)				
		Nominal loading rate (mg/l)				
		0.1	1.	5.0	10	25
0 (new)	ND	0.12	0.31	1.7	3.1	7.7
1 (old)	ND	0.12	0.41	1.6	3.3	7.1
1 (new)	ND	0.16	0.44	1.7	1.9	6.5
2 (old)	ND	0.15	0.45	1.6	2.1	6.8
2 (new)	ND	0.07	0.43	1.6	3.2	NA
3 (old)	ND	0.12	0.43	1.6	3.1	NA
3 (new)	ND	0.16	0.57	1.8	3.3	NA
4 (old)	ND	ND	0.56	1.8	3.5	NA

ND=not detected, NA=not analyzed due to 100% mortality

Guideline/protocol deviations: Body length smaller than recommended range of 4-6 cm; smaller fish used to minimize DO depletion in closed vessel (no-headspace) systems.

**Test condition** : Test solutions were prepared as water accommodated fractions

## 4. Ecotoxicity

**Id** Gasoline  
**Date** 11.12.2001

(WAF). The control and dilution water was a laboratory blend water prepared from carbon filtered well water and ion exchange softened, reverse osmosis dialyzed well water aged for >24 hrs. To determine contact time required to achieve maximum solubility between test substance and aqueous solution, samples of WAF test solutions equilibrated for 20, 24 27 and 49 hours at 100 mg/l loading were analyzed by GC-FID for concentrations of the following: benzene, toluene, ethylbenzene, xylene isomers and naphthalene (BTEXN). Maximum WAF solubility for these components reached equilibrium within 24 hrs of stirring. Nominal loading rates of 0, 0.1, 1, 5, 10, and 25 mg/l were used to prepare test solutions for the fish toxicity tests. Test substance, added volumetrically, was mixed for each individual treatment in dilution water for 24 hours in 20 liter stoppered containers with less than 10% headspace volume. The mixtures were allowed to settle for 1-2 hours prior to drawing off the aqueous solutions for testing. Fish were approximately five weeks old at test initiation and were obtained from Thomas Fish company, Anderson, CA, Lot 297. Loading of fish body mass to treatment was 0.3 g fish per liter of aqueous solution, mean length at termination was 3.3 cm (sd=0.2), and mean weight was 0.271 g (sd=0.064). Test vessels were 4 liter glass aspirator bottles with foil covered neoprene stoppers. Three replicates per treatment and 5 organisms per replicate were tested for each treatment and the control. Exposure containers were filled ( no headspace volume) and tightly sealed to prevent volatilization. Test solution renewal was performed daily by removing 80% of the test solution and replacing it with fresh WAF solution prepared at least 24 hrs prior to use. Freshly prepared and old WAF test solutions were analyzed by GC-FID for concentrations of BTEXN. Water temperature was 15 °C (0.1sd). Test photoperiod was 16 hrs. light and 8 hr dark, light intensity approx 609-614 Lux during full daylight periods. Dissolved oxygen measurements ranged from 5.4 to 9.7 ppm, pH values between 6.8 and 8.2.

**Test substance** : Gasoline Sample W94/814, Blend; Detailed hydrocarbon analysis:  
N-paraffins: 16% total C4-C8  
Iso-paraffins: 25% total C4-C11  
Olefins: 12%, C4-C7  
Naphthenes: 5% C6-C10  
Aromatics: 42% C6-C11

**Reliability** : (1) valid without restriction  
21.11.2001

(31)

**Type** : static  
**Species** : Oncorhynchus mykiss (Fish, fresh water)  
**Exposure period** : 96 hour(s)  
**Unit** : mg/l  
**Analytical monitoring** : yes  
**Method** : OECD Guide-line 203 "Fish, Acute Toxicity Test"  
**Year** : 1995  
**GLP** : yes  
**Test substance** : Gasoline CAS No. 86290-81-5  
**Method** : LL<sub>50</sub> at 96 hr calculated using Probit procedure (Finney, D.J., 1971. Probit Analysis, Third Edition, London: Cambridge University Press, and SAS computer statistics software.

## 4. Ecotoxicity

**Id** Gasoline  
**Date** 11.12.2001

### Result

Mortality (no. of deaths/treatment) at 96 hrs:

Treatment	No. of deaths
0	1
1.0	0
5	0
10	7
25	15
50	5

96-hr  $LL_{50}$  = 11 mg/l, 95% C.I: 8.7-16 mg/l (as nominal loading rate)  
96-hour No Observed Effect Loading (NOEL) was 5 mg/l, both calculated (Dunnett's Procedure) and observed.  
Results are quoted in terms of 50% Lethal Loading ( $LL_{50}$ ), the loading rate of test substance resulting in 50% mortality of the test species exposed to the WAF. At termination, abnormal behavior/appearance (lethargy, erratic swimming) was observed in all surviving fish at the 10 mg/l treatment. Losses of the soluble components from the WAF over each 24 hour period ranged from 5 to 25% for the 5, 10 and 25 mg/l loadings. Up to 57% loss was observed in the 1.0 mg/l treatment in 24 hrs samples. BTEXN concentrations on 24hour samples of the 50 mg/l treatments due to complete mortality on day 0 were not determined.

Day	Analytical results Measured BTEXN (mg/l)					
	Nominal loading rate (mg/l)					
	Control	1.0	5.0	10	25	50
0 (new)	ND	0.54	2.3	4.2	9.5	20
1 (old)	ND	0.50	2.3	4.0	10	NA
1 (new)	ND	0.47	1.7	4.2	NA	NA
2 (old)	ND	0.20	2.1	4.0	NA	NA
2 (new)	ND	0.52	2.0	4.1	NA	NA
3 (old)	ND	0.25	2.0	4.3	NA	NA
3 (new)	ND	0.57	1.6	4.0	NA	NA
4 (old)	ND	0.38	1.2	3.2	NA	NA

ND=not detected, NA=not analyzed due to 100% mortality

Guideline/protocol deviations: Body length (2.7cm av.) smaller than recommended range of 4-6 cm; smaller fish used to minimize DO depletion in closed vessel (no-headspace) systems.

### Test condition

: Test solutions were prepared as water accommodated fractions (WAF). The control and dilution water was a laboratory blend water prepared from carbon filtered well water and ion exchange softened, reverse osmosis dialyzed well water aged for >24 hrs. To determine contact time required to achieve maximum solubility between test substance and aqueous solution, samples of WAF test solutions equilibrated for 20, 24 27 and 49 hours at 100 mg/l loading were analyzed by GC-FID for concentrations of the following: benzene, toluene, ethylbenzene, xylene isomers and naphthalene (BTEXN). Maximum WAF solubility for these components reached equilibrium within 24 hrs of stirring. Nominal loading rates of 0, 1, 5, 10, 25 and 50 mg/l were used to prepare test solutions for the fish toxicity tests. Test substance, added volumetrically, was mixed for each individual treatment in dilution water for 24 hours in 20 liter stoppered containers with less than 10% headspace volume. The mixtures were allowed to settle for 1-2 hours prior to drawing off the aqueous solutions for testing. Fish

## 4. Ecotoxicity

**Id** Gasoline  
**Date** 11.12.2001

were approximately four weeks old at test initiation and were obtained from Thomas Fish company, Anderson, CA, Lot 297. Loading of fish body mass to treatment was 0.2 g fish per liter of aqueous solution, mean length at termination was 2.7 cm (sd=0.2), and mean weight was 0.136 g (sd=0.034). Test vessels were 4 liter glass aspirator bottles with foil covered neoprene stoppers. Three replicates per treatment and 5 organisms per replicate were tested for each treatment and the control. Exposure containers were filled (no headspace volume) and tightly sealed to prevent volatilization. Test solution renewal was performed daily by removing 80% of the test solution and replacing it with fresh WAF solution prepared at least 24 hrs prior to use. Freshly prepared and old WAF test solutions were analyzed by GC-FID for concentrations of BTEXN. Water temperature was 14.1 °C (0.03sd). Test photoperiod was 16 hrs. light and 8 hr dark, light intensity approx 619-622 Lux during full daylight periods. Dissolved oxygen measurements ranged from 7.4 to 9.8 ppm, pH values between 7.8 and 8.1.

**Test substance** : Gasoline Sample W94/813, Blend

Detailed hydrocarbon analysis:  
N-paraffins: 20% total C3-C8,  
Iso-paraffins: 28% total C4-C9  
Olefins: 1%, C5-C7  
Naphthenes: 5% C5-C10  
Aromatics: 46% C6-C9

**Reliability** : (1) valid without restriction  
21.11.2001

(12)

### 4.2 ACUTE TOXICITY TO AQUATIC INVERTEBRATES

**Type** : Acute immobilization test  
**Species** : Daphnia magna (Crustacea)  
**Exposure period** : 48 hour(s)  
**Unit** : mg/l  
**Analytical monitoring** : yes  
**Method** : OECD Guide-line 202, part 1 "Daphnia sp., Acute Immobilisation Test"  
**Year** : 1995  
**GLP** : yes  
**Test substance** : Gasoline CAS No. 86290-81-5  
**Method** : EL<sub>50</sub> calculated using the probit procedure (Finney, D.J., 1971. Probit Analysis, 3rd Ed. London: Cambridge Univ. Press)  
**Result** : 48 hr results-number of organisms affected and analytical results

Treatment	Measured	
	Immobilization	BTEXN
Control	2	ND
0.1 mg/l	1	0.12
1.0 mg/l	1	0.31
5.0 mg/l	1	1.7
10 mg/l	5	3.1
25 mg/l	20	7.7

based upon nominal loading rate 48-hr EL<sub>50</sub> = 12 mg/l (95% C.I. 7.3 to 22 mg/l)

**Test condition** : Test solutions were prepared as water accommodated fractions (WAF). The control and dilution water was a laboratory blend

## 4. Ecotoxicity

**Id** Gasoline  
**Date** 11.12.2001

water prepared from carbon filtered well water and ion exchange softened, reverse osmosis dialyzed well water aged for >24 hrs. To determine contact time required to achieve maximum solubility between test substance and aqueous solution, samples of WAF test solutions equilibrated for 20, 24, 27 and 49 hours at 100 mg/l loading were analyzed by GC-FID for concentrations of the following: benzene, toluene, ethylbenzene, xylene isomers and naphthalene (BTEXN). Maximum WAF solubility for these components reached equilibrium within 24 hrs of stirring. Nominal loading rates of 0, 0.1, 1, 5, 10, and 25 mg/l were used to prepare test solutions for the toxicity tests. Test substance, added volumetrically, was mixed for each individual treatment in dilution water for 24 hours in 4 liter stoppered containers with less than 10% headspace volume. The WAF mixtures were allowed to settle for 1-2 hours prior to drawing off the aqueous solutions for testing. WAF test solutions were analyzed by GC-FID for concentrations of BTEXN on day 0 and at termination. Test vessels for daphnid testing were 125 ml glass erlenmeyer flasks with foil covered neoprene stoppers. Four replicates per treatment and 5 organisms per replicate were tested for each treatment and the control. Exposure containers were filled (no headspace volume) and tightly sealed to prevent volatilization. During the study test system solutions: dissolved oxygen concentration range: 7.2 to 9.2; pH ranged from 7.5 to 7.8; temperature was 19 °C (sd:0.2). Daphnia magna were supplied by testing laboratory; age < 24 hours old; obtained from culture maintained in-house.

**Test substance** : Gasoline Sample W94/814, Blend  
Detailed hydrocarbon analysis:  
N-paraffins: 16% total C<sub>4</sub>-C<sub>8</sub>  
Iso-paraffins: 25% total C<sub>4</sub>-C<sub>11</sub>  
Olefins: 12% C<sub>4</sub>-C<sub>7</sub>  
Naphthenes: 5% C<sub>6</sub>-C<sub>10</sub>  
Aromatics: 42% C<sub>6</sub>-C<sub>11</sub>

**Reliability** : (2) valid with restrictions. Three previous attempts to conduct study were invalidated due to excessive (>20%) control mortality.

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(11)

**Type** : Acute immobilization test  
**Species** : Daphnia magna (Crustacea)  
**Exposure period** : 48 hour(s)  
**Unit** : mg/l  
**Analytical monitoring** : yes  
**Method** : OECD Guide-line 202, part 1 "Daphnia sp., Acute Immobilisation Test"  
**Year** : 1995  
**GLP** : yes  
**Test substance** : Gasoline CAS No. 86290-81-5  
**Method** : EL<sub>50</sub> calculated using the probit procedure (Finney, D.J., 1971. Probit Analysis, 3rd Ed. London: Cambridge Univ. Press)  
**Result** : 48 hr results-number of organisms affected and analytical results

Treatment	Immobilization	Measured BTEXN-day 0	Measured BTEXN-day 2
Control	0	ND	ND
0.5 mg/l	0	0.29	0.10
1.0 mg/l	0	0.28	0.10
5.0 mg/l	3	2.3	1.7
10 mg/l	16	3.9	3.1



## 4. Ecotoxicity

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### Result

- calculated using ANOVA (Duncan D.B., 1975, Biometrics, 31, 339-359)
- : Percent inhibition:
    - 72 hour EL50 for average growth rate=3.1 mg/l  
(0.15 to >25 mg/l CI @95%)
    - 72 hour EL50 for area under the growth curve=1.4 mg/l  
(0 to 20 mg/l CI @95%)
    - 96 hour EL50 for average growth rate=3.7 mg/l  
(0.34 to >25 mg/l CI @95%)
    - 96 hour EL50 for area under the growth curve=1.1 mg/l  
(0 to 22 mg/l CI @95%)
    - 72 hour NOEL for average growth rate=0.5 mg/l
    - 72 hour NOEL for area under the growth curve =<0.5 mg/l
    - 96 hour NOEL for average growth rate =1.0 mg/l
    - 96 hour NOEL for area under the growth curve =<0.5 mg/l

Nominal (mg/l)	Average cell density (cells/ml)		% Inhibition Average growth rate		Area under growth curve	
	72hr	96hr	72hr	96 hr	72hr	96hr
Control	1.6E5	3.9 E5	0	0	0	0
0.5	1.0E5	2.8E5	6.2	3.4	36	33
1.0	6.8E4	2.6 E5	12	5.5	51	44
5.0	4.6E4	1.2E5	21	22	53	65
10	2.4E3	3.7E3	90	89	93	98
25	BMDL	BMDL	99	99	98	99

BMDL=below method detection limit

### Analytical results

Nominal (mg/l) Measured Concentration (mg/l as BTEXN)

	Day0	Day 4
	Control	none detected
0.5	0.12	0.30
1.0	0.58	0.67
5.0	2.4	0.65
10	5.2	4.72
5	12	9.6

### Test condition

- : Individual test treatment solutions were prepared as Water Accommodated Fractions (WAFs). To determine contact time required to achieve maximum solubility between test substance and aqueous solution, samples of WAF test solutions equilibrated for 20, 24, 27 and 49 hours at 100 mg/l loading were analyzed by GC-FID for concentrations of the following: benzene, toluene, ethylbenzene, xylene isomers and naphthalene (BTEXN). Maximum WAF solubility for these components reached equilibrium within 24 hrs of stirring. Nominal loading rates of 0, 0.5, 1, 5, 10, and 25 mg/l were used to prepare test solutions for the algal toxicity tests. Test material was added volumetrically to 2.0 liters of sterilized algal nutrient media (enriched with 100 mg/l of sodium bicarbonate) in 2.0 liter aspirator bottles covered with aluminum foil. The mixing vessels were sealed with foil covered stoppers and mixed on magnetic stir plates with teflon coated stir bars for approximately 24 hours at room temperature. After mixing the solutions were allowed to settle for one hour and the WAF was removed and used for testing. Test vessels were 125ml glass Erlenmeyer flasks containing ten 4mm glass balls that were completely

## 4. Ecotoxicity

**Id** Gasoline  
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filled (140 ml) with treatment solution, inoculated with algae and sealed with glass stoppers. Algal cells were obtained from 6 day old laboratory stock cultures maintained in nutrient enriched media, at 24 °C ( $\pm 2^\circ$ ) C under continuous illumination of 4300( $\pm 10\%$ ) lux. Original algal cultures (Strain 1648) were provided by the Department of Botany, University of Texas. Cell density of the algal stock culture inoculum was determined prior to study initiation with a Turner filter-fluorometer. Fluorometer readings were converted to cell numbers using a regression formula developed through cell counts. Three replicates were prepared for each treatment level and six replicates were prepared as control systems. The initial algal concentration was approximately  $1.0 \times 10^3$  cells/ml in each replicate chamber. All test replicates were placed on a shaker table at 150 oscillations per minute during the study and exposed to continuous fluorescent light, illumination at 4300 to 4400 Lux as measured daily using a Licor photometric sensor. A sample volume of 3.5 ml was taken daily for density determinations, and an equivalent volume of reserve 24 hour WAF was used to replenish the displaced sample volume. WAF test solutions were analyzed by GC-FID for concentrations of BTEXN at day 0 and 96 hr termination. BTEXN total concentration at termination was at least 80% of the initial concentration for all treatments, with the exception of the 5.0 mg/l exposure, which showed a loss of 73%. This excessive loss compared to the other treatments was determined to be due to sampling technique. Test temperature was 23 °C (sd=0.08)C. The average pH was 7.5 at initiation; and ranged from 9.2 (control) to 7.8 (25 mg/l loading) at termination.

**Test substance** : Gasoline Sample W94/813, Blend

Detailed hydrocarbon analysis:

N-paraffins: 20% total C3-C8

Iso-paraffins: 28% total C4-C9

Olefins: 1% C5-C7

Naphthenes: 5% C5-C10

Aromatics: 46% C6-C9

**Reliability** : (1) valid without restriction  
21.11.2001

(8)

**Species** : Selenastrum capricornutum (Algae)

**Endpoint** : growth rate

**Exposure period** : 96 hour(s)

**Unit** : mg/l

**Analytical monitoring** : yes

**Method** : OECD Guide-line 201 "Algae, Growth Inhibition Test"

**Year** : 1995

**GLP** : yes

**Test substance** : Gasoline CAS No. 86290-81-5

**Method** : EL<sub>50</sub> values were calculated using the inverse extrapolation method of Snedecor and Cochran, Statistical Methods, 8th Ed., 1989, Iowa State University Press/Ames. NOEL values calculated using ANOVA (Duncan D.B., 1975, Biometrics, 31, 339-359).

**Result** : Percent inhibition:

72 hour EL<sub>50</sub> for average growth rate=3.3 mg/l  
(0.24 to >25 mg/l CI @95% )

72 hour EL<sub>50</sub> for area under the growth curve=4.2 mg/l  
(0 to 24 mg/l CI @95%)

96 hour EL<sub>50</sub> for average growth rate=2.5 mg/l  
(0.62 to 14 mg/l CI @95%)

## 4. Ecotoxicity

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96 hour EL<sub>50</sub> for area under the growth curve=0.25 mg/l  
(0 to 26 mg/l CI @95%)  
72 hour NOEL for average growth rate and area under the  
growth curve =0.5 mg/l  
96 hour NOEL for average growth rate = 0.5 mg/l  
96 hour NOEL for area under the growth curve =<0.5 mg/l

Nominal (mg/l)	Avg cell density (cells/ml)		% Inhibition		area under growth curve	
	72hr	96hr	72hr	96 hr	72hr	96 hr
	Control	9.9E4	3.8 E5	0	0	0
0.5	7.7E4	2.6 E5	7.7	7.8	21	27
1.0	5.5E4	1.7 E5	15	17	36	50
5.0	2.5E4	2.2 E4	33	51	54	81
10	3.7E3	2.0 E3	76	95	90	97
25	BMDL	BMDL	99	100	98	99

BMDL=below method detection limit

### Analytical results

Nominal (mg/l)    Measured Concentration (mg/l as BTEXN)

	Day 0	Day 4
Control	none detected	none detected
0.5	0.22	0.23
1.0	0.47	0.51
5.0	1.5	1.3
10	3.5	3.3
25	9.5	7.7

**Test condition** : Individual test treatment solutions were prepared as Water Accommodated Fractions (WAFs). To determine contact time required to achieve maximum solubility between test substance and aqueous solution, samples of WAF test solutions equilibrated for 20, 24, 27 and 49 hours at 100 mg/l loading were analyzed by GC-FID for concentrations of the following: benzene, toluene, ethylbenzene, xylene isomers and naphthalene (BTEXN). Maximum WAF solubility for these components reached equilibrium within 24 hrs of stirring. Nominal loading rates of 0, 0.5, 1, 5, 10, and 25 mg/l were used to prepare test solutions for the algal toxicity tests. Test material was added volumetrically to 2.0 liters of sterilized algal nutrient media (enriched with 100 mg/l of sodium bicarbonate) in 2.0 liter aspirator bottles covered with aluminum foil. The mixing vessels were sealed with foil covered stoppers and mixed on magnetic stir plates with teflon coated stir bars for approximately 24 hours at room temperature. After mixing the solutions were allowed to settle for one hour and the WAF was removed and used for testing. Test vessels were 125ml glass Erlenmeyer flasks containing ten 4mm glass balls that were completely filled (140 ml) with treatment solution, inoculated with algae and sealed with glass stoppers. Algal cells were obtained from 5 day old laboratory stock cultures maintained in nutrient enriched media, at 24 °C (±2°) under continuous illumination of 4300(±10%) lux. Original algal cultures (Strain 1648) were provided by the Department of Botany, University of Texas. Cell density of the algal stock culture inoculum was determined prior to study initiation with a Turner filter-fluorometer. Fluorometer readings were converted to cell numbers using a regression formula

## 4. Ecotoxicity

**Id** Gasoline  
**Date** 11.12.2001

developed through cell counts. Three replicates were prepared for each treatment level and six replicates were prepared as control systems. The initial algal concentration was approximately  $1.0 \times 10^3$  cells/ml in each replicate chamber. All test replicates were placed on a shaker table at 150 oscillations per minute during the study and exposed to continuous fluorescent light, illumination at 4300 to 4400 Lux as measured daily using a Licor photometric sensor. A sample volume of 3.5 ml was taken daily for density determinations, and an equivalent volume of reserve 24 hour WAF was used to replenish the displaced sample volume. WAF test solutions were analyzed by GC-FID for concentrations of BTEXN at day 0 and 96 hr termination. BTEXN total concentration at termination was at least 80% of the initial concentration. Test temperature was 23 °C (sd=0.08). The average pH was 7.6 at initiation; and ranged from 9.5 (control) to 8.1 (25 mg/l loading) at termination.

**Test substance** : Gasoline Sample W94/814, Blend  
Detailed hydrocarbon analysis:  
N-paraffins: 16% total C4-C8  
Iso-paraffins: 25% total C4-C11  
Olefins: 12% C4-C7  
Naphthenes: 5% C6-C10  
Aromatics: 42% C6-C11

**Reliability** : (1) valid without restriction  
21.11.2001

(9)

**5.1.1 ACUTE ORAL TOXICITY**

**Type** : LD<sub>50</sub>  
**Species** : rat  
**Strain** : Sprague-Dawley  
**Sex** : male/female  
**Number of animals** : 10  
**Vehicle** : undiluted  
**Value** : = 18.75 ml/kg bw  
**Year** : 1980  
**GLP** : yes  
**Test substance** : API PS-6  
**Method** :

Groups of 10 fasted rats (five male and five female) were given API PS-6 at doses of 10, 15, 17.5, 20 and 25 ml/kg as a single oral dose. The animals were then allowed food and water ad libitum and were observed hourly for clinical signs for the first 6 hours after dosing. Observation was twice daily thereafter for 14 days. Body weights were recorded at 7 and 14 days after administration of test material. At the end of the study, the animals were killed and subjected to a gross necropsy and any abnormalities were recorded. [In addition 2 extra males and one female were given 15 ml/kg because 3 of the original animals died soon after dosing and this was believed to be due to dosing injury. However, at necropsy, no evidence of injury was found and therefore all animals were included in the calculations for an LD50]

**Result** : Toxic signs were the same in all dose groups, increasing in severity with increasing dose. There were oily urine stains, but most of the test material seemed to be excreted via the feces. The area around the anus became very irritated. Diarrhea was common in each dose level and blood was commonly seen around the eyes, nose and mouth. Observations at necropsy were similar for all dose groups. Animals surviving to 14 days had very few abnormalities and these were usually of a minor nature such as enlarged Peyer's patches on the intestines. There were numerous instances of lung involvement in both surviving animals and those dying before 14 days. These changes consisted of mild irritation and congestion, to fluid filled abscesses. Almost all animals that died before 14 days had intestinal damage. The intestines, and often the stomach, became hemorrhagic and sometimes blood was observed in the intestine or stomach. The intestine wall became thin and there was an increased amount of gas in the gastro intestinal tract. The heart was enlarged or irregularly shaped in some rats.

Mortality and body weight changes are summarized in the following table.

Dose group (ml/kg)	Mortality dying/dosed	Weight change (over 14days)
Males		
10	0/5	weight gain
15	2/7	weight loss
17.5	3/5	weight loss
20	4/5	weight loss
25	5/5	weight loss

## 5. Toxicity

**Id** Gasoline  
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Females		
10	0/5	weight gain
15	1/6	weight gain
17.5	3/5	weight loss
20	0/5	weight gain*
25	4/5	weight loss

\* one animal had a weight loss over the 14 day period.

The oral LD<sub>50</sub> was determined to be 18.75 ml/kg. The 95% confidence limits were 16.3 to 21.6 ml/kg.

**Reliability** : (1) valid without restriction (5)  
19.11.2001

### 5.1.3 ACUTE DERMAL TOXICITY

**Type** : LD<sub>50</sub>  
**Species** : rabbit  
**Strain** : New Zealand white  
**Sex** : male/female  
**Number of animals** : 4  
**Vehicle** : undiluted  
**Value** : > 5 ml/kg bw  
**Year** : 1979  
**GLP** : yes  
**Test substance** : API PS-6  
**Method** : The skin of the patched area of two rabbits of each sex had been abraded whilst the other two had intact skin. A single dose of undiluted test material (5 ml/kg) was applied to the dorsal skin of each of 4 male and 4 female rabbits. The applied dose was covered with an occlusive dressing (gauze and an impermeable covering). 24 hours after dosing, the patches were removed, the skin wiped and collars fitted to the rabbits to prevent oral intake of any residual test material. The collars were removed 24 hours later and the animals were observed for a total of 14 days post-dosing. At study termination the animals were killed with carbon dioxide and a gross necropsy was performed. Any abnormalities were recorded.

**Result** : When the patches were removed following dosing dark red to almost purple skin was seen in all animals. Slight erythema and dry skin was observed in all rabbits during the study. With the exception of one animal all animals weighed more at the end of the study than they did at study commencement. One female rabbit died on day 6 of the 14 day study and the gross necropsy revealed slightly congested lungs, no food in the stomach and white areas in the liver. At necropsy of the surviving animals four rabbits had congested lungs, one had pale kidneys, one had an irritated stomach lining and one had enlarged Peyer's patches on the jejunum. These observations were considered to be normal and not dose-related.

**Reliability** : (1) valid without restriction (5)  
19.11.2001

## 5. Toxicity

**Id** Gasoline  
**Date** 11.12.2001

### 5.1.4 ACUTE TOXICITY, OTHER ROUTES

**Remark** : Not appropriate  
04.07.2001

### 5.2.1 SKIN IRRITATION

**Species** : rabbit  
**Concentration** : undiluted  
**Exposure** : Occlusive  
**Exposure time** : 24 hour(s)  
**Number of animals** : 6  
**PDII** : .98  
**Result** : slightly irritating

**Method** : Draize Test  
**Year** : 1979  
**GLP** : yes  
**Test substance** : API PS-6  
**Method** :

0.5 ml of undiluted test material was applied to the shorn skin in two areas on each of 3 male and 3 female rabbits. One area was intact and the other abraded skin. The treated area was then covered with an occlusive dressing. After 24 hours the dressing was removed and the treated skin was wiped to remove any residue of test material. The degree of erythema and edema was recorded according to the Draize scale. A second reading of skin responses was made at 72 hours and again at 96 hours, 7 and 14 days. Results of the 24 and 72 hour readings were used to determine the Primary Irritation Index.

**Result** : A summary of the dermal irritation scores is given below.

	Exposure time (hours)	Average value of all animals
Erythema		
Intact skin	24	0
	72	0.92
Abraded skin	24	0
	72	1.0
Edema		
Intact skin	24	0.5
	72	0.5
Abraded skin	24	0.5
	72	0.5
		3.92
Primary irritation score =Total÷4=		0.98

Edema but no erythema was noted at 24 hours, although the test area was whiter than the surrounding skin.  
At 72 hours erythema and edema were observed.  
By 7 days almost all erythema had cleared but some edema was still present and the test site was dry and flaky.  
By day 14 all edema and erythema had cleared but there was no hair growth at this time.

**Reliability** : (1) valid without restriction  
19.11.2001

(5)

## 5. Toxicity

Id Gasoline  
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### 5.2.2 EYE IRRITATION

<b>Species</b>	:	rabbit
<b>Concentration</b>	:	undiluted
<b>Dose</b>	:	.1 ml
<b>Exposure Time</b>	:	See method
<b>Comment</b>	:	rinsed after (see exposure time)
<b>Number of animals</b>	:	9
<b>Result</b>	:	not irritating
<b>Method</b>	:	Draize Test
<b>Year</b>	:	1979
<b>GLP</b>	:	yes
<b>Test substance</b>	:	API PS-6
<b>Method</b>	:	0.1 ml of undiluted test material was applied to the corneal surface of one eye of each of 9 rabbits (4 male, 5 female), the other eye was untreated and served as control. After 20 to 30 seconds the treated eyes of 3 rabbits were washed with lukewarm water for 1 minute. Eyes of the other 6 rabbits were not washed. Readings of ocular lesions for all animals were made at 1, 24, 48, 72 hours and 7 days after treatment. Sodium fluorescein was used to aid in revealing possible corneal injury.
<b>Result</b>	:	No irritation was observed in any animal at any of the three observation times. Animals whose eyes had been irrigated following instillation of test material were no different from those whose eyes had not been washed.
<b>Reliability</b>	:	(1) valid without restriction
19.11.2001		(5)

### 5.3 SENSITIZATION

<b>Type</b>	:	Buehler Test
<b>Species</b>	:	guinea pig
<b>Concentration</b>	:	Induction 50 % occlusive epicutaneous Challenge 50 % occlusive epicutaneous
<b>Number of animals</b>	:	10
<b>Vehicle</b>	:	mineral oil
<b>Result</b>	:	not sensitizing
<b>Year</b>	:	1979
<b>GLP</b>	:	yes
<b>Test substance</b>	:	API PS-6
<b>Method</b>	:	0.5 ml of undiluted test material was applied under an occlusive dressing to the shorn skin of 10 male and 10 female animals. 6 hours after application, the dressings were removed and the skin wiped to remove residues of test material. After the first application, irritation was sufficiently severe that for further dosing a 50% dilution in mineral oil was used. The animals received one application 3 times each week for 3 weeks. The same application site was used each time. 2 weeks following the third application a challenge dose (0.5 ml of a 50% dilution in mineral oil) was applied in the same manner as the sensitizing doses. A previously untreated site was used for the challenge application. The application sites for sensitizing and challenge doses were read for erythema and edema 24 and 48 hours after patch removal. To assist in the reading of the response to the final challenge dose the

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- test site was depilated 3 hours prior to reading by using a commercially available depilatory cream.
- Positive control (0.05% 2,4-dinitrochlorobenzene in ethanol), vehicle control and naive control groups were included in this study and the procedure for these was the same as for the test groups.
- Result** : On a subjective basis, the challenge treatment did not appear to be more reactive than the sensitizing treatments. The average scores for erythema and edema following induction and challenge are summarized below.
- | Average scores | PS-6 gasoline Erythema | Edema | Positive control Erythema | Edema |
|----------------|------------------------|-------|---------------------------|-------|
| Induction      | 0.9                    | 0.3   | 1.3                       | 0.3   |
| Challenge      | 0.1                    | 0     | 1.9                       | 1.7   |
- Reliability** : The authors concluded that the test material was not sensitizing. (2) valid with restrictions  
Although the study was conducted to GLP, there was no vehicle control and the results from the positive control were not convincing.
- 21.11.2001 (5)

### 5.4 REPEATED DOSE TOXICITY

- Species** : Rats and monkeys  
**Sex** : male/female  
**Strain** : Sprague Dawley rats and squirrel monkeys  
**Route of admin.** : inhalation  
**Exposure period** : 6 hours each day  
**Frequency of treatment** : 5 days a week for 13 weeks  
**Post obs. period** :  
**Doses** : leaded gasoline: 0, 100 & 400 ppm. Unleaded gasoline: 0, 400 & 1500 ppm  
**Control group** : yes, concurrent no treatment  
**Year** : 1984  
**GLP** : no data  
**Test substance** : leaded and unleaded gasoline  
**Method** : This study was conducted as a preliminary range finding study prior to conducting a two year study on the same test materials.

20 rats and 4 monkeys of each sex were housed in 1m<sup>3</sup> glass and stainless steel exposure chambers 24 hours a day and were only removed for cleaning purposes. Target exposure vapor concentrations of the test materials were:  
Unleaded gasoline: 400 and 1500 ppm  
Leaded gasoline: 100 and 400 ppm  
A control group of 20 rats and 4 monkeys of each sex were exposed to air only.  
Exposures were for 6 hours each day, 5 days each week for 13 weeks.

Blood was taken from 10 rats of each sex at the end of the study from the highest dose groups only for hematological evaluation. Blood was taken from all monkeys in the highest dose group at 1.5, and 3 months.

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Urine samples were analyzed for all animals at 1.5 and 3 months for levels of protein, glucose, ketones, bilirubin, blood and lead.

CNS evaluations were conducted on the monkeys in the control and high level dose groups at before exposure and at 3 months. The CNS evaluations consisted of recording simultaneous and evoked responses and this was accomplished using electrodes that had been implanted permanently in the visual cortex.

Pulmonary function tests similar to those reported by Alarie were conducted on all monkeys prior to exposure and at 1.5 and 3 months on the control and high level unleaded groups. All animals that died or were sacrificed at termination of the study were subjected to a gross necropsy. Organ weights were recorded and lungs, kidneys, spleen, heart, brain and bone marrow from the control and high dose groups were evaluated for histopathology.

All male and female animals from the control and high exposure groups were also evaluated for the presence of IgG in the renal glomerulus and lungs.

A lead analysis was also made on rat brain, kidney, liver, urine and blood from both the leaded dose groups and controls.

### Result

: The mean exposure concentrations were found to be as follows:

Group	Gasoline vapor exposure concentration		
	Mg/l ±SD	ppm	Alkyl lead µg Pb/l ±SD
Control	-	-	-
Unleaded 1500 ppm	6.35±0.44	1552	-
Unleaded 400 ppm	1.57±0.15	384	-
Leaded 400 ppm	1.53±0.23	374	0.72±0.1
Leaded 100 ppm	0.42±0.04	103	0.19±0.04

Three rats at different dose levels and three monkeys also at different dose levels died during the study. These deaths were not considered to be treatment-related.

Two female monkeys in each of the high dose groups exhibited emesis, 13 and 17 days after commencing exposure for the 1500 ppm unleaded and 400 ppm leaded groups respectively. Although there was a reduction in body weights in males in the lowest dose group of each of the test materials but by the end of the study they were demonstrating increased weights. No differences were observed in any of the other treated groups.

The hematological values for the monkeys exposed to either test material at either dose level were similar to those of the control animals. In the rats the only changes observed were:

unleaded (1500 ppm males)	64% increase in thrombocytes
unleaded (1500 ppm females)	150% increase in reticulocytes
leaded (400 ppm males)	4% decrease in MCHC
leaded (400 ppm females)	10% increase in hematocrit
leaded (400 ppm females)	11% increase in MCV
leaded (400 ppm females)	decrease in WBC

Mean flash-evoked response time for the monkeys was measured prior to exposure and was unaffected by exposure.

## 5. Toxicity

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The results of the mean pulmonary function data are summarised in the following table. Only increases (% I) or decreases (% D) compared to controls are shown in the table. All other parameters were similar for treated and control animals.

	Pre-exposure	42 days	90 days
<b>Respiratory rate</b>			
Unleaded 1500 ppm F	-	-	-
Unleaded 1500 ppm M	30% D	21% D	-
Leaded 400 ppm F	-	-	-
Leaded 400 ppm M	-	-	-
<b>Tidal volume</b>			
Unleaded 1500 ppm F	-	-	22% D
Unleaded 1500 ppm M	-	-	-
Leaded 400 ppm F	-	-	-
Leaded 400 ppm M	-	-	-
<b>Minute volume</b>			
Unleaded 1500 ppm F	-	-	-
Unleaded 1500 ppm M	-	-	36% I
Leaded 400 ppm F	-	-	-
Leaded 400 ppm M	-	-	53% I

There were no effects on airway resistance, dynamic compliance or breaths to 1% nitrogen.

Urinalysis showed no differences between treated and control animals in either species.

There was no evidence of IgG deposition in the kidneys of rats or monkeys of either sex following exposure to the test materials for 90 days.

Group mean lead levels in the rat tissues were as follows:

	Control	Leaded 400 ppm	Unleaded 100 ppm
Brain	M	1.26	9.49
	F	1.44	5.39
Kidney	M	1.71	12.4
	F	2.97	9.57
Liver	M	0.71	17.9
	F	1.21	19.7
Blood	M	0.61	6.1
	F	0.24	1.32
Urine	M	0.17	0.21
	F	0.31	0.18

No actual values are given on organ weights or organ/body weight ratios but the following effects are reported:

### Rats

	Liver wt	Kidney wt
Unleaded 400 ppm M	increased	
Unleaded 400 ppm F		
Unleaded 1500 ppm M		
Unleaded 1500 ppm F		
Leaded 400 ppm M		
Leaded 400 ppm F	decreased	
Leaded 100 ppm M	increased	
Leaded 100 ppm F		increased

## 5. Toxicity

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### Monkeys

Unleaded 400 ppm M	Thyroid increased	Kidney
Unleaded 400 ppm F		
Unleaded 1500 ppm M	increased	
Unleaded 1500 ppm F		
Leaded 400 ppm M		decreased
Leaded 400 ppm F		
Leaded 100 ppm M		
Leaded 100 ppm F		

Organ weights were also expressed as % of body weight and the following effects were recorded:

#### Rats:

Decreased heart weight in both male leaded groups  
Decreased brain weight in both male unleaded groups  
Decreased liver weight in 400 ppm female leaded group  
Decreased adrenal weight in 1500 ppm female unleaded group.

#### Monkeys:

Decreased kidney weight in 400 ppm male unleaded group.

No evidence of treatment-related histopathology was observed in either rats or monkeys, with the exception of lesions noted in the kidneys of all male rats. The lesions were characterized by subtle but discernible increases in the incidence and severity of regenerative epithelium and dilated tubules. The latter were seen to contain protein in their lumens.

### Test substance

: An unleaded EPA reference fuel and a commercially available leaded gasoline were tested. The compositional properties of the two fuels were as follows:

	<u>Unleaded fuel</u>	<u>Leaded fuel</u>
<u>Calculated data</u>		
Research octane No.	93	87
Motor octane No.	88	86
Reid vapor pressure (PSIA)	6.9	6.3
Distillation °F (ASTM D-86)		
Initial boiling point	80	80
10%	135	160
50%	210	217
90%	275	295
100%	345	340
FIA analysis		
% aromatics	30.1	27.4
% olefins	8.2	7.8
% saturates	61.7	64.8

#### Experimental data

API gravity at 60°F	57.0	58.4
Sulfur, ppm	240	75
Lead, g/gallon	<0.005	1.94
Benzene, LV%	0.2	0.4
Toluene, LV%	16.7	11.4
n-Butane, LV%	1.0	0.4
Isopentane, LV%	5.4	5.5
n-Pentane, LV%	4.8	4.0

**Reliability**  
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: (2) valid with restrictions

(20)

## 5.5 GENETIC TOXICITY 'IN VITRO'

**Type** : Ames test  
**System of testing** : Microbial mutation assay in Salmonella typhimurium and Saccharomyces cerevisiae  
**Metabolic activation** : with and without  
**Result** : Negative  
**Year** : 1977  
**GLP** : no data  
**Test substance** : Unleaded gasoline  
**Method** :

The solubility, toxicity and dose levels for the test material were determined prior to the mutagenicity screening.

DMSO was used as solvent.

Based on the preliminary studies the following concentrations of test material were used in the mutagenicity assays:

Test doses	% Concentration	
	Bacteria	Yeast
1/8 50% survival	0.375	0.625
1/4 50% survival	0.75	1.25
1/2 50% survival	1.5	2.5
50% survival	3	5

## Plate tests

For non-activation assays cells in broth were exposed to the test material at the concentrations shown above. The contents of the tubes of broth plus test material were poured over selective agar plates which were then incubated. The test was conducted with and without metabolic activation.

Positive control substances (see results section) were also run in the same assay.

The following evaluation criteria were used in this plate test.

## Strains TA1535, 1537 and 1538

If the solvent control value is within the normal range a chemical which produces a positive response over three concentrations with the lowest increase equal to twice the solvent control value is considered to be mutagenic.

## Strains TA98, 100 and D4

If the solvent control value is within the normal range, a chemical which produces a positive response over three concentrations with the highest increase equal to twice the solvent control value for TA100 and two to three times the solvent control value for strains TA98 and D4 is considered to be mutagenic. For these strains, the dose response increase should start at approximately the solvent control value.

## Pattern

Because TA1535 and TA100 were both derived from the same parental strain (G-46) and because TA1538 and TA98 were both derived from the same parental strain (D3052), there is a built-in redundancy in the microbial assay. In general the

two strains of a set respond to the same mutagen and such a pattern is sought. It is also anticipated that if a given strain responds to a mutagen in non-activation tests it will generally do so in activation tests, but the converse of this is not anticipated.

While similar response patterns are not required for all mutagens, they can be used to enhance the reliability of an evaluation decision.

#### Reproducibility

If a chemical produces a response in a single test which cannot be repeated in one or more additional runs, the initial positive test data loses significance.

The above criteria are not absolute and other extenuating factors may enter into a final evaluation decision.

#### Suspension tests

Bacteria and yeast cultures were grown in complete broth. The cells were removed, washed and exposed to the test material at the concentrations shown in the results section. For the yeast cells exposure to the test material was for 4 hours whereas for the bacterial cells exposure was for 1 hour. Aliquots of the cells were plated onto the appropriate complete media. After suitable incubation periods, the number of revertant colonies were counted. This assay was also conducted with and without metabolic activation and positive control substances were also included.

The following criteria were used in the suspension assay.

#### Surviving population counts

A certain level of chemically-induced toxicity is anticipated, but occasionally isolated tests show very low (<25%) survival compared to the tissue controls. Data of this type are generally unacceptable and these experiments are repeated at a lower dose level.

#### Total mutant counts

For non mutagens, the ratio of mutant to surviving population should be roughly equivalent for each test point in a given experiment.

A mutagenic chemical will produce an altered mutant/surviving population ratio. An attempt is made to keep the surviving population of cells high and to look for positive responses that show increases in both numbers of mutants and mutation frequencies.

#### Dose-response

Dose-related increases in mutants and mutation frequencies are the most convincing data when assessing mutagenic activity. To ensure a proper dose response, dose levels are kept within a relatively low range o

**Result****:** Plate test

There was no increase in revertants caused by exposure to the test material at any concentration. The results in this assay were negative both with and without metabolic activation.

Suspension test

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The mutation frequencies are summarized in the following table for assays with and without metabolic activation.

### Non activation assay

Dose level	Salmonella strains					Yeast D4**
	TA100	TA1535	TA1537	TA1538	TA98*	
-ve control	5.48	3.59	6.15	7.1	41.99	23.69
+ve control	125.51	185.65	161.54	84.75	100	66.29
1 (low)	18.18	2.26	12.54	27.78	233.33	9.52
2	2.9	2.15	8.97	11.76	63.04	36.99
3	3.1	2.98	7.19	10	9.56	30.02
4 (high)	4.13	2.66	9.68	3.21	35.74	32.38

\* Assay repeated for negative control and lowest 2 doses.  
Results were 54.59 for -ve control  
10.84 for lowest dose  
14.11 for next highest dose

\*\* Assay repeated at all dose levels  
Results were: -ve control 4.66  
+ve control 97.73  
dose level 1 1.3  
dose level 2 8.33  
dose level 4 12.65

Slight increases are observed at the high dose levels with TA100, TA1537 and TA1538. However the responses are not adequate enough to be considered positive. The increases with TA98 could not be reproduced.

### With activation

Dose level	Salmonella strains					Yeast D4**
	TA100	TA1535	TA1537	TA1538	TA98*	
-ve controls*						
A+C	17.08	5.25	6.01	4.8	21.01	52.66
A-C	17.29	8.77	9.29	8.25	62.02	7.96
AL1	17.34	7.32	3.99	6.48	45.03	30.06
+ve control	25.51	89.92	0.22	1253.4	555.35	115.3
1 (low)	22.97	41.67	100	71.43	100	
2	15.64	7.21	0	300	30.66	27.22
3	17.26	9.57	20	15.38	83.33	27.03
4	22.31	7.21	5.43	6.93	60.13	29.04

\* Controls were  
A+C No activation system but including positive control  
A-C Solvent control, no test chemical or activation system  
AL1 Liver homogenate control plus solvent

Scattered increases were found at one or more dose levels (see table above). All apparent positive effects were repeated and were not reproducible indicating problems associated with the initial runs. When the raw data were inspected it was observed that the increases were due to anomalous reductions in viable cell counts. The results of this assay were therefore considered to be negative.

### Reliability

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: (2) valid with restrictions  
Valid with restrictions due to poor quality of initial assay.

(2)

## 5. Toxicity

**Id** Gasoline  
**Date** 11.12.2001

**Type** : Mouse lymphoma assay  
**System of testing** : Forward mutation assay using cell line L5178Y TK+/-  
**Metabolic activation** : with and without  
**Result** : negative  
**Year** : 1977  
**GLP** : no data  
**Test substance** : Unleaded gasoline  
**Method** : The test material was dissolved in acetone for this assay. The positive control substances were Ethyl methane sulphonate (EMS) and Dimethylnitrosamine (DMN).

A cytotoxicity study was carried out prior to the mutagenicity assay. For the mutation assay the lymphoma cells were exposed for 5 hours to test material at concentrations ranging from 0.065 to 1.04 µl/ml for both the activation and non-activation assays. After exposure to the test material, the cells were allowed to recover for 3 days and then cultures were selected for cloning and mutant selection. Surviving cell populations were determined by plating diluted aliquots in non-selective growth medium.

A mutation index was derived by dividing the number of clones formed in the BUdR-containing selection medium by the number found in the same medium without BUdR. The ratio was then compared to that obtained from other dose levels and from positive and negative controls.

A compound is considered mutagenic if:

A dose response relationship is observed over 3 of the 4 dose levels employed.

The minimum increase at the high level of the dose response curve is at least 2.5 times greater than the solvent control value.

The solvent control data are within the normal range of the spontaneous background for the TK locus.

**Result** : Little toxicity was observed with the test material. Positive control values exhibited significant responses over the negative controls, and the negative controls were within the normal range. All results for the test material from the non-activation assay were negative. The results from the activation assay were also considered to be negative. There was an increase in the number of mutants at the 0.52 µl/ml concentration but this appeared to result from a slight increase in the number of viable clones. There was no trend indicating a dose-related response and, therefore, the increases were not believed to be compound related.

The results are summarized below.

## 5. Toxicity

**Id** Gasoline  
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Dose (µl/ml)	Rel. susp. growth	Mutant clones	Viable clones	% Rel. growth	Mutant frequency	
Non-activation						
0.065	121.8	76	159	139.3	0.478	
0.13	103.7	29	215	160.4	0.1349	
0.26	114.6	44	211	174	0.2085	
0.52	141.8	66	161	164.3	0.4099	
1.04	107.5	58	270	208.9	0.2148	
Solvent		100	14	139	100	0.1007
Negative	129.9	41	140	130.8	0.2929	
EMS	58.7	227	67	28.3	3.3881	
Activation						
0.065	120.6	66	87	79.5	0.7586	
0.13	108.6	46	126	103.7	0.3651	
0.26	106	70	130	104.4	0.5385	
0.52	112.4	92	108	92	0.8519	
1.04	68.9	21	193	100.8	0.1088	
Solvent		100	30	132	100	0.2273
Negative	92.1	41	150	104.7	0.2733	
DMN	16.7	91	7	0.9	13	

**Reliability** : (1) valid without restriction  
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(2)

### 5.6 GENETIC TOXICITY 'IN VIVO'

**Type** : Cytogenetic assay  
**Species** : rat  
**Sex** : male  
**Route of admin.** : i.p.  
**Doses** : Acute study: 0.024, 0.08 & 0.24 ml/rat; subacute study: 0.01, 0.03 & 0.1 ml/rat  
**Result** : negative  
**Year** : 1977  
**GLP** : no data  
**Test substance** : API PS-6 unleaded gasoline  
**Method** : Two studies were conducted viz an acute and a subacute study. The test material was administered to the animals intraperitoneally in acetone.

Acute study  
Groups of 15 rats were given either acetone (0.1 ml/rat), or test material at doses of 0.024, 0.80 or 0.24 ml/rat. An additional group of 5 rats were given Triethylenemelamine(TEM) at a dose of 0.3 mg/kg. 6, 24 and 48 hours after administration of the test material 5 animals in each dose group were killed. For the TEM group, all five animals were killed 24 hours after administration of the substance.  
Two hours prior to being killed, cells were arrested in metaphase by the administration of a single i.p. dose of colchicine (4 mg/kg).  
Bone marrow was aspirated from the femurs and tibias of the lower limbs of the animals after they had been killed. The marrow plug was washed and then fixed. Slides of the cells were prepared and stained with Giemsa for microscopic examination. Fifty spreads were located for each animal and when of suitable quality, the chromosomes were counted and

## 5. Toxicity

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evaluated for the presence of abnormalities.

### Subchronic study

18 animals were used in this study. They were dosed with three levels of test compound (0.01, 0.03 & 0.1 ml/rat) once each day for 5 days. All animals were killed 6 hours after administration of the last dose and 2 hours prior to being killed they were treated with colchicine in the same way as the animals in the acute study.

Slides were prepared and examined as for the acute study. A negative and positive control group were also included, again the same as for the acute study.

### Remark

: A subsequent study (API ref 26-60099) was also carried out and this supported the negative conclusion of the original study. The second study is not summarized here.

### Result

: The results of the acute and repeat dose studies are summarized in the following table:

Material & dose (ml/rat)	Time after dose	No of rats	Total No of cells	% cells with aber*	Mitotic index	
<u>Acute study</u>						
Acetone (0.1)	6	3	100	1	3.4	
	24	3	100	0	4.3	
	48	3	150	0	3.8	
TEM (0.3 mg/kg)	24	5	200	30	3.8	
	PS-6 (0.024)	6	5	250	6	3.9
		24	5	250	1	4.9
48		5	250	3	4.7	
(0.08)	6	5	250	1	5.6	
	24	5	200	3	4.7	
	48	5	100	5	2.7	
(0.24)	6	5	187	2	3.2	
	24	3	100	0	4.1	
	48	5	200	3	4.5	
<u>Subacute study</u>						
Acetone (0.1)	5 days	3	150	1	5.8	
PS-6 (0.01)	5 days	5	200	1	3.5	
(0.03)	5 days	4	159	2	2.9	
(0.1)	5 days	5	174	2	3.2	

\* = aberrations

The results of the acute study were considered to be negative. There was an increase in aberrations at the 48 hour sacrifice period of the intermediate dose. The increases of 5% was significant, but did not fit into a trend suggestive of a compound-related mutagenic response. No other increases were observed at any dose level or sacrifice time.

The results of the subacute study were considered to be negative. There was no indication of an increased number of cells with aberrations.

Reliability  
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: (1) valid without restriction

(2) (3)

## 5. Toxicity

**Id** Gasoline  
**Date** 11.12.2001

**Type** : Dominant lethal assay  
**Species** : mouse  
**Sex** : male/female  
**Strain** : CD-1  
**Route of admin.** : inhalation  
**Exposure period** : 6 hours/day, 5 days/week for 8 weeks  
**Doses** : 400 & 1600 ppm  
**Result** : negative  
**Year** : 1980  
**GLP** : yes  
**Test substance** : API PS-6 unleaded gasoline  
**Method** : Groups of 10 male mice were exposed to either filtered air (negative controls) or test material at concentrations of 400 or 1600 ppm. Generation of test atmospheres was accomplished by bubbling air through the test material. Exposures were for 6 hours a day, 5 days each week for 8 weeks. On the final day of exposure a positive control group of 10 male mice were given Triethylenemelamine (TEM) intraperitoneally as a single i.p. dose, at a dose level of 0.3 mg/kg. The dose volume was 0.1 ml/mouse and the TEM was dissolved in 0.9% saline. Chamber concentrations were monitored at least hourly during the exposure periods. After 2 days rest following termination of exposures, each male was caged with 2 unexposed virgin female mice. At the end of 5 days, the females were removed. This weekly mating sequence was continued for 2 weeks. Each pair of mated females were transferred to a fresh cage and after 14 days after the midweek of being caged with the male were sacrificed. The uterine contents of the females were examined and scored for the numbers of dead and living implants and total implants.

### Evaluation Criteria

Dominant lethality was determined from a) a mutation index derived from the ratio of total to dead implants; or b) the number of dead implants per pregnant female. If true dominant lethality is observed then a significant increase in the number of dead implants per pregnant female should be accompanied by a significant decrease in the number of living implants per pregnant female. The two ratios are compared with both concurrent and comparable historical control values. Dose-related trends are also looked for. Any statistically significant differences must also be strongly evaluated for their biological significance.

In this study the following parameters were determined:

Fertility index ie. Proportion of pregnant females.  
Average No of implants/pregnant female.  
Average No. of dead implants/pregnant female.  
Proportion of females with one or more dead implants.  
Proportion of females with two or more dead implants.

**Result** : During the exposure phase actual chamber concentrations were found to be 0, 396.4 and 1524.6 ppm. One male died in the 1600 ppm group and another animal in the same group exhibited excessive lacrimation in the seventh

## 5. Toxicity

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week but this cleared in the final week.

The data for each of the parameters determined are as follows for untreated control, historical control, positive control and the two groups exposed to test material.

Week	Hist.	-ve	-ve	+ve	400 ppm	1600 ppm
------	-------	-----	-----	-----	---------	----------

### Fertility index

1	22/24	21/23	19/24	17/20	21/22
2	16/24	19/24	13/24	18/19	16/22

### Av. No. of implants/pregnant female

1	267/22	240/21	140/19	203/17	214/21
2	193/16	220/19	91/13	219/18	183/16

### Av. No. of dead implants/pregnant female

1	12/22	14/21	83/19	9/17	9/21
2	13/16	5/19	66/13	9/18	12/16

### Proportion of females with one or more dead implants

1	11/22	9/21	19/19	6/17	8/21
2	9/16	4/19	13/13	8/18	7/16

### Proportion of females with two or more dead implants

1	1/22	3/21	17/19	3/17	1/21
2	2/16	1/19	13/13	1/18	3/16

### No of dead implants/total implants

1	12/267	14/240	83/140	9/203	9/214
2	13/193	5/220	66/91	9/219	12/183

### Interpretation of the results:

The test material did not cause any significant reduction in the fertility index.

The test material had no effect on the average number of implants per pregnant female.

With respect to the number of dead implants per pregnant female, the test material showed no significant differences from the values of the concurrent as well as the negative controls.

The results support the conclusion that the test material did not cause increases in post-implantation deaths.

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: (1) valid without restriction

(6)

## 5.7 CARCINOGENITY

<b>Species</b>	:	rat and mouse
<b>Sex</b>	:	male/female
<b>Strain</b>	:	F 344 rat B6C3F mouse
<b>Route of admin.</b>	:	inhalation
<b>Exposure period</b>	:	Up to 113 weeks
<b>Frequency of treatment</b>	:	6 hours/day, 5 days/week
<b>Doses</b>	:	50, 275 & 1500 ppm nominal concentration
<b>Control group</b>	:	yes
<b>Method</b>	:	Similar to NCI guidelines
<b>Year</b>	:	1984
<b>GLP</b>	:	yes
<b>Test substance</b>	:	API PS-6 gasoline
<b>Method</b>	:	Groups of 100 rats of each sex and 100 mice of each sex were exposed to wholly vaporized gasoline at nominal concentrations of 50, 275 and 1500 ppm. 100 mice and 100 rats of each sex were exposed to air only and served as controls. Whole body exposures were in 16 m <sup>3</sup> glass and stainless steel chambers. Exposures were for 6 hours a day, 5 days each week for up to 113 weeks. All animals were individually housed and were allowed free access to food and water except during the exposure periods. Any animals that died during the first 10 days of exposure were replaced but thereafter no replacements were made. All animals were observed twice daily, once before and once after the exposure period. Animals found moribund were removed from the study and sacrificed. All animals were examined once per month for clinical signs and palpable tissue masses. Body weights were recorded monthly for the first 17 months and bi-weekly thereafter. After approximately 18 and 24 months exposure 7 male and 7 female rats from each dose group were selected and hematological and clinical evaluations were conducted on these. After 3, 6, 12 and 18 months exposure 10 rats and 10 mice of each sex from each dose group were sacrificed and underwent complete post mortem examinations. At study termination all surviving animals were sacrificed. Body weight were recorded and after gross examination a wide range of organs/tissues were removed, weighed and fixed for subsequent histopathological examination.
<b>Result</b>	:	Monitoring of the exposure chamber concentrations established that actual concentrations for the study were: 0, 67, 292 and 2056 ppm.

Results of study in rats

There were very few pharmacotoxic signs that occurred in only a few animals and insufficiently frequently to be considered treatment-related. Mortality rates were also unaffected by exposure to gasoline vapor.

Male rats in the highest dose group had lower body weights than controls from week 5 throughout the study. The difference amounted to 33 g at week 44 and this remained throughout. Females at the highest dose also weighed less than controls. A difference of 30 g had occurred by week 66 and this remained throughout the study.

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The few differences in hematological data between controls and several treatment groups were within the normal range for rats of similar age and not considered to be treatment-related.

Similarly, small changes in a few clinical chemical parameters were not considered to be treatment-related.

At gross necropsy at the 3, 6 12 and 18 month sacrifice the only significant macroscopic findings were in the kidneys of the high dose group male rats and these consisted of tan color, foci, mottling, discolored and granular surface. Although the incidence was small it was considered significant in the light of the histopathology findings. Additionally, masses or nodules were observed in mid (3 masses/nodules) and high (5 masses/nodules) dose male kidneys that died between 18 months and study termination. There were no other gross findings.

Microscopic pathology examination revealed an increase in the incidence of renal disease with tubular degeneration and regeneration or cystic dilatation in the mid and high dose males from 3 months onwards. At 24 months primary renal neoplasms were observed in the following incidence:

Dose group	Neoplasm	Males	Females
0 ppm		0	0
50 ppm	renal carcinoma	1	0
275 ppm	renal adenoma	2	0
	renal carcinoma	2	0
	renal sarcoma	1	1
2056 ppm	renal carcinoma	6	0
	renal adenoma	1*	

\* Occurred at 18 months

With the exception of one renal sarcoma all other tumors occurred in males.

### Results of study in mice

There were no consistent signs of toxicity attributable to treatment and mortality rates were considered to be unaffected by treatment. Growth rates were similar for treated and control groups up until approximately week 70 after which the highest dose group males and females had lower body weights than controls. The difference amounted to approximately 2.3 to 4.4g on a body weight of 35g for males and 2 to 3g on a weight of 33g for females. The investigators considered this reduced body weight to be attributable to treatment. Organ weights were unaffected by treatment.

There was an increased incidence of liver nodules and masses in treated females in the high dose group that died on the study from 18 months to termination and which were terminally sacrificed. The incidence is tabulated as follows:

	Dose group (ppm)			
	0	67	292	2056
<u>MALES</u>				
Dead on study				
plus 18mth group	3/5	2/7	3/14	4/8
Terminal sacrifice	14/46	12/35	13/30	21/46
Total	17/51	14/42	16/44	25/54

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### FEMALES

Dead on study				
plus 18mth group	4/16	4/19	7/20	8/14
Terminal sacrifice	5/41	6/33	8/37	18/42
Total	9/57	10/52	15/57	26/56

There was a possible reduction in the incidence of cystic or enlarged uteri for female mice.

The incidences were

0 ppm	38/41
67ppm	26/33
292ppm	19/37
2056ppm	12/42

There were no other treatment-related findings at necropsy.

Microscopic examination of the tissues of animals up to and including the 18 month sacrifice did not reveal any compound-related effects.

At 24 months, however, there was an increased incidence of hepatocellular tumors in the high dose group females when compared to controls. The actual incidence of liver tumors is shown in the following table.

	Dose group (ppm)			
	0	67	292	2056
Number examined	57	52	57	56
Hepatocellular adenoma	1	4	4	8
Hepatocellular csarcinoma	7	6	9	20
Animals with hepatocellular tumors*	8	10	12	27

\* Some animals had more than 1 tumor.

No other compound-related lesions were observed.

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(18) (19) (21)

### 5.8 TOXICITY TO REPRODUCTION

Type	: Two generation study
Species	: rat
Sex	: male/female
Strain	: Sprague-Dawley
Route of admin.	: inhalation
Exposure period	: 6 hours
Frequency of treatment	: Daily
Premating exposure period	
Male	: 10 weeks
Female	: 10 weeks
Duration of test	: Two generations
Doses	: 5000, 10 000 & 20 000 mg/m <sup>3</sup>
Control group	: yes
NOAEL Parental	: = 20000 mg/m <sup>3</sup>
NOAEL F1 Offspr.	: = 20000 mg/m <sup>3</sup>
Method	: OECD Guide-line 416 "Two-generation Reproduction Toxicity Study"
Year	: 2000
GLP	: yes
Test substance	: Volatile fraction of gasoline

## 5. Toxicity

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### Method

- : Groups of 30 male and 30 female Sprague Dawley rats were exposed 6 hours/day, seven days/week to volatilized test material at target concentrations of 5000, 10 000 and 20 000 mg/m<sup>3</sup>. Singly housed animals were exposed for 10 weeks prior to mating. There was then a 3 week mating period and mating was confirmed by either presence of sperm in a vaginal rinse or by the presence of a vaginal plug. Exposure of females was continued until gestation day 20. Exposure was then suspended until post partum day 5 to avoid unduly stressing the dams during birth and was then re-commenced and continued until sacrifice of parental females after weaning. The pups were culled on a random basis to approximately 5/sex/litter. At weaning on postnatal day 28, the F1 pups were selected for the second generation. Among the pups not selected, 3/sex/litter were sacrificed and examined for internal abnormalities. The remainder were examined for external abnormalities, sacrificed and discarded.
- The pups selected for F1 were exposed for a 13 week pre-mating period and then for a 3 week mating period as described above. The males were sacrificed at this time and the females continued to be exposed until gestation day 20. As described above exposures were resumed on post partum day 5 and was continued until weaning, when all remaining animals were sacrificed. Other than during the period from gestation day 20 until post partum day 5, all F1 offspring were exposed from conception to sacrifice.
- All animals were examined regularly for viability and clinical observations. Body weights and food intakes were also recorded regularly throughout the study. All pups were counted and examined externally on a daily basis and weighed at regular intervals until post natal day 21. F1 pups were examined regularly between post natal days 21 to 28 and were weighed on days 28 and 35. All surviving F1 and F2 pups were examined for developmental landmarks, including pinna detachment, hair growth, incisor eruption, eye opening and the development of the surface righting reflex. Surviving F1 female offspring were monitored for vaginal opening and males were examined for preputial separation. Reproductive parameters evaluated included: male and female fertility indices, male mating index, female fecundity and gestational indices, mean litter size, mean days of gestation, female estrous cycle length and number of females cycling normally. Live birth index, survival index, survival indices (post partum days 1, 4, 7, 14 and 21), viability index at weaning, mean live and dead offspring on day 0, sex ratio at day 0, offspring in-life observations, offspring body weight and offspring gross postmortem findings were also assessed.
- All animals dying or sacrificed in a moribund condition were necropsied. Culled pups were examined externally but were only necropsied if external evidence warranted it. Randomly selected pups were necropsied and the weight of the following organs was determined: ovaries, liver, adrenals, testes, kidneys, spleen and brain. Additionally a wide range

**Result**

of tissues were taken for histology. Similar evaluations were also carried out on all adults surviving to scheduled sacrifice. Tissues taken from the high dose group and controls were evaluated histologically and since there were no untoward findings, tissues from the lower dose groups were not examined. Samples of sperm from the left distal cauda epididymis were collected from all males at terminal sacrifice for evaluation of sperm parameters. These included assessments of total caudal epididymal sperm numbers, % progressively motile sperm and homogenization resistant spermatid count, % morphologically normal sperm and % sperm with an identified abnormality. An ovarian examination was carried out in the females that included confirmation of growing follicles and corpora lutea and quantification of primordial oocytes. This was done in the high dose and control groups and since there were no abnormal findings other groups were not evaluated.

: There were no treatment-related clinical signs, or effects on mortality, body weight or food intake in either parents or pups. Furthermore, there were no treatment-related post mortem findings.

There were no significant differences in absolute organ weights in either males or females from the first parental generation. In the second parental generation, however, there were some statistically significant increases in absolute organ weights, including liver, kidneys and testis in the males and lungs in the females, but none of the differences between controls and the high dose group was statistically significant. In the absence of a clear dose-response relationship the significance of the result was unclear. When expressed as organ/body weight ratios, the only significant difference was seen in male kidney weights in the lowest dose group of the first parental generation and an increase in the highest dose group of the second parental generation. Although this latter may have been treatment related it was not considered to be of clinical importance.

There were no compound-related microscopic changes in any of the reproductive tissues or in the upper or lower respiratory tract from any of the P1 or P2 rats exposed to 20 000 mg/m<sup>3</sup>. The only microscopic changes seen were in the kidneys of males of both generations. There was an exposure related increase in the amount and size of hyaline droplets. In three male rats of the high exposure group from both P1 and P2 animals granular casts were observed in the medullary tubules of the kidneys. These kidney changes and the accompanying weight increases are regarded as a sex and species specific effect and of no relevance for man.

In the first generation, there were no differences in mating index, fecundity, pregnancy or length of gestation. Among the offspring there were no differences in litter size, fraction of live births or sex ratio. Results in the second generation were similar. There were no differences in survival of offspring through weaning in the first generation and in the second generation early survival was slightly higher among the offspring from the exposed dams.

## 5. Toxicity

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### Test substance

There were no differences in the weight of the offspring through weaning in either generation.  
There were no unusual post mortem observations.

The sperm analysis carried out on both P1 and P2 (F1) males revealed no effects on sperm count, progressive motility or gross appearance.

No effects were found on the estrous cycle length, quantification of primordial oocytes or % females with abnormal cycles in the P1 or P2 generations.

There were no significant differences in incisor eruption, pinna detachment, or surface righting reflex in the F1 or F2 offspring. Hair growth was delayed by just less than one day in males only of the F1 pups and in both sexes of the lowest dose group (approx half day) for the F2 pups. Eye opening was advanced by approximately one-half day for the high dose males of the F2 offspring.

: The test material was a condensate of gasoline vapor that had been collected from a vapor recovery unit during normal operations. This test material was selected since it was representative of the exposures that normally occur for the general public during self-service refueling. Analytical studies were conducted on the condensate and the results compared with exposure studies that had been carried out during refueling operations. The results confirmed that the vapor recovery condensate was similar in composition to the vapors to which the public are exposed during refueling.

Test atmospheres for the inhalation study were generated by fully volatilizing the condensate and diluting with air to achieve target concentrations of 5000, 10 000 and 20 000 mg/m<sup>3</sup>. The highest concentration was approximately 50% of the lower explosive limit and several orders of magnitude greater than the concentrations to which the public are exposed..

Chamber analyses of the test atmospheres confirmed the actual concentrations to be: 5076, 10 274 and 20 241 mg/m<sup>3</sup>.

Analysis of the vapor recovery condensate gave the following results:

<u>Component</u>	<u>Vol %</u>
<u>Non aromatics</u>	
C3	1.0
C4	51.7
C5	37.2
C6	8.3
C7	0.4
C8	0.2
C9	-
C9+	-
Total saturates (vol%)	-
Total olefins (vol%)	-
<u>Aromatics</u>	
Benzene	0.7
Toluene	0.7
C8	-

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C9 -  
C9+ -  
Total aromatics (vol%) -

**Reliability** : NB - denotes no data available.  
21.11.2001 : (1) valid without restriction (25)

### 5.9 DEVELOPMENTAL TOXICITY/TERATOGENICITY

**Species** : rat  
**Sex** : female  
**Strain** : Sprague-Dawley  
**Route of admin.** : inhalation  
**Exposure period** : 6 hours each day  
**Frequency of treatment** : Daily  
**Duration of test** : Days 6 through 15 of gestation  
**Doses** : 400 and 1600 ppm  
**Control group** : yes  
**NOAEL Maternalt.** : = 1600 ppm  
**NOAEL Teratogen** : = 1600 ppm  
**Year** : 1978  
**GLP** : no data  
**Test substance** : Unleaded gasoline as described in section 1.1 above  
**Method** : Female rats were mated with sexually mature males of the same strain. The females were examined daily for evidence of a copulatory plug and when this was observed it was designated day 0 of gestation. The mated female rats were assigned sequentially into three groups of 25 animals for the 0, 400 and 1600 ppm dose groups and were caged individually. The animals were subjected to whole body exposure to gasoline vapors at the concentrations shown above for 6 hours each day from day 6 through day 15 of gestation. Mated females were weighed on days 0, 6, 15 and 20 of gestation. Food consumption was recorded daily during the periods 0-6, 6-15 and 15-20 days of gestation. Observations were made daily for clinical signs. On day 20 of gestation the female rats were anesthetized and their visceral and thoracic organs were examined. The uterus was removed and opened and the number of implantation sites, their placement in the uterine horns, live and dead fetuses and resorption sites recorded. The fetuses were removed, examined externally for abnormalities and weighed. One third of the fetuses from each litter were fixed in Bouin's and examined later for changes in the soft tissues of the head, thoracic and visceral organs. The remaining fetuses in each litter were stained with Alizarin Red S and examined for skeletal abnormalities. The uterus and ovaries from the adult females were preserved for possible future examination.

**Result** : Chamber concentrations were found to be:

Nominal (ppm)	Actual (ppm)
0	0
400	442±42
1600	1573±80

## 5. Toxicity

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There were no deaths during the study and all animals appeared normal throughout. There were no treatment-related effects on body weight or food consumption. There were no treatment related effects on any of the reproductive parameters recorded. These data are summarized as follows:

	Historical control	0 ppm	400 ppm	1600 ppm
Pregnancy ratio (pregnant/bred)	-	20/22	22/22	20/21
Live litters	99%	20	22	20
Implantation sites (left /right horn)	46/54%	123/145	149/158	143/152
Resorptions	252	16	22	15
Litters with resorptions	50%	65%	41%	55%
Dead fetuses	1	0	0	0
Litters with dead fetuses	1	0	0	0
Live fetuses/Implantaion site	92%	95%	93%	95%
Mean live litter size	12.2	13	13	14
Average fetal wt. (g)	3.5	3.8	3.7	3.6

No treatment related effects were observed during the examination for soft tissue changes in the fetuses. Results of the skeletal examination of the stained fetuses are summarized below:

Dose (ppm)	Fetuses examined	Fetuses normal	Fetuses with commonly encountered changes only	Fetuses with Unusual skeletal variations
0	177 (20)*	112	60 (18)	5 (5)
400	197** (22)	128	55 (16)	14 (4)
1600	196 (20)	131	47*** (14)	18*** (7)

\* Average No. of litters in parenthesis

\*\* Two specimens of one litter lost on processing

\*\*\* p<0.05

The unusual changes were mainly related to retarded ossification and were not considered as malformations.

Statistical analysis of data on a pup basis revealed a significant difference between the 1600 and 0 ppm groups. However when analyzed on a litter basis no statistically significant differences were found.

### Conclusion

: Exposure of pregnant rats to vapors of unleaded gasoline at concentrations of 400 or 1600 ppm did not cause effects on pregnant dams. There was no evidence of variation in sex ratio, embryo toxicity, inhibition of fetal growth or development or teratogenic potential.

**Reliability**  
19.11.2001

: (1) valid without restriction

(4)

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**Id** Gasoline  
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- (30) Springborn Laboratories, Inc.(1993). CWE5 (Blended Gasoline) Toxicity to Freshwater Alga, *Selenastrum capricornutum*SLI Report # 93-5-4783.Springborn Laboratories, Inc Environmental Sciences division, 790 Main Street, Wareham, Massachusetts, USA
- (31) Study conducted by Exxon Biomedical Sciences Inc. Fish - acute toxicity test: study no. 104958, test substance MRD-95-049. CONCAWE, Brussels, 1995

**ROBUST SUMMARY  
OF INFORMATION ON**

**Substance Group:** **NAPHTHENIC NAPHTHAS**

**Summary prepared by:** American Petroleum Institute

**Creation date:** 22 FEBRUARY 2001

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**Date of last Update:** 10 DECEMBER 2001

**Number of Pages:** 28

NB. Reliability of data included in this summary has been assessed using the approach described by Klimisch, et al.

Klimisch, H. J., Andreae, M. and Tillman, U, (1997)

A systematic approach for evaluating the quality of experimental toxicological and ecotoxicological data.  
Regulatory Toxicology and Pharmacology 25, 1-5.

## 1.1 GENERAL SUBSTANCE INFORMATION

**Substance type** : petroleum product  
**Physical status** : liquid  
**Remark** : The naphtha streams that are rich in naphthenes are obtained from the atmospheric distillation of crude oil. The streams contain saturated and aromatic hydrocarbons, mainly in the range C4 to C12 and boil in the range of approximately minus 20 to 230 °C

Sweetened Naphtha (CAS No. 64741-87-3) is typical of the streams in this category.

An API sample of Sweetened naphtha that has been used for some of the toxicology studies for this group has been characterized as follows:

### SAMPLE API 81-08

Gravity (°API)	76.6
Sulfur (wt %)	0.1
Nitrogen (ppm)	<1
RVP (psia)	01.2
IBP (°F)	98
FBP (°F)	262
Paraffins (% by MS)	76.5
Olefins (% by MS)	1.0
Naphthenes (% by MS)	16.5
Aromatics (% by MS)	4.0
Saturates (% by MS)	-

12.11.2001

## 2. Physico-Chemical Data

**Id** N. Naphthas  
**Date** 10.12.2001

### 2.1 MELTING POINT

: Not relevant

### 2.2 BOILING POINT

**Value** : Approximately 49 - 177 ° C at 1013 hPa  
**Decomposition** : no  
**Remark** : The samples, which were used by the API in its toxicity assessments for this Group were prepared by the fractionation of two types of crude oil, using a pilot plant still and separating cuts in a distillation range of 120 to 350°F (49 to 177°C).

These figures represent a typical boiling range for light straight-run naphtha, CAS No. 64741-46-4.

The standard oil industry method for determination of boiling range is ASTM D86.

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(10) (18)

### 2.5 PARTITION COEFFICIENT

**Log pow** : 2.13 - 4 at 25° C  
**Method** : Calculated by LOGKOWWIN ver. 1.65  
**Year** : 2000  
**GLP** : no  
**Test substance** : Light Straight Run Naphtha (LSRN)-Low Naphthenic, CONCAWE sample CWE3, CAS No. 64741-46-4  
**Remark** : Log P values represent the spread of calculated and/or measured values for C5 to C7 hydrocarbon components found in LSRN, CAS No 64741-46-4. Detailed hydrocarbon analysis was used to identify the components of this specific low naphthenic LSRN sample. Calculation based on an atom/fragment contribution method of W. Meylan and P. Howard. Calculated SAR result for surrogate structure contained in program database (smilecas.dat).

**Reliability** : (2) valid with restrictions

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(13) (20)

**Log pow** : 2.13 - 4.76 at 25° C  
**Method** : Calculated by LOGKOWWIN ver. 1.65.  
**Year** : 2000  
**GLP** : no  
**Test substance** : Light Straight Run Naphtha (LSRN)-Moderate (19.7%) Naphthenic, CAS No. 64741-46-4  
**Remark** : Log P values represent the spread of calculated and/or measured values for C5 to C9 hydrocarbon components found in LSRN, CAS No 64741-46-4. Detailed hydrocarbon analysis performed by Chevron Research was used to identify the components of this specific moderate naphthenic (19.7%) LSRN sample. Calculated SAR result for surrogate structures contained in program database (smilecas.dat). Calculation based on an atom/fragment contribution method of W. Meylan and P. Howard.

**Reliability** : (2) valid with restrictions

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(20)

## 2. Physico-Chemical Data

**Id** N. Naphthas  
**Date** 10.12.2001

**Log pow** : 2.73 - 4.85 at 25° C  
**Method** : Calculated by LOGKOWWIN ver. 1.65  
**Year** : 2000  
**GLP** : No  
**Test substance** : Light Straight Run Naphtha (LSRN)-Hi Naphthenic, CONCAWE sample W94/809, CAS No. 64741-46-4  
**Remark** : Log P values represent the spread of calculated and/or measured values for C5 to C9 hydrocarbon components found in LSRN, CAS No 64741-46-4. Detailed hydrocarbon analysis was used to identify the components of this specific high naphthenic LSRN sample. Calculation based on an atom/fragment contribution method of W. Meylan and P. Howard. Calculated SAR result for surrogate structure contained in program database (smilecas.dat).  
**Reliability** : (2) valid with restrictions  
27.10.2001 (13) (20)

### 2.6.1 WATER SOLUBILITY

**Method** : Preparation of Water Soluble Fractions  
**Year** : 1995  
**GLP** : yes  
**Test substance** : Light Straight Run Naphtha (LSRN)-Hi Naphthenic, CONCAWE sample W94/809, CAS No. 64741-46-4  
**Method** : Water Accomodated Fractions (WAFs) of LSRN were prepared at 100 mg/L loading in freshwater and equilibrated for 48 hours in tightly closed systems with minimal headspace.  
**Result** : Gas chromatographic analysis of TEX (toluene, ethyl benzene, and xylenes) components indicated freshwater solubility of 5.7-7.9 ppm (as TEX). Measured test concentrations of the LSRN were based on the total combined concentrations of TEXN which represent approximately 13% composition of the test substance. Concentrations for these components reached equilibrium by 19 hours.  
**Conclusion** : Individual components of complex petroleum substances have specific and differing solubilities. Calculated and measured water solubilities for LSRN components range from approximately 3 to 2000 mg/l. At any particular loading rate, the resulting aqueous concentration of each chemical constituent is a function of the relative volume of the two phases (aqueous and the petroleum mixture), the partition coefficient between the phases, the amount of component present and the maximum water solubility of each component. Initially as the petroleum mixture is added in amounts below the solubility limit of the least soluble component the aqueous concentration increases proportionally until the least soluble component reaches a saturation concentration, and only the more soluble components continue to dissolve, resulting in a two phase system. Further addition of the petroleum mixture results in an aqueous concentration that is a non-linear function of the amount added.  
**Reliability** : (2) valid with restrictions  
27.10.2001 (11) (12) (14) (15) (16)

**Method** : Preparation of Water Soluble Fraction  
**Year** : 1995  
**GLP** : yes  
**Test substance** : Light Straight Run Naphtha (LSRN)-Low Naphthenic, CONCAWE sample CWE39, CAS No. 64741-46-4  
**Method** : Water Accomodated Fractions (WAFs) of LSRN were prepared at 1000 mg/l loading in freshwater and equilibrated for 24 hours in tightly closed systems with minimal headspace.  
**Result** : Gas chromatographic analysis of BTEX components indicated freshwater solubility at 24 hours of 4.9 ppm as benzene.

## 2. Physico-Chemical Data

Id N. Naphthas  
Date 10.12.2001

**Conclusion** : Individual components of complex petroleum substances have specific and differing solubilities. Calculated and measured water solubilities for gasoline components range from approximately 3 to 2000 mg/l. At any particular loading rate, the resulting aqueous concentration of each chemical constituent is a function of the relative volume of the two phases (aqueous and the petroleum mixture), the partition coefficient between the phases, the amount of component present and the maximum water solubility of each component. Initially as the petroleum mixture is added in amounts below the solubility limit of the least soluble component the aqueous concentration increases proportionally until the least soluble component reaches a saturation concentration, and only the more soluble components continue to dissolve, resulting in a two phase system. Further addition of the petroleum mixture results in an aqueous concentration that is a non-linear function of the amount added.

**Reliability** : (2) valid with restrictions  
27.10.2001 (11) (14) (15) (16) (22)

**Method** : Preparation of water soluble fraction

**Year** : 1995

**GLP** : Yes

**Test substance** : Light Straight Run Naphtha (LSRN)-Moderate (19.7%) Naphthenic, CAS No. 64741-46-4

**Method** : Water Accomodated Fractions (WAFs) of LSRN were prepared at 50 mg/L loading in freshwater and saltwater and equilibrated for 48 hours in tightly closed systems with minimal headspace

**Result** : Gas chromatographic analysis of LSRN components 2-methyl-pentane, cyclohexane, benzene, toluene, ethylbenzene, ortho, meta and para-xylene in WAFs indicated freshwater solubility of 1.62-1.81 ppm and 1.50 mg/l in saltwater. Concentrations for these components reached equilibrium by 24 hours.

**Conclusion** : Individual components of complex petroleum substances have specific and differing solubilities. Calculated and measured water solubilities for LSRN components range from approximately 3 to 2000 mg/l. At any particular loading rate, the resulting aqueous concentration of each chemical constituent is a function of the relative volume of the two phases (aqueous and the petroleum mixture), the partition coefficient between the phases, the amount of component present and the maximum water solubility of each component. Initially as the petroleum mixture is added in amounts below the solubility limit of the least soluble component the aqueous concentration increases proportionally until the least soluble component reaches a saturation concentration, and only the more soluble components continue to dissolve, resulting in a two phase system. Further addition of the petroleum mixture results in an aqueous concentration that is a non-linear function of the amount added.

**Reliability** : (2) valid with restrictions  
11.11.2001 (4) (11) (14) (15) (16)

### 3. Environmental Fate and Pathways

Id N. Naphthas  
Date 10.12.2001

#### 3.1.1 PHOTODEGRADATION

<b>Type</b>	:	Calculated	
<b>Light source</b>	:	Sun light	
<b>Indirect photolysis</b>			
<b>Sensitizer</b>	:	OH	
<b>Method</b>	:	Calculated by AOPWIN ver. 1.89. Method based on the work of R. Atkinson	
<b>Year</b>	:	2000	
<b>GLP</b>	:	No	
<b>Test substance</b>	:	Light Straight Run Naphtha (LSRN)-Low Naphthenic, CONCAWE sample CWE39, CAS No. 64741-46-4	
<b>Remark</b>	:	AOPWIN ver. 1.89 calculates atmospheric oxidation half lives of hydrocarbons in contact with hydroxyl radicals in the troposphere, under the influence of sunlight and in contact with O <sub>3</sub> . Atmospheric oxidation rates were calculated for the C <sub>5</sub> to C <sub>9</sub> hydrocarbon components found in LSRN, CAS No. 64741-46-4. Detailed hydrocarbon analysis was used to identify the components of this specific low naphthenic LSRN sample. Based on a 12-hour day, the range for atmospheric half-lives for LSRN constituents is: 1.262 days (cyclohexane) to 15.985 days (isopentane).	
<b>Result</b>	:	Indirect Photolysis	
		Sensitizer:	OH radical
		Conc. of sensitizer:	1.50E+06 OH radicals/cm <sup>3</sup>
		Rate Constant:	0.6991 E-12 (isopentane) to 8.4783 E-12 (cyclohexane) cm <sup>3</sup> /molecule-sec
		Half-life	1.262 to 15.985 days
<b>Reliability</b>	:	(2) valid with restrictions	(13) (21)
27.10.2001			
<b>Type</b>	:	Calculation	
<b>Light source</b>	:	Sun light	
<b>Indirect photolysis</b>			
<b>Sensitizer</b>	:	OH	
<b>Deg. Product</b>	:		
<b>Method</b>	:	Calculated by AOPWIN ver. 1.89. Method based on the work of R. Atkinson	
<b>Year</b>	:	2000	
<b>GLP</b>	:	No	
<b>Test substance</b>	:	Light Straight Run Naphtha (LSRN)-Hi Naphthenic, CONCAWE sample W94/809, CAS No. 64741-46-4	
<b>Remark</b>	:	AOPWIN ver. 1.89 calculates atmospheric oxidation half lives of hydrocarbons in contact with hydroxyl radicals in the troposphere, under the influence of sunlight and in contact with O <sub>3</sub> . Atmospheric oxidation rates were calculated for the C <sub>5</sub> to C <sub>9</sub> hydrocarbon components found in LSRN, CAS No. 64741-46-4. Detailed hydrocarbon analysis was used to identify the components of this specific high naphthenic LSRN sample. Based on a 12-hour day, the range for atmospheric half-lives for LSRN constituents is: 0.789 days (m-xylene) to 5.486 days (benzene).	
<b>Result</b>	:	Indirect Photolysis	
		Sensitizer:	OH radical
		Conc. of sensitizer:	1.50E+06 OH radicals/cm <sup>3</sup>
		Rate Constant:	1.9498 E-12 (benzene) to 13.5606 E-12 (m-xylene) cm <sup>3</sup> /molecule-sec
		Half-life:	0.789 to 5.486 days
<b>Reliability</b>	:	(2) valid with restrictions	(13) (21)
27.10.2001			

### 3. Environmental Fate and Pathways

Id N. Naphthas  
Date 10.12.2001

**Type** : Calculation  
**Light source** : Sun light  
**Rel. intensity** : 1 based on Intensity of Sunlight  
**Indirect photolysis**  
**Sensitizer** : OH  
**Method** : Calculated Calculated by AOPWIN ver. 1.89. Method based on the work of R. Atkinson  
**Year** : 2000  
**GLP** : No  
**Test substance** : Light Straight Run Naphtha (LSRN)-Moderate (19.7%) Naphthenic, CAS No. 64741-46-4  
**Remark** : AOPWIN ver. 1.89 calculates atmospheric oxidation half lives of hydrocarbons in contact with hydroxyl radicals in the troposphere, under the influence of sunlight and in contact with O<sub>3</sub>. Atmospheric oxidation rates were calculated for the C5 to C9 hydrocarbon components found in LSRN, CAS No. 64741-46-4. Detailed hydrocarbon analysis was used to identify the components of this specific moderate naphthenic LSRN sample. Based on a 12-hour day, the range for atmospheric half-lives for LSRN constituents is: 0.789 days (m-xylene) to 15.985 days (isopentane).  
**Result** : Indirect Photolysis  
Sensitizer: OH radical  
Conc. of sensitizer: 1.50E+06 OH radicals/cm<sup>3</sup>  
Rate Constant: 0.6691E-12 (isopentane) to 13.5606E-12 (m-xylene) cm<sup>3</sup>/molecule-sec  
Half-life: 0.789 to 15.985 days  
**Reliability** : (2) valid with restrictions  
11.11.2001 (21)

#### 3.1.2 STABILITY IN WATER

**Test substance** : Light Straight Run Naphtha (LSRN)-Hi Naphthenic, CONCAWE sample W94/809, CAS No. 64741-46-4  
**Conclusion** : Hydrolysis unlikely  
**Reliability** : (1) valid without restriction  
27.10.2001 (17)

**Test substance** : Light Straight Run Naphtha (LSRN)-Low Naphthenic, CONCAWE sample CWE39, CAS No. 64741-46-4  
**Conclusion** : Hydrolysis unlikely  
**Reliability** : (1) valid without restriction  
27.10.2001 (17)

**Test substance** : Light Straight Run Naphtha (LSRN)-Moderate (19.7%) Naphthenic, CAS No. 64741-46-4  
**Conclusion** : Hydrolysis unlikely  
**Reliability** : (1) valid without restriction  
11.11.2001 (17)

### 3. Environmental Fate and Pathways

Id N. Naphthas  
Date 10.12.2001

#### 3.3.1 TRANSPORT BETWEEN ENVIRONMENTAL COMPARTMENTS

**Type** : Calculated according to Mackay Level 1  
**Media** : Soil, air, water, suspended sediment, sediment  
**Year** : 2000  
**Remark** : Model based on chemical fugacity. Physical properties input are those calculated by the EPIWIN Estimation 3.04 program and included in this summary. Values represent the spread of calculated values for C5 to C9 hydrocarbon components found in LSRN, CAS No 64741-46-4. Detailed hydrocarbon analysis was used to identify the components of this specific high naphthenic LSRN sample.

The majority of LSRN components will partition rapidly to air, where these hydrocarbons will be rapidly oxidized by OH radicals.

**Result** : 

<u>Medium</u>	<u>% distribution</u>
Air:	97 to 99.97
Soil:	0.03 to 1.2
Water:	0.008 to 2.7
Sediment	0.00 to 0.02
Suspended sediment	0.00

**Conclusion** : The constituents of this complex petroleum mixture are expected to partition primarily to air.

**Reliability** : (2) valid with restrictions  
24.09.2001

(13) (19)

**Type** : Calculated according to Mackay Level 1  
**Media** : Soil, air, water, suspended sediment, sediment  
**Method** : Calculated  
**Year** : 2000  
**Remark** : Model based on chemical fugacity. Physical properties input are those calculated by the EPIWIN Estimation 3.04 program and included in this summary. Values represent the spread of calculated values for C5 to C9 hydrocarbon components found in LSRN, CAS No 64741-46-4. Detailed hydrocarbon analysis was used to identify the components of this specific low naphthenic LSRN sample.

The majority of LSRN components will partition rapidly to air, where these hydrocarbons will be rapidly oxidized by OH radicals.

**Result** : 

<u>Medium</u>	<u>% distribution</u>
Air:	98.89 to 99.98
Soil:	0.01 to 0.11
Water:	0.01 to 1.00
Sediment	<0.001
Suspended sediment	<0.001

**Reliability** : (2) valid with restrictions  
27.10.2001

(13) (19)

### 3. Environmental Fate and Pathways

Id N. Naphthas  
Date 10.12.2001

**Type** : Calculated according to Mackay Level 1  
**Media** : Soil, air, water, suspended sediment, sediment  
**Method** : Calculated  
**Year** : 2000  
**Remark** : Model based on chemical fugacity. Physical properties input are those calculated by the EPIWIN Estimation 3.04 program and included in this summary. Values represent the spread of calculated values for C5 to C9 hydrocarbon components found in LSRN, CAS No 64741-46-4. Detailed hydrocarbon analysis was used to identify the components of this specific moderate naphthenic (19.7%) LSRN sample.  
The majority of LSRN components will partition rapidly to air, where these hydrocarbons will be rapidly oxidized by OH radicals.

**Result** : 

<u>Medium</u>	<u>% distribution</u>
Air:	97 to 99.99
Soil:	0.00 to 1.2
Water:	0.013 to 2.7
Sediment	0.00 to 0.03
Suspended Sediment	0.00

**Test substance** : Light Straight Run Naphtha (LSRN)-Moderate (19.7%) Naphthenic, No. 64741-46-4

**Conclusion** : The constituents of this complex petroleum mixture are expected to partition primarily to air.

**Reliability** : (2) valid with restrictions  
11.11.2001

(19)

## 4.1 ACUTE/PROLONGED TOXICITY TO FISH

<b>Type</b>	: Static with daily renewal
<b>Species</b>	: Pimephales promelas (Fish, fresh water)
<b>Exposure period</b>	: 96 hour(s)
<b>Unit</b>	: mg/l
<b>Analytical monitoring</b>	: Yes
<b>Year</b>	: 1996
<b>GLP</b>	: Yes
<b>Test substance</b>	: Light Straight Run Naphtha
<b>Method</b>	: No specific guideline was described as being used to conduct the test, however report references 1991 ASTM method E729-88a and 1982 EPA Support Documents for Environmental Testing: EPA 560/6-82-002. LL <sub>50</sub> and LC <sub>50</sub> calculated using binomial probability analysis. ASTM Special Technical Publication 634. 1977, pp 65-84.
<b>Result</b>	: Mortality (no. of deaths/treatment) at 96 hrs: 0, 0, 0, 6, 20 and 21 in 0, 3.1, 6.3, 13, 25 and 50 mg/l treatments. Abnormal behavior (surfacing, erratic swimming, quiescence) was observed at 96 hrs for 6 organisms in the 13 mg/l treatment. 96-hr LL <sub>50</sub> = 15 mg/l, 6.3-25 mg/l w/ 95% C.I. (as nominal loading rate) 96-hr LC <sub>50</sub> = 0.689 mg/l, 0.289-0.962 mg/l w/ 95% C.I. (measured concentrations) 96-hr NOEL = 6.3 mg/l (nominal); 96-hr NOEC = 0.287 mg/l (measured) based on lack of mortality and abnormal effects for these treatments.
<b>Test condition</b>	: A low boiling point naphtha sample w/ CAS no. 8030-30-6 (different from the sample used in toxicity testing, but similar in composition) was used to validate the analytical method being developed to identify water-soluble hydrocarbons in aqueous 24-hour equilibrated samples. This does not appear to have affected the results of the study. Test solutions were prepared as water accommodated fractions (WAF). Control and dilution water was prepared by blending naturally hard well water with water that had been demineralized by reverse osmosis. Nominal loading rates of 0, 3.1, 6.3, 13, 25 and 50 mg/l were used to prepare test solutions.  WAFS were prepared for each test concentration by mixing the appropriate mass of substance in 9.4 liters of water for 24 hr in 9 liter glass bottles. The bottles were filled to neck height with water to minimize volatility. A measured amount of test substance was added into each bottle, and the bottles were capped tightly with a positive pressure siphoning apparatus. The siphoning apparatus consisted of a Teflon lined neoprene stopper housing two Teflon tubes. One tube extended to the bottom of the bottle for removal of the WAF solution, the other tube ended above the WAF surface, and was used to control air pressure during siphoning. During WAF preparation, Para film was used to seal the external joint between the neoprene stopper and glass bottle, and the bottles were covered with aluminum foil. After stirring for 24 hrs using 25% or less vortex, the contents of the WAF solution bottles were allowed to settle for approximately 45 minutes to two hours, then siphoned by the positive pressure apparatus port and used for testing. Samples were also analyzed by Purge & trap/GC-FID for concentrations of the following: 2-methyl-pentane, cyclohexane, benzene, toluene, ethylbenzene, ortho, meta and para-xylene. Measured test concentrations of the light straight run naphtha were based on the total combined concentrations of all analytes.  Fish were hatched and raised from ABC Laboratories' in-house culture, and were acclimated prior to experimentation for a minimum of 14 days on

## 4. Ecotoxicity

**Id** N. Naphthas  
**Date** 10.12.2001

a 16/8hr light/dark cycle. Test vessels were 3.8 liter glass containers with Teflon lined caps. Fish were acclimated to the test water and temperature approximately 72 hr before the test, and were not fed during this 72 hr period. Two replicates per treatment and 10 organisms per replicate were tested for each treatment and the control, with the exception of the 50 mg/l treatment, where 11 organisms instead of 10 were placed in one replicate. Exposure containers were filled with no headspace and tightly sealed to prevent volatilization. Test solution renewal was performed daily by removal of most of the test solution (leaving approximately one liter of solution to avoid stressing test organisms) and replacing it with fresh WAF solution prepared at least 24 hrs prior to use.

Water temperature was 21-22 °C. Test photoperiod was 16 hrs. light and 8 hr dark. Dissolved oxygen measurements were between 7.3 and 8.8, pH values between 8.1 and 8.3. Hardness values ranged from 134 to 144 mg/l; alkalinity values ranged from 144 to 154 mg/l and conductivity values ranged from 300 to 340 microsiemens.

**Reliability** : (2) valid with restrictions  
Measured concentrations may not represent 100% of components, remaining hydrocarbon components in WAF may be equally toxic and should have been quantitated to determine total measured concentrations.  
12.11.2001 (3)

### 4.2 ACUTE TOXICITY TO AQUATIC INVERTEBRATES

**Type** : 48 Hr Static Acute Toxicity Test with 24Hr Renewal in Closed Systems  
**Species** : Daphnia magna (Crustacea)  
**Exposure period** : 48 hour(s)  
**Unit** : mg/l  
**Analytical monitoring Method** : yes  
: Procedure patterned after:1991 ASTM method E729-88a and 1985 USEPA TSCA Test Guidelines: Daphnid Acute Toxicity Test. Fed. Reg., vol. 50 (No. 188) Sept 27, 1985, 797.1300.  
**Year** : 1996  
**GLP** : yes  
**Test substance** : Light Straight Run Naphtha  
**Method** : Statistical Method: (FT - ME) EL<sub>50</sub> and EC<sub>50</sub> calculated using binomial probability analysis. ASTM Special Technical Publication 634. 1977, pp 65-84.  
**Result** : Immobility (no. of organisms) at 48 hrs: 1, 3, 0, 0, 19 and 20 for 0, 3.0, 6.0, 12, 24 and 48 mg/l treatments.

At the 3 and 12mg/l nominal treatments, 1 and 20 organisms were observed at the bottom of the test chambers, respectively.

48-hr EL<sub>50</sub> = 18 mg/l based upon nominal loading rate (95% C.I. 12 to 24 mg/l)

48 hr EC<sub>50</sub> was 0.65 mg/l (95% C.I. 0.47 to 0.83 mg/L); based on total measured concentrations.

48-hr NOEL = 6.0 mg/l based upon nominal loading rate. 48 hr NOEC was 0.24 ppm based on total measured concentrations.

**Test condition** : Test solutions were prepared as water accommodated fractions (WAF). Control and dilution water were a blend of aged well water and reverse osmosis well water.

WAFS were prepared for each test concentration by mixing the appropriate mass of substance in 2.4L of water for 24 hr in aluminum foil covered 2.5 liter aspirator bottles fitted w/ a port near the bottle bottom. The bottles

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were filled to neck height with water to minimize volatility. A measured amount of test substance was pipetted below the water surface. After stirring for 24 hrs using 25% or less vortex, the contents of the aspirator bottles were allowed to settle for approximately one hour, then drained from the port and used for testing. Samples were also analyzed by purge & trap/GC-FID for concentrations of the following: 2-methyl-pentane, cyclohexane, benzene, toluene, ethylbenzene, ortho, meta and para-xylene. Measured test concentrations of the light straight run naphtha were based on the total combined concentrations of all analytes.

Range finding toxicity studies were conducted at 0.5, 1.0, 10 and 100 mg/l loading, using WAFS which were divided into duplicate aliquots and tested. Definitive toxicity studies were conducted at 3.0, 6.0, 12, 24 and 48 mg/l loading, using WAFS which were divided into duplicate aliquots and tested.

Test vessels were Teflon cap-sealed 8 oz. glass jars with 10 daphnids per jar and were completely filled to overflowing with approximately 273 ml test solution.

During the study test system solutions: dissolved oxygen concentration range: 8.0 to 8.5; pH ranged from 8.3 to 8.4; temperature was 20 to 21 °C; hardness (mg/l) ranged from 132 - 140; alkalinity (mg/l) was 142-150 and conductivity (umhos) values were 280 - 300.

Daphnia magna, were supplied by testing laboratory; age < 24 hours old; obtained from 11 day culture maintained in-house since October 1996.

**Reliability** : (2) valid with restrictions: Measured concentrations may not represent 100% of components, remaining hydrocarbon components in WAF may be equally toxic and should have been quantitated to determine total measured concentrations.

27.10.2001

(1)

### 4.3 TOXICITY TO AQUATIC PLANTS E.G. ALGAE

**Species** : Selenastrum capricornutum (Algae)  
**Exposure period** : 96 hour(s)  
**Unit** : mg/l  
**Analytical monitoring Method** : yes  
: No specific guideline was described as being used to conduct the test, however report references 1991 ASTM method E729-88a and 1982 EPA Support Documents for Environmental Testing: EPA 560/6-82-002.  
**Year** : 1997  
**GLP** : yes  
**Test substance** : Light Straight Run Naphtha  
**Method** : Statistical Method: EL<sub>50</sub> and EC<sub>50</sub> calculated using nonlinear logistics sigmoid model (SAS). All NOEL/NOEC values based on visual review and Dunnett's test for significance.  
**Result** : Percent inhibition on growth determined by cell density (cells/ml):  
96 hour EL<sub>10</sub>=2.7 mg/l (1.9-3.5 mg/l CI @95% )  
96 hour EL<sub>50</sub>=6.4mg/l (5.7-7.1 mg/l CI @95% )  
96 hour EL<sub>90</sub>=15 mg/l (12-18 mg/l CI @95% )  
96 hour NOEL=1.9 mg/l  
  
96 hour EC<sub>10</sub>=0.1 mg/l (0.061-0.15 mg/l CI @95% )  
96 hour EC<sub>50</sub>= 0.26 mg/l (0.22-0.30 mg/l CI @95% )  
96 hour EC<sub>90</sub>=0.66 mg/l (0.50-0.83 mg/l CI @95% )  
96 hour NOEC=0.0326 mg/l

Subcultures of the 31 mg/l treatment cultures were placed in fresh media (no test substance) after acute testing for ten days and indicated that

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growth inhibition was algistatic in this treatment. Conduct of the range-finder and definitive tests were acceptable (no repeats). No excursions from the protocol were noted which would have affected the integrity of the study.

	Nominal (mg/l)
Conc (meas mg/l).	96hr cell density (cells/ml)
Control (<LOQ)	43.58 x10 <sup>4</sup>
1.9 (0.0322)	42.332 x10 <sup>4</sup>
4.0 (0.130)	29.25 x10 <sup>4</sup>
7.8 (0.329)	18.42 x10 <sup>4</sup>
16 (0.704)	1.74 x10 <sup>4</sup>
31 (1.29)	0.04 x10 <sup>4</sup>

**Test condition** : Individual test treatment solutions were prepared as Water Accommodated Fractions (WAFs). Test material was added to 9.4-9.6l of sterilized AAP test media (enriched with 515 mg/l of sodium bicarbonate, 300 µg/l EDTA chelator, pH adjusted to 7.5 + 0.1 with 0.1 NHCl and sterilized by 0.45 micron filtration) in 9.5 liter aspirator bottles. A measured amount of test substance was added into each bottle, and the bottles were capped tightly with a positive pressure siphoning apparatus. The siphoning apparatus consisted of a Teflon lined neoprene stopper housing two Teflon tubes. One tube extended to the bottom of the bottle for removal of the WAF solution, the other tube ended above the WAF surface, and was used to control air pressure during siphoning. During WAF preparation, Para film was used to seal the external joint between the neoprene stopper and glass bottle, and the bottles were covered with aluminum foil. The contents were stirred with Teflon coated stir bars in the mixing vessels which were placed on magnetic stir plates at room temperature. After stirring for 24 hrs using 25% or less vortex, the contents of the WAF solution bottles were allowed to settle for approximately 45 minutes to two hours, then siphoned by the positive pressure apparatus port and used for testing. Test vessels were 125ml glass Erlenmeyer flasks that were completely filled (148 ml) with treatment solution and inoculated with 3 day old algae. Algal cells obtained from testing laboratory cultures grown in sterile, nutrient enriched AAP media. Original algal cultures (stock UTEX-1648) obtained from Dept of Botany, Culture Collection of Algae, University of Texas at Austin, 1996. Cell density of the algal stock culture inoculate was determined prior to study initiation with a hemacytometer cell and compound microscope. Twelve replicates were prepared for each treatment level. Nominal treatment levels were 0, 1.9, 4.0, 7.8, 16 and 31 mg/l The initial algal concentration was 1.0 x 10<sup>3</sup> cells/ml. All test replicates were placed on a shaker table at 100 oscillations per minute during the study and exposed to continuous fluorescent light, illumination 400 +50 ft candles. Triplicate samples were taken daily for cell counts and analytical testing. Cell densities were determined by direct microscopic examination. Samples at 0,24, 48, 72 and 96 hrs were also analyzed by Purge & trap/GC-FID for concentrations of the following: 2-methyl-pentane, cyclohexane, benzene, toluene, ethylbenzene, ortho, meta and para-xylene. Measured test concentrations of the light straight run naphtha were based on the total combined concentrations of all analytes.

**Reliability** : Test temperature was 24-26 °C. Test solution pH ranged from 8.0 to 8.5.  
(2) valid with restrictions  
Measured concentrations may not represent 100% of components, remaining hydrocarbon components in WAFs may be equally toxic and should have been quantitated to determine total measured concentrations.

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(2)

**5.1.1 ACUTE ORAL TOXICITY**

<b>Type</b>	: LD <sub>50</sub>
<b>Species</b>	: rat
<b>Strain</b>	: Sprague-Dawley
<b>Sex</b>	: male/female
<b>Number of animals</b>	: 10
<b>Vehicle</b>	: undiluted
<b>Value</b>	: > 5000 mg/kg bw
<b>Year</b>	: 1982
<b>GLP</b>	: yes
<b>Test substance</b>	: API 81-08
<b>Method</b>	: The test material was administered undiluted, as a single oral dose to groups of 5 male 5 female rats at a single dose level of 5 g/Kg. The dose volume was 7.35 ml/Kg based on an average bulk density of 0.68 g/ml. Food had been withheld from the rats overnight prior to dosing, but they had free access to water. Following dosing, food and water were available ad-lib for a period of 14 days. The animals were observed for clinical signs of toxicity and mortality every hour for the first 6 hours after dosing and twice daily thereafter for 14 days. The rats were weighed the day before dosing and then at 7 and 14 days after dosing. At study termination, all animals were killed with carbon dioxide and subjected to a gross necropsy and abnormalities were recorded.
<b>Result</b>	: No animals died during the study. Clinical signs of intoxication included diarrhea and mucoid diarrhea. Although there was a reduction in body weight following fasting, body weights were increasing by seven and 14 days post dosing. At necropsy, no visible lesions were observed in 4 of 5 males and 2 of 5 females. In the right kidney of one male, the renal pelvis was mildly dilated and a cervical lymph node was enlarged. In the females dilation of the pelvis of the kidney was observed in one animal, a cervical lymph node was enlarged in another animal and in a third animal mild hydrometra of the uterus was observed. The Oral LD50 was greater than 5 g/Kg.
<b>Reliability</b> 27.10.2001	: (1) valid without restriction

(5)

**5.1.2 ACUTE INHALATION TOXICITY**

<b>Type</b>	:	LC <sub>50</sub>
<b>Species</b>	:	rat
<b>Strain</b>	:	Sprague-Dawley
<b>Sex</b>	:	male/female
<b>Number of animals</b>	:	10
<b>Vehicle</b>	:	air
<b>Exposure time</b>	:	4 hour(s)
<b>Value</b>	:	> 5.2 mg/l
<b>Year</b>	:	1986
<b>GLP</b>	:	yes
<b>Test substance</b>	:	API 81-08
<b>Method</b>	:	A group of 5 male and 5 female rats were exposed by whole body inhalation to API 81-08 at a nominal concentration of 5mg/l for 4 hours. After the 4-hour exposure, the rats were observed twice daily for mortality. The animals were weighed prior to exposure and again on days 7 and 14 post exposure. On day 14, all surviving animals were killed by exsanguination following sodium pentobarbital anesthesia and were subjected to a full necropsy. For all animals, including those found dead during the study the lungs were removed, fixed and examined histologically.
<b>Result</b>	:	The actual chamber concentrations were found to be 5.2 mg/l. No deaths occurred during the study. There were no unusual pharmacotoxic signs or behavior observed in the control animals. There was however, a slight incidence of nasal discharge (2/5 males and 1/5 females) during the exposure period but none during the following 14-day observation period. The body weight gains for the males exposed to API 81-08 was considered normal but the female body weight gains were marginally less than that of the controls on day 14 post exposure (8.2% compared to 13.8% increase over pre-exposure body weight). No significant macro or microscopic changes were observed that were considered treatment related.
<b>Reliability</b>	:	(1) valid without restriction
27.10.2001		(7)

## 5.1.3 ACUTE DERMAL TOXICITY

<b>Type</b>	:	LD50
<b>Species</b>	:	rabbit
<b>Strain</b>	:	New Zealand white
<b>Sex</b>	:	male/female
<b>Number of animals</b>	:	8
<b>Vehicle</b>	:	undiluted
<b>Value</b>	:	> 2000 mg/kg bw
<b>Year</b>	:	1982
<b>GLP</b>	:	yes
<b>Test substance</b>	:	API 81-08
<b>Method</b>	:	The skin of the patched area of two rabbits of each sex had been abraded whilst the other two had intact skin. A weighed quantity of undiluted test material (equivalent to a dose of 2 g/Kg) was applied to the dorsal skin of each of 4 male and 4 female rabbits. The applied dose was covered with an occlusive dressing (gauze and an impermeable covering). 24 hours after dosing, the patches were removed, the skin wiped and collars fitted to the rabbits to prevent oral intake of any residual test material. Collars were used to restrain the animals during the application period. The animals were observed for a total of 14 days post-dosing. Body weights were recorded just prior to dosing and again seven and 14 days after dosing. At study termination the animals were killed with carbon dioxide and a gross necropsy was performed. Any abnormalities were recorded.
<b>Result</b>	:	No animals died during the study and no clinical signs of intoxication were observed. Normal growth was observed throughout the study. At necropsy, the only visible lesions seen were on the skin of two animals in which the test site was reddened in one together with crusted appearance and mild crusting was observed in the other rabbit.
<b>Reliability</b>	:	(1) valid without restriction
27.10.2001		(5)

## 5.2.1 SKIN IRRITATION

<b>Species</b>	:	rabbit
<b>Concentration</b>	:	undiluted
<b>Exposure</b>	:	Occlusive
<b>Exposure time</b>	:	24 hour(s)
<b>Number of animals</b>	:	6
<b>PDII</b>	:	1.2
<b>Result</b>	:	slightly irritating
<b>Method</b>	:	Draize Test
<b>Year</b>	:	1982
<b>GLP</b>	:	yes
<b>Test substance</b>	:	API 81-08
<b>Method</b>	:	0.5 ml of undiluted test material was applied to two areas on each rabbit. One area was intact and the other abraded skin. The treated area was then covered with an occlusive dressing. After 24 hours, the dressing was removed and the treated skin was wiped to remove any residue of test material. The degree

of erythema and edema was recorded according to the Draize scale. A second reading of skin responses was made at 72 hours again at 96 hours, 7 and 14 days. Results of the 24 and 72-hour readings were used to determine the Primary Irritation Index.

Body weights were recorded just prior to application of the test material and weekly thereafter throughout the study.

At study termination, all surviving animals were euthanized with an overexposure of carbon dioxide, subjected to a gross necropsy and abnormalities were recorded.

**Result** : The scores for erythema and edema at each of the observation times were as follows:

	Erythema		Edema	
	Intact	Abraded	Intact	Abraded
24 h	0.7	0.7	1.0	0.7
72 h	1.0	0.7	0	0
96 h	0.5	0.2	0	0
7 days	0.3	0.3	0	0
14 days	0	0	0	0

The Primary dermal Irritation index was 1.2

Growth rates were normal throughout the study and there were no visible lesions at necropsy.

**Reliability** : (1) valid without restriction  
27.10.2001

(5)

### 5.2.2 EYE IRRITATION

**Species** : rabbit  
**Concentration** : undiluted  
**Dose** : 0.1 ml  
**Comment** : rinsed after 30 sec exposure in 3 rabbits; 6 rabbits unwashed  
**Number of animals** : 9  
**Result** : not irritating  
**Method** : Draize Test  
**Year** : 1982  
**GLP** : yes  
**Test substance** : API 81-08  
**Method** : 0.1 ml of undiluted test material was placed in the everted lower eyelid of one eye of each of 9 rabbits, the other eye was untreated and served as control.

After 30 seconds the treated eyes of 3 rabbits were washed with lukewarm water for 1 minute. Eyes of the other 6 rabbits were not washed.

Readings of ocular lesions for all animals were made at 1, 24, 48, 72 hours and 7 days after treatment. At the 72 hour and seven day readings, sodium fluorescein was used to aid in revealing possible corneal injury.

Body weights were recorded just prior to treatment and one week afterwards. At termination of the study, the rabbits were euthanized by an overexposure of carbon dioxide and were subjected to a gross necropsy. Any abnormalities found were recorded.

**Result** : One hour after application of the test material the average score for irritation was 2.0 and 0.7 for unwashed and washed eyes respectively and the 24 hour readings were 0.3 and 0 respectively. All other scores throughout the study were 0. Growth was normal throughout the study and there were no visible lesions at necropsy.

**Reliability** : (1) valid without restriction  
27.10.2001

(5)

**5.3 SENSITIZATION**

: No data

**5.4 REPEATED DOSE TOXICITY**

: No data

**5.5 GENETIC TOXICITY 'IN VITRO'**

**Type** : Mouse lymphoma assay  
**System of testing** : Forward mutation assay using cell line L5178Y TK+/-  
**Concentration** : 12.5 - 300 µl/ml  
**Cycotoxic conc.** : 0.05µl/ml without activation; 0.5µl/ml with activation  
**Metabolic activation** : with and without  
**Result** : negative  
**Year** : 1985  
**GLP** : yes  
**Test substance** : API 81-08  
**Method** : Based on a preliminary test, ethanol was selected as solvent for this assay. Concentrations of 0.061 to 1000 µl/ml appeared soluble in the assay medium and no change in color was noted.  
Two positive control substances were used viz Ethyl methane sulphonate (EMS) at a concentration of 0.5 µl/ml in the assay without activation and Dimethylnitrosamine (DMN) at a concentrations of 0.3 µl/ml.

A cytotoxicity study was carried out prior to the mutagenicity assay. The results were difficult to interpret and as a consequence a second study was carried out and the results from this were used to determine the concentrations to be used in the subsequent lymphoma assay. It was established that complete toxicity occurred at 0.05 µl/ml for the non-activated cultures and at 0.5 µl/ml for S-9 activated cultures.

For the mutation assay the lymphoma cells were exposed for 4 hours to test material at concentrations ranging from 0.005 to 0.08 µl/ml without activation and 0.00004 to 0.8 µl/ml with S-9 activation. After exposure to the test material, the cells were allowed to recover for 2 days and then cultures were selected for cloning and mutant selection; TFT was used as the restrictive agent.

Eight non-activated and nine activated cultures were selected for cloning based on their degree of toxicity. The non-activated cultures that were cloned were treated with 0.005, 0.01, 0.015, 0.02, 0.025, 0.03, 0.035 or 0.04 µl/ml and resulted in a range of growth of 6 to 97%. The activated cultures that were cloned were treated with 0.0002, 0.0009, 0.0028, 0.008, 0.02, 0.045, 0.09, 0.7 or 0.75 µl/ml and produced a range of growth from 24 to 109%. Plates were prepared from TFT and from the VC cultures and after 10 to 12 days incubation these plates were scored for total number of colonies per plate. A mutation frequency was

then determined.

The following criteria were used in judging the significance of the activity of the test article.

Positive - if there is a positive dose response and one or more of the 3 highest doses exhibit a mutant frequency which is two-fold greater than background level.

Equivocal - if there is no dose response but any one or more doses exhibit a two-fold increase in mutant frequency over background.

Negative - if there is no dose response and none of the test cultures exhibit mutant frequencies which are two-fold greater than background.

**Result** : The data from each of the 3 trials that were considered valid are tabulated below.

Conc. ( $\mu$ l.ml)	Relative growth (%)	Mutant frequency (10 <sup>-6</sup> units)
------------------------	------------------------	--

TRIAL 1 (No activation)

15.6	118.6	18.1
31.3	64.4	27
62.5	97.8	15.7
125	78.2	24.2
250	20.1	48.8
Solvent control 1	100	13.9
Solvent control 2	100	20.3
Untreated control	191.7	21.5
EMS 0.5 $\mu$ l/ml	17.4	258.2

TRIAL 1 (with S-9 activation)

15.6	78.1	59.1
31.3	53.8	49.3
62.5	63.3	49
125	46.5	79.7
250	46.3	41.6
Solvent control 1	100	34
Solvent control 2	100	24
Untreated control	100.7	30.5
DMN 0.3 $\mu$ l/ml	5	327.5

TRIAL 4 (No activation)

12.5	47.8	19.5
25	49.7	19.2
50	37.7	13.5
100	113.3	8.5
200	86.2	9.3
300	19.8	36.4
Solvent control 1	100	18.3
Solvent control 2	100	18.5
Untreated control	163.9	16.2
EMS 0.5 $\mu$ l/ml	13.5	700

TRIAL 4 (with S-9 activation)

12.5	81.6	52.3
25	60.2	85.7
50	57.3	59.1
100	44.7	63.8
200	71.8	21

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300	3.1	19.3
Solvent control 1	100	23.2
Solvent control 2	100	22.9
Untreated control	78.2	22.2
DMN 0.3 µl/ml	8.8	469.4
<u>TRIAL 5 (with S-9 activation)</u>		
150	76.9	13.6
150	28.4	25.2
200	42.5	24
200	41.9	15.3
250	59.6	24.2
250	15.6	31.1
300	4.9	30.2
300	7.3	32
Solvent control 1	100	27.1
Solvent control 2	100	19.2
Solvent control 3	100	22.4
Solvent control 4	100	24.5
Untreated control 1	63.8	31
Untreated control 2	49.9	29.2
DMN 0.3 µl/ml	16.6	352.9
DMN 0.3 µl/ml	2.2	333.3

The authors judged the data using the following criteria.

### TRIAL 1

Non activation conditions.

The percent relative growths of the assayed treatments ranged from 118.6% to 20.1% which demonstrated non-detectable to moderate toxicities.

The minimum criterion for mutagenesis in this assay was a mutant frequency that exceeding  $37.8 \times 10^{-6}$ . The highest, most toxic treatment (250 µl/ml) induced a mutant frequency that exceeded the minimum criterion, but the increase in the mutant frequency was not accompanied by an increase in the total mutant clones. In order to determine if the increase was repeatable, another non-activation assay was performed.

### Activated assay

Test material was assayed at concentrations ranging from 15.6 to 250 µl.ml. The minimum criterion for mutagenesis in his assay was a mutant frequency exceeding  $54.2 \times 10^{-6}$ . Two treatments induced mutant frequencies that exceeded the minimum criterion, but the increases were sporadic and unrelated to dose or toxicity. Another assay was therefore performed.

### TRIAL 4

Non activated assay.

The test material was assayed at concentrations ranging from 12.5 to 300 µl/ml. In order for a treatment to be considered mutagenic in this assay, a mutant frequency of  $36.5 \times 10^{-6}$  was required. None of the assayed treatments induced mutant frequencies that exceeded the minimum criterion. The observed toxicities ranged from non toxic to moderate toxicity. Although it is preferable to consider results from treatments that induce high toxicity, it was not possible in this assay because of a sharp toxicity curve. The test material was therefore considered non mutagenic without activation in this assay at treatments that approached lethality.

Activated assay.

Concentrations ranging from 12.5 to 300 µl/ml were used in this assay and low to very high toxicity was induced. Sporadic increases in the mutant frequency were induced. The minimum criterion for mutagenesis in this assay was a mutant frequency exceeding  $44.2 \times 10^{-6}$  and three treatments did exceed the minimum criterion (25, 50 & 100 nl/ml). However, the highest concentrations assayed were non-mutagenic. A further assay was therefore performed.

#### TRIAL 5

Activated assay

The test material was assayed in duplicate at concentrations ranging from 150 to 300 nl/ml. A wide range of toxicities were induced.

The sporadic increases in mutant frequency observed in Trials 1 and 4 were not repeatable. None of the treatments induced mutant frequencies that exceeded the minimum criterion of  $48.4 \times 10^{-6}$ .

The test material was therefore considered non-mutagenic with activation in this assay.

**Reliability** : (2) valid with restrictions (multiple assays needed to get usable studies) (6)  
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### 5.6 GENETIC TOXICITY 'IN VIVO'

**Type** : Cytogenetic assay  
**Species** : rat  
**Sex** : male/female  
**Strain** : Sprague-Dawley  
**Route of admin.** : inhalation  
**Exposure period** : 6 hours/day, for 5 days  
**Doses** : 0, 65, 300 & 2050 ppm, nominal concentrations  
**Result** : negative  
**Year** : 1986  
**GLP** : yes  
**Test substance** : API 81-08  
**Method** : Groups of 10 male and 10 female Sprague Dawley rats were exposed (whole body) to nominal concentrations of 65, 300 and 2050 ppm of test material. Exposures were exposed to vapor of the test material 6 hours each day for 5 consecutive days.  
 A positive control group of 10 rats of each sex was given a single dose (0.8 mg/kg) of TEM intraperitoneally 24 hours before sacrifice. A negative control group of 10 rats of each sex was exposed to air only.  
 For the treated and negative control groups bone marrow was harvested 6 hours after the final exposure. For the positive control group the bone marrow was harvested 24 hours after administration of the TEM.  
 Three hours prior to sacrifice by carbon monoxide the rats were given a single intraperitoneal dose of colchicine (4 mg/kg).  
 Immediately after sacrifice, bone marrow was obtained from the tibiae of the animals. The marrow was washed and the cells were fixed before being spread on slides for examination. Routinely 50 spreads were prepared for each animal. The location of cells bearing aberrations was identified. A

mitotic index based on at least 500 cells counted was also recorded. It was calculated by scoring the number of cells in mitosis per 500 cells on each slide read. Slides were scored for chromosomal aberrations.

The authors give the following as the criteria for a positive response and data interpretation. Gaps were not counted as significant aberrations. Indicators of genetic damage were considered to be: Open breaks, configurations resulting from the repair of breaks. The latter included translocations, multiradials, rings, multacentrics etc. Reunion figures such as these were weighted slightly higher than breaks since they usually resulted from more than one break. The number of cells with aberrations per animal was also considered to indicate more genetic damage than those containing evidence of single events. Consistent variations from the euploid number were also considered in the evaluation of mutagenic potential. Often it is not possible to locate 50 suitable metaphase spreads for each animal, even after preparing additional spreads. Possible causes for this appear to be related to cytotoxic effects, which alter the duration of the cell cycle, kill the cell or cause clumping of the chromosomes. Additional information can be gained from the mitotic index, which also appears to reflect cytotoxic effects. The type of aberration, its frequency and its correlation to dose in a given time period was considered in evaluating a test article as being mutagenically positive or negative. Statistical analysis employed a Kruskal-Wallis test of aberrations per cell on a per animal basis.

**Result** : The mean exposure chamber concentrations were found to be: 0, 69±18, 293±42 and 2012±16 ppm.

No signs of toxicity were observed in the rats during the exposure phase of the study. The results of the cytogenetic evaluation are summarized in the following table. NB. Mean values without standard errors are given in the table, although these data are available in the report.

	Exposure concentration (ppm)			Control	
	69	293	2012	Positive	Negative
<u>Total No. of cells</u>					
Male	470	500	410	400	500
Female	500	500	500	474	500
M+F	970	1000	910	874	1000
<u>Frequency of structural aberrations</u>					
Male	.009	.006	.029	>.708	.016
Female	0	>.014	.030	>.970	.008
M+F	.005	>0.01	.029	>.853	.012
<u>Frequency of numerical aberrations</u>					
Male	.012	0	.013	.023	.01
Female	.012	.016	.006	.015	.008
M+F	.01	.008	.01	.019	.009
<u>% Cells with structural aberrations/animal</u>					
1 or more					
Male	.9	.4	2.2	20	.6

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Female	0	1.4	2.6	19	.8
M+F	.5	.9	2.4	19.5	.7
2 or more					
Male	0	.2	.4	11.3	.4
Female	0	.2	.4	14.4	0
M+F	0	.2	.4	12.9	.2
%MI					
Male	6.5	6.6	3.8	1	5.7
Female	4.1	4.8	4.5	1	4.2
M+F	5.3	5.7	4.2	12.9	.2

On the basis of the above data, the authors concluded that there was no evidence of a clastogenic effect of the test material and that there was no significant increase in chromosomal aberration in the dosed animals when compared to the negative controls.

**Reliability** : (1) valid without restriction  
12.11.2001

(8)

### 5.7 CARCINOGENITY

**Species** : Mouse  
**Sex** : Male  
**Strain** : C3H  
**Route of admin.** : Dermal  
**Exposure period** : Lifetime  
**Frequency of treatment** : Twice weekly.  
**Post. obs. period** : None  
**Doses** : 50 µl/application  
**Control group** : Untreated, solvent and positive controls  
**Year** : 1989  
**GLP** : Yes  
**Test substance** : API 81-08  
**Method** : The study summarized here was designed to evaluate the carcinogenicity of 12 different petroleum refinery streams. Only the information relating to the control groups and the group exposed to API 81-08 is included in this summary.

50 µl of undiluted test material was applied twice weekly to the shorn dorsal skin to a group of 47 male mice for 139 weeks. An untreated group of 50 male mice served as untreated controls. A further 50 male mice used as solvent controls received 50 µl of toluene twice weekly for 2 years and BaP at concentrations of 0.01% and 0.05% in toluene was applied twice weekly to a further two groups of 50 male mice.

Body weights of the mice were recorded prior to study initiation, weekly for the first 13 weeks of the study and every 4 weeks until termination at 139 weeks. Observations were made daily for morbidity, mortality and any clinical signs of toxicity. All tumors that developed were recorded and their progression noted. A gross necropsy was performed on all animals dying during the study or killed at termination. Special attention was paid to any dermal and subcutaneous masses.

**Result**

Liver, kidneys, lungs and gonads were weighed for each animal at necropsy and group mean organ weight and organ/body weight ratios were calculated. The test skin site (including dermal and subcutaneous tumors) and control skin site were examined histopathologically as were any suspected dermal and systemic neoplasms.

: The body weights of the mice treated with 81-08 did not differ from those of controls throughout the study. No clinical signs of systemic toxicity were observed in animals treated with 81-08. Observations of preputial gland swelling and penile prolapse increased in all groups with age. Penile prolapse occurred in virtually all mice by 2 years.

Virtually no dermal lesions were observed in the untreated control group. However, mice treated with toluene had an average of 100% incidence of mild or moderate desquamation and an average of 10 to 20% incidence of mild irritation and scabbing. The incidence of scabbing increased up to 40% in older mice.

Dermal lesions in mice treated with 0.01% BaP were similar to the toluene controls. Although they initially had less irritation than the animals treated with toluene, the incidence of mild irritation increased to approximately 50% after 2 years.

To begin with, the mice treated with 0.05% BaP had similar lesions to the toluene controls. However, the incidence of irritation increased to 50 to 100% from weeks 60 to 78.

Dermal lesions in mice treated with 81-08 were very similar to the toluene controls, but with slightly less irritation and scabbing.

Survival of the mice treated with 81-08 was better than that for any of the control groups as shown in the following table

Group	Survival % at month				
	6	12	18	24	30
Untreated	90	90	86	62	18
Toluene	96	94	76	52	10
BaP 0.01%	100	98	84	38	0
BaP 0.05%	100	86	2	0	0
81-08	98	98	89	56	19

A variety of non-neoplastic lesions other than those at the treated skin site were observed at histopathological examination but these occurred in all groups and were not considered to be treatment related.

Lesions at the treated skin site are summarized as follows:

Lesion	Solvent Toluene		Group		
	control	control	BaP 0.01%	BaP 0.05%	81-08
<b>Dermal inflammation</b>					
% affected	0	36	20	24	6
Severity		1.7	1.6	1.6	1.3
<b>Hyperkeratosis</b>					
% affected	0	96	90	92	94
Severity		2	2	2	2

## 5. Toxicity

Id N. Naphthas  
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Acanthosis					
% affected	0	76	80	78	87
Severity		1.8	1.7	1.7	1.9
Epidermal crusting					
% affected	2	84	82	86	75
Severity	1	1.8	1.7	2.1	1.9
Dermal pigmentation					
% affected	0	78	12	2	87
Severity		1.6	1	1	1.9
Dermal fibrosis					
% affected	0	68	54	36	96
Severity		1.6	1.4	1.4	2
Ulceration					
% affected	0	18	12	6	0
Severity		1.6	1.7	1.7	
Dermal neoplasms					
% affected ***	0	8	64	98	6

- \* % mice affected  
 \*\* Severity on a scale  
 1 = minimal  
 2 = mild  
 3 = moderate  
 4 = severe  
 \*\*\* % mice with neoplasms

The percent of mice with systemic neoplasms in the control and 81-08 animals was as follows:

Neoplasm	Group				
	Solvent control	Toluene control	BaP 0.01%	BaP 0.05%	81-08
Primary liver neoplasm					
Benign	4	2	6	0	4
Malignant	32	20	30	4	23
Primary lung neoplasm					
Benign	2	4	2	2	0
Malignant	0	0	0	0	0
Other neoplasms*					
Benign	0	2	0	0	2
Malignant	2	0	0	0	4
Total neoplasms	40	28	38	6	33
Group mean longevity**103		100	96	61	108

- \* Includes malignant lymphomas that were observed in multiple sites, including treated skin  
 \*\* Longevity shown in weeks

The data on dermal neoplasms that developed during the study are summarized in the following table

## 5. Toxicity

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	Solvent Toluene control control	BaP 0.01%	BaP 0.05%	81-08
% mice developing benign dermal neoplasm	0	0	4	4
% mice developing malignant dermal neoplasm	0	8	56	98
& mice with multiple tumors	0	0	28	60
Average tumors/mouse	0	.08	1	1.82
No. mice with metastases	0	0	6	12
Mean latency (weeks)		111	86	49
Tumorigenic activity				113
FEN *		43	49	47
% **		9	65	100

\* FEN = No. of animals alive at the time of appearance of the median tumor plus any mice that died from tumor before that time OR when median latency is over 60 weeks, FEN = No. of animals alive at 60 weeks plus any mice that died with tumor before 60 weeks.

\*\* % of mice developing tumors = No. developing tumors divided by FENx100

The tumor data of the 81-08 group were compared using a Chi square test with the data from the untreated and solvent controls and no significant differences were found.

**Test substance**

: API 81-08 was applied undiluted.  
The solvent control group was treated with toluene.  
Benzo(a)pyrene was applied at concentrations of 0.01 and 0.05% in toluene.

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(9)

### 5.8 TOXICITY TO REPRODUCTION

: No data

### 5.9 DEVELOPMENTAL TOXICITY/TERATOGENICITY

: No data

## 6. References

Id N. Naphthas  
Date 15.11.2001

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## 6. References

Id N. Naphthas  
Date 15.11.2001

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Massachusetts, USA.

**ROBUST SUMMARY  
OF INFORMATION ON**

**Substance Group:** **OLEFINIC NAPHTHAS**

**Summary prepared by:** American Petroleum Institute

**Creation date:** 20 DECEMBER 2000

**Printing date:** 26 OCTOBER 2001

**Date of last Update:** 10 DECEMBER 2001

**Number of Pages:** 27

NB. Reliability of data included in this summary has been assessed using the approach described by Klimisch, et al.

Klimisch, H. J., Andreae, M. and Tillman, U, (1997)

A systematic approach for evaluating the quality of experimental toxicological and ecotoxicological data.  
Regulatory Toxicology and Pharmacology 25, 1-5.

# 1. General Information

Id O. Naphthas  
Date 10.12.2001

## 1.1 GENERAL SUBSTANCE INFORMATION

**Substance type** : petroleum product  
**Physical status** : liquid  
**Remark** : Naphtha streams obtained from the catalytic cracking of heavy distillates into lighter fractions contain saturated, olefinic and aromatic hydrocarbons, mainly in the range C4 to C12 and boil in the range of approximately -20 to 230°C.

The streams that are rich on olefinic components are covered in this robust summary.

Information on several samples of olefinic naphtha streams is included here and their analytical characteristics are listed below.

	API <sup>1</sup> 83-09	LCCN <sup>2</sup>	LCCN-D <sup>3</sup> (vapor)
Parameter			
Gravity (°API)	69.5		
Sulfur (wt %)	0.02		
RVP (psia)	10.3		
IBP (°F)	87		
FBP (°F)	262		
Total Olefins (% by MS)	46.5	42.44	60.09
Total Naphthenes (% by MS)	-	9.14	2.66
Total Aromatics (% by MS)	9.0	15.78	1.85
Total Paraffins (% by MS)	44.5	29.77	35.14
n-Paraffins		5.32	7.11
Benzene (%)	1.2	1.21	0.79
Carbon No. (vol%)			
4		0.44	1.55
5		24.65	62.35
6		28.22	28.50
7		22.52	6.22
8		16.07	1.27
9		6.85	0.10

<sup>1</sup> API 83-09 is a sample of LCCN

<sup>2</sup> LCCN is Light Catalytically cracked Naphtha

<sup>3</sup> LCCN-D is a distillate of LCCN

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## 2. Physico-Chemical Data

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### 2.2 BOILING POINT

The olefinic naphtha streams boil in the range of approximately minus 20 to 230 °C (See section 1.1)

### 2.5 PARTITION COEFFICIENT

**Log pow** : 2.13 - 4 at 25° C  
**Method** : Calculated by LOGKOWWIN ver. 1.65.  
**Year** : 2000  
**GLP** : No  
**Test substance** : Light Catalytically Cracked Naphtha  
**Remark** : Log P values represent the spread of calculated and/or measured values for the C5 to C9 hydrocarbon components found in LCCN (CAS No. 64741-55-5). Detailed hydrocarbon analysis performed by Chevron Research was used to identify the components of this specific LCCN sample. Calculated SAR result for surrogate structures contained in program database (smilecas.dat). Calculation based on an atom/fragment contribution method of W. Meylan and P. Howard.

**Reliability** : (2) Valid with restrictions  
26.06.2001 (11)

### 2.6.1 WATER SOLUBILITY

**Method** : Preparation of Water Soluble Fraction  
**Year** : 1995  
**GLP** : yes  
**Test substance** : Light Catalytically Cracked Naphtha  
**Method** : Water Accommodated Fractions (WAFs) of LCCN were prepared at 50 mg/l loading in freshwater and saltwater and equilibrated for 72 hours in tightly closed systems with minimal headspace.

**Result** : Gas chromatographic analysis of selected components indicated freshwater and saltwater solubilities of 4.6 and 4.3 ppm respectively. Measured test concentrations of the light alkylate naphtha were based on the total combined concentrations of benzene, toluene, ethylbenzene, o-xylene and p-xylene, which represent 13% composition of the test substance. Concentrations for these components reached equilibrium in freshwater and saltwater by 24 and 12 hours respectively.

**Conclusion** : Individual components of complex petroleum substances have specific and differing solubilities. Calculated and measured water solubilities for LCCN components range from approximately 3 to 2000 mg/l. At any particular loading rate, the resulting aqueous concentration of each chemical constituent is a function of the relative volume of the two phases (aqueous and the petroleum mixture), the partition coefficient between the phases, the amount of component present and the maximum water solubility of each component. Initially as the petroleum mixture is added in amounts below the solubility limit of the least soluble component the aqueous concentration increases proportionally until the least soluble component reaches a saturation concentration, and only the more soluble components continue to dissolve, resulting in a two phase system. Further addition of the petroleum mixture results in an aqueous concentration that is a non-linear function of the amount added.

**Reliability** : (2) Valid with restrictions  
26.10.2001 (3) (4) (6) (7) (18)

## 3.1.1 PHOTODEGRADATION

<b>Type</b>	: Calculation
<b>Light source</b>	: Sun light
<b>Rel. intensity</b>	: Based on Intensity of Sunlight
<b>Sensitizer</b>	: OH
<b>Rate constant</b>	: $\text{cm}^3/(\text{molecule}\cdot\text{sec})$
<b>Method</b>	: Calculated by AOPWIN ver. 1.89. Method based on the work of R. Atkinson
<b>Year</b>	: 2000
<b>GLP</b>	: no
<b>Test substance</b>	: Light Catalytically Cracked Naphtha
<b>Remark</b>	: AOPWIN ver. 1.89 calculates atmospheric oxidation half-lives of hydrocarbons in contact with hydroxyl radicals in the troposphere, under the influence of sunlight and in contact with $\text{O}_3$ . Atmospheric oxidation rates were calculated for the C5 to C9 hydrocarbon components found in LCCN CAS No. 64741-55-5. Detailed hydrocarbon analysis performed by Chevron Research was used to identify the components of this specific LCCN sample. Based on a 12-hour day, the range for atmospheric half-lives for LCCN constituents due to OH reactions is: 1.44 hours (1-methyl cyclopentene) to 15.985 days (isopentane). The range for atmospheric half-lives due to $\text{O}_3$ reactions for LCCN olefinic constituents (accounting for approximately 30% composition) is 38.378 min (1-methyl cyclopentene) to 22.920 Hrs (C5 olefins).
<b>Result</b>	: Indirect photolysis: Sensitizer: OH radical Conc. of sensitizer: $1.50\text{E}+06 \text{ OH radicals}/\text{cm}^3$ Rate constant: $0.6691\text{E}-12 \text{ cm}^3/\text{mol}\cdot\text{sec}$ (isopentane) to $89.41 \text{ E}-12$ (1-methyl cyclopentene) Half life: 1.44 hours to 15.985 days  Sensitizer: $\text{O}_3$ radical  Conc. of sensitizer: $7\text{E}1103/\text{cm}^3$ Rate constant: $1.2 \text{ E}-17$ to $43\text{-}17 \text{ cm}^3/\text{molecule}\cdot\text{sec}$ Half life: 38.378 min to 22.920 Hrs.
<b>Reliability</b>	: (2) valid with restrictions
26.06.2001	(12)

## 3.1.2 STABILITY IN WATER

<b>Test substance</b>	: Light Catalytically Cracked Naphtha
<b>Conclusion</b>	: Hydrolysis unlikely
<b>Reliability</b>	: (1) Valid without restriction
26.06.2001	(8)

## 3.3.1 TRANSPORT BETWEEN ENVIRONMENTAL COMPARTMENTS

<b>Type</b>	: Calculated
<b>Media</b>	: soil, air, water, suspended sediment and sediment
<b>Method</b>	: Calculated according to Mackay Level 1
<b>Year</b>	: 2000
<b>Remark</b>	: Model based on chemical fugacity. Physical properties input are those calculated by the EPIWIN Estimation 3.04 program and included in this summary. Values represent the spread of calculated values for C5 to C9 hydrocarbon components found in LCCN, CAS No 64741-55-5. Detailed

### 3. Environmental Fate and Pathways

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hydrocarbon analysis performed by Chevron Research was used to identify the components of this specific LCCN sample.

The majority of LCCN components will partition rapidly to air, where these hydrocarbons will be rapidly oxidized by OH radicals and ozone.

**Result** : Medium % distribution  
Air 97 to 100  
Soil 0.00 to 1.2  
Water 0.01 to 2.7  
Sediment <0.001 to 0.02  
Suspended sediment <0.001 to 0.02

**Conclusion** : This complex petroleum mixture is expected to partition primarily to air.  
**Reliability** : (2) Valid with restrictions  
05.10.2001 (10)

#### 3.5 BIODEGRADATION

**Type** : Aerobic  
**Inoculum** : Mixed, adapted inoculum of domestic activated sludge and soil  
**Contact time** : 56 day  
**Result** : Inherently biodegradable  
**Method** : CONCAWE test method for determining the inherent aerobic biodegradability of oil products. 1996/1997, and modification of ISO/DIS 14593  
**Year** : 1999  
**GLP** : Yes  
**Test substance** : Light Catalytically Cracked Naphtha  
**Method** : Water quality-Evaluation of ultimate aerobic biodegradability of organic compounds in aqueous medium-Method by analysis of inorganic carbon in sealed vessels (CO<sub>2</sub> headspace test)

**Result** : Test material was inherently biodegradable since it achieved >20% biodegradability based on CO<sub>2</sub> production. By day 28 approximately 74% of the test material was degraded, then essentially reached a plateau in degradation rate until day 56. The test was considered valid according to CONCAWE criteria, as >60% biodegradation of positive control (63% actual) was observed by day 14, and total blank CO<sub>2</sub> production at termination was less than 15% of the organic carbon added as test substance. Temperature ranged from 18 to 21 °C, which deviated from the protocol value of 22 ±2°C. This deviation was not expected to have affected the outcome of this study.

Test Day	% Degradation (sd)	
	Hexadecane	Test Material
3	13.93 (1.85)	16.83 (9.56)
7	34.40 (4.54)	30.99 (0.56)
14	63.17 (0.94)	51.66 (3.33)
21	77.26 (6.52)	54.82 (6.24)
28	90.35 (7.14)	74.30 (1.24)
35	85.13 (n=1)	65.02 (1.37)
42	85.21 (n=1)	74.82 (0.54)
49	96.93 (8.94)	70.78 (6.48)
56	94.69 (4.10)	79.22 (12.28)

**Test condition** : Mixed inoculum prepared from soil and activated sludge was incubated with test substance or hexadecane (positive control) during a two-week adaptation period. Triplicate test systems were incubated for both the test substance and hexadecane fed inoculum. Two additional, similar test substances were concurrently incubated in separate 160 ml test systems using the same inoculum and acclimation procedure. Duplicate blank control test systems were prepared which consisted of the mixed inocula in mineral medium but no test or positive control substance. Test medium consisted of glass-distilled water and mineral salts (phosphate buffer, ferric

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chloride, magnesium sulfate, calcium chloride) prepared as described in ISO method.

Acclimation procedure-Activated sludge from aeration basin of Wareham Wastewater Treatment Plant (Mass., U.S.A.) was sieved through 2 mm and centrifuged at 1000 rpm for 10 minutes. After removal of supernatant the concentrated solids were diluted to 5 mg/ml suspended solids with reagent grade water. Soil was collected from a site located in a mixed hardwood and pine forest (Mass., U.S.A.). Site of sampling was cleared of debris and approximately 500 g of soil was obtained at a depth between 5-10cm from the soil surface. Soil was air-dried, sieved through a 2 mm sieve, and analyzed for moisture content (38%).

Test vessels (160 ml serum bottles) were filled with 103 ml of mineral medium containing 50 mg/l of yeast extract and 50 mg/l (dry weight) washed activated sludge, then approximately 0.16g of sieved soil (0.1 g dry wt) was added to each bottle. Test or reference substance was added directly to test systems using a 10 µl Hamilton gas tight syringe. The volume required to achieve the specified mg carbon/L concentrations were calculated based on %carbon and specific gravity of the respective substance. The test substance %carbon (0.8724) and specific gravity (0.7220 mg/µl) information was supplied by the Sponsor. Hexadecane %carbon (0.8496) was calculated from the empirical formula and specific gravity (0.7749 mg/µl) was obtained from Verschuere (1983). Addition of respective substance was performed on an incremental basis to the appropriate vessels as follows: 4, 8 and 8 mg C/l were added on days 0, 7 and 11, respectively. Test vessels were sealed with butyl rubber septa/aluminum crimp caps and incubated at 22 °C (± 2°C) in the dark.

Biodegradation by CO<sub>2</sub> determination-test initiation and procedure. On day 14 of the acclimation phase, all test system inoculum from blanks, positive control, and each of the three test substances was combined and filtered through glass wool, and aerated prior to use. The aerated mixed inoculum was then added to mineral medium to achieve 10% concentration based on total volume(100 ml inoculum/l).

Test vessels (160 ml serum bottles) were filled with 103 ml of inoculated mineral medium. Respective test systems were dosed with either test substance or hexadecane as described for the acclimation procedure to achieve 20 mg carbon/l concentration.

Duplicate test systems for each test substance, positive control and blank treatments were prepared for sacrifice at weekly sampling intervals for subsequent CO<sub>2</sub> analysis. After test system preparation, all vessels were placed in a walk-in chamber and incubated in the dark at 22 °C (± 2°C).

On days 3, 7, 14, 21, 28, 35, 42, 49 and 56, 1ml of conc. H<sub>3</sub>PO<sub>4</sub> was injected through the septum of each sacrificed test vessel. The acidified samples were shaken for 1 hr at 200 ppm, and then analyzed for CO<sub>2</sub> using gas chromatography-thermal conductivity detection. Quantitation of inorganic mg C/l evolved was determined by linear regression analysis based on response factors for sodium carbonate standards spanning 1-30 mg carbon/l concentrations.

Reliability  
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: (1) valid without restriction

(14)

## 4. Ecotoxicity

**Id** O. Naphthas  
**Date** 10.12.2001

### 4.1 ACUTE/PROLONGED TOXICITY TO FISH

**Type** : 96 Hr Static Acute Toxicity Test w/ Daily Renewal  
**Species** : Pimephales promelas (Fish, fresh water)  
**Exposure period** : 96 hour(s)  
**Unit** : mg/l  
**Analytical monitoring Method** : Yes  
: No specific guideline was described as being used to conduct the test, however report references 1991 ASTM method E729-88a and 1982 EPA Support Documents for Environmental Testing: EPA 560/6-82-002.

**Year** : 1995  
**GLP** : Yes  
**Test substance** : TS: Light Catalytically Cracked Naphtha  
**Method** : Statistical Method: (FT - ME) LL<sub>50</sub> and LC<sub>50</sub> calculated using binomial probability analysis. ASTM Special Technical Publication 634. 1977, pp 65-84.

**Result** : Mortality (no. of deaths/treatment) at 96 hrs: 0, 1, 0, 0, 4 and 20, respectively 0, 3.0, 7.4, 15, 37 and 74 mg/l treatments. All surviving organisms exhibited normal behavior.

96-hr LL<sub>50</sub> = 46 mg/l, 37-74 mg/l w/ 95% C.I. (as nominal loading rate)

96-hr LC<sub>50</sub> = 4.1 mg/l, 3.2-7.0 mg/l w/ 95% C.I. (measured concentrations)

**Test condition** : Test solutions were prepared as water accommodated fractions (WAF). Control and dilution water was Mobil Technical Center well water. Nominal loading rates of 0, 3.0, 7.4, 15, 37 and 74 mg/l were used to prepare test solutions.

WAFS were prepared for each test concentration by mixing the appropriate mass of substance in 9.4 liters of water for 24 hr in 9 liter glass bottles. The bottles were filled to neck height with water to minimize volatility. A measured amount of test substance was added into each bottle, and the bottles were capped tightly with a positive pressure siphoning apparatus. The siphoning apparatus consisted of a Teflon lined neoprene stopper housing two Teflon tubes. One tube extended to the bottom of the bottle for removal of the WAF solution, the other tube ended above the WAF surface, and was used to control air pressure during siphoning. During WAF preparation, Para film was used to seal the external joint between the neoprene stopper and glass bottle. After stirring for 24 hrs using 25% or less vortex, the contents of the WAF solution bottles were allowed to settle for approximately 45 minutes to two hours, then siphoned by the positive pressure apparatus port and used for testing. Samples were also analyzed by Purge & trap/GC-PID for concentrations of the following: benzene, toluene, ethylbenzene, and p-xylene, which represent 13% composition of the test substance. Measured test concentrations of the light catalytically cracked naphtha were based on the total combined concentrations of all analytes.

Fish were hatched and raised in-house, and were acclimated prior to experimentation for a minimum of 14 days on a 16/8hr light/dark cycle. Test vessels were 3.8L glass containers with Teflon lined caps. Two replicates per treatment and 10 organisms per replicate were tested for each treatment and the control. Exposure containers were filled with no headspace and tightly sealed to prevent volatilization. Test solution renewal was performed daily by removal of most of the test solution (leaving adequate volume to avoid stressing test organisms) and replacing it with fresh WAF solution prepared at least 24 hrs prior to use.

Water temperature was 21.4-21.8 °C. Test photoperiod was 16 hrs. light

## 4. Ecotoxicity

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**Reliability** : and 8 hr dark. Dissolved oxygen measurements were between 5.2 and 8.6, pH values between 7.61 and 8.2.  
: (2) Valid with restrictions  
NOEC values not reported as sublethal effects and moderate mortality (20%) were observed at the 37-ppm (nominal loading) treatment, which is reported to be the NOEC.

26.10.2001 (15)

### 4.2 ACUTE TOXICITY TO AQUATIC INVERTEBRATES

**Type** : 48 Hr Static Acute Toxicity Test with 24Hr Renewal in Closed Systems  
**Species** : Daphnia magna (Crustacean)  
**Exposure period** : 48 hour(s)  
**Unit** : mg/l  
**Analytical monitoring** : yes  
**Method** : No specific guideline was described as being used to conduct the test, however report references 1991 ASTM method E729-88a and 1982 EPA Support Documents for Environmental Testing: EPA 560/6-82-002.

**Year** : 1995  
**GLP** : yes  
**Test substance** : TS: Light Catalytically Cracked Naphtha  
**Method** : Statistical Method: (FT - ME) EL<sub>50</sub> and EC<sub>50</sub> calculated using binomial probability analysis. ASTM Special Technical Publication 634. 1977, pp 65-84.

**Result** : Mortality (no. of deaths/treatment) at 48 hrs: 0, 0, 0, 20, 20 and 20 for 0, 6.4, 13, 25, 51 and 102 mg/l treatments.

48-hr EL<sub>50</sub> = 18 mg/l (95% C.I. 13 to 25 mg/l) based upon nominal loading rate.  
48 hr EC<sub>50</sub> was 1.4 ppm (95% C.I. 0.99 to 1.95 mg/l); based on total measured concentrations.

48-hr NOEC = 13 mg/l based upon nominal loading rate  
48 hr EC<sub>50</sub> was 0.99 ppm based on total measured concentrations.

**Test condition** : Test solutions were prepared as water accommodated fractions (WAF). Control and dilution water was aged well water.

WAFS were prepared for each test concentration by mixing the appropriate mass of substance in 1.2L of water for 24 hr in aluminum foil covered 1 liter aspirator bottles fitted w/ a port near the bottle bottom. The bottles were filled to neck height with water to minimize volatility. A measured amount of test substance was pipetted below the water surface. After stirring for 24 hrs using 25% or less vortex, the contents of the aspirator bottles were allowed to settle for approximately 45 minutes, then drained from the port and used for testing. Samples were also analyzed by Purge & trap/GC-FID for concentrations of the following: benzene, toluene, ethylbenzene, and p-xylene, which represent 13% composition of the test substance. Measured test concentrations of the light catalytically cracked naphtha were based on the total combined concentrations of all analytes.

Range finding toxicity studies were conducted at 1.3, 10 and 102 mg/l loading, using WAFS, which were divided into duplicate aliquots and tested. Definitive toxicity studies were conducted at 6.4, 13, 25, 51 and 102 mg/l loading, using WAFS, which were divided into duplicate aliquots and tested.

Test vessels were teflon cap-sealed 265 ml glass jars with 10 daphnids per jar and were completely filled with test solution.

During the study test system solutions: dissolved oxygen concentration

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range: 8.0 to 8.6; pH ranged from 7.94 to 8.40; temperature was 19.1 to 20.2 Deg C; hardness (mg/l) ranged from 172 - 180; alkalinity (mg/l) was 124-132 and conductivity (umhos) values were 360 - 405.

**Reliability** : (2) valid with restrictions  
Measured concentrations represent only 13-20% of components, remaining hydrocarbon components in WAF may be equally toxic and should have been quantitated to determine total measured concentrations.

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### 4.3 TOXICITY TO AQUATIC PLANTS E.G. ALGAE

**Species** : Selenastrum capricornutum (Algae)  
**Exposure period** : 96 hour(s)  
**Unit** : mg/l  
**Analytical monitoring Method** : yes  
EPA. 1982. Guidelines and Support Documents for Environmental Effects Testing. EPA 560/6-82-002. Sections EG-8, ES-5.  
**Year** : 1995  
**GLP** : yes  
**Test substance** : Light Catalytically Cracked Naphtha  
**Method** : Statistical Method: LL<sub>50</sub> and LC<sub>50</sub> calculated using probit analysis. ASTM Special Technical Publication 634. 1977, pp 65-84. All NOEL/NOEC values calculated using Fisher's exact test.

**Result** : Percent inhibition on growth determined by cell density (cells/ml):  
96 hour EL<sub>50</sub>=64 mg/l (44-111 mg/l CI @95% )  
96 hour EC<sub>50</sub>= 4.6mg/l (2.9-8.8 mg/l CI @95% )  
96 hour NOEL=51 mg/l  
96 hour NOEC=3.5 mg/l  
Subcultures placed in fresh media (no test substance) after acute testing for six days indicated that growth inhibition was algistatic in all treatments, with the exception of the 102 ppm, which was determined to be algicidal. No excursions from the protocol were noted. However, range finding and two previous definitive tests were performed and considered inconclusive due to inconsistencies in control and treatment cell densities, which presumably were resolved by modification of the AAP media. Additionally, control growth showed a lag during the first 72 hours of the study.

Nominal (mg/l)		
Conc. (meas.mg/l).	96hr cell density (Cells/ml)	(% Inhibition)
Control	8.4 x10 <sup>3</sup>	na
6.4 (0.093)	3.2 x10 <sup>4</sup>	- 281.1
13 (0.130)	9.73x10 <sup>3</sup>	- 16.0
25 (0.429)	1.99x10 <sup>4</sup>	-136.9
51 (1.87)	1.36x10 <sup>3</sup>	53.0
102 (4.85)	2.59x10 <sup>3</sup>	69.2

**Test condition** : Individual test treatment solutions were prepared as Water Accommodated Fractions (WAFs). Test material was added to 4.4L of sterilized AAP test media (enriched with 515 mg/l of sodium bicarbonate, pH adjusted to 7.5 + 0.1 with 0.1NHCl and sterilized by 0.22 micron filtration) in 4.0 liter aspirator bottles. The mixing vessels were sealed with foil covered stoppers and mixed on magnetic stir plates with Teflon coated stir bars for approximately 24 hours at room temperature in a hood darkened with aluminum foil. After mixing the solutions were allowed to settle for one hour and the WAF was removed and used for testing. Test vessels were

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125ml glass Erlenmeyer flasks that were completely filled (140 ml) with treatment solution and inoculated with algae. Algal cells obtained from testing laboratory cultures grown in sterile, nutrient enriched AAP media, and transferred every 4-8 days to fresh media. Original algal cultures obtained from American Type Culture Collection (ATCC Strain 22662), Rockville, MD, September 1995. Cell density of the algal stock culture inoculate was determined prior to study initiation with a hemacytometer cell and compound microscope. Twelve replicates were prepared for each treatment level. Nominal treatment levels were 0, 6.4, 13, 25, 51 and 102 mg/l The initial algal concentration was  $1.0 \times 10^3$  cells/ml. All test replicates were placed on a shaker table at 100 oscillations per minute during the study and exposed to continuous fluorescent light, illumination at  $400 \pm 50$ -ft candles. Triplicate samples were taken daily for cell counts and analytical testing. Cell densities were determined by direct microscopic examination. Samples at 0,24, 48, 72 and 96 hrs were also analyzed by Purge & trap/GC-FID for concentrations of the following: benzene, toluene, ethylbenzene, o-xylene and p-xylene, which represent 13% composition of the test substance. Measured test concentrations of the light catalytically cracked naphtha were based on the total combined concentrations of all analytes.

Test temperature was  $24 \pm 2$  °C. The pH was 7.5 at test initiation, pH value at test termination not included in report.

### Reliability

: (2) valid with restrictions  
Measured concentrations may not represent 100% of components, remaining hydrocarbon components in WAFs may be equally toxic and should have been quantitated to determine total measured concentrations. (17)

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### 5.1.1 ACUTE ORAL TOXICITY

<b>Type</b>	:	LD <sub>50</sub>
<b>Species</b>	:	Rat
<b>Strain</b>	:	Sprague-Dawley
<b>Sex</b>	:	Male/female
<b>Number of animals</b>	:	5
<b>Vehicle</b>	:	Undiluted
<b>Value</b>	:	> 5000 mg/kg bw
<b>Year</b>	:	1986
<b>GLP</b>	:	Yes
<b>Test substance</b>	:	LCCN (Sample API 83-20)
<b>Method</b>	:	Groups of five male and five female fasted rats were given API 83-20 as a single oral dose of 5 g/kg. The animals were then allowed food and water <i>ad-libitum</i> and were observed hourly for clinical signs for the first 6 hours after dosing. Observation was twice daily thereafter for 14 days. Body weights were recorded at 7 and 14 days after administration of test material. At the end of the study, the animals were killed and subjected to a gross necropsy and any abnormalities were recorded.
<b>Result</b>	:	There were no mortalities during the study. Body weights had increased by day 7 after dosing and further increases were recorded 14 days after dosing. Clinical signs of toxicity were observed during the 24 hours immediately after dosing and appeared normal thereafter. Clinical signs included: hypoactivity, ataxia, diarrhea, lacrimation, yellow-stained anal area, excessive salivation and respiratory congestion. There were no treatment-related lesions observed at necropsy.
<b>Conclusion</b>	:	Oral LD <sub>50</sub> was greater than 5 g/kg for males and females.
<b>Reliability</b>	:	(1) Valid without restriction
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(1)

### 5.1.2 ACUTE INHALATION TOXICITY

<b>Type</b>	:	LC <sub>50</sub>
<b>Species</b>	:	Rat
<b>Strain</b>	:	Sprague-Dawley
<b>Sex</b>	:	Male/female
<b>Number of animals</b>	:	5
<b>Vehicle</b>	:	Air
<b>Exposure time</b>	:	4 hour(s)
<b>Value</b>	:	> 5.3 mg/l
<b>Year</b>	:	1987
<b>GLP</b>	:	Yes
<b>Test substance</b>	:	LCCN (Sample API 83-20)
<b>Method</b>	:	A group of 5 male and 5 female rats were exposed by whole body inhalation to API 83-20 at a nominal concentration of 5mg/l for 4 hours. After the 4 hour exposure the rats were observed twice daily for mortality. The animals were weighed prior to exposure and again on days 7 and 14 post exposure. On day 14 all surviving animals were killed and subjected to a gross post-mortem examination. For all animals, including those found dead during the study, the lungs were removed,

- Result** : fixed and examined histologically.  
: The mean analytical exposure concentration was measured and found to be  $5.28 \pm 0.55$  mg/l. Gravimetric samples, collected on glass fiber filters suggested little or no aerosol in the chamber. Most animals exhibited languid behavior and squinted eyes during the second hour of the exposure. Polypnea was observed in all animals when removed from the chamber at the one-hour post exposure observation period. Rhinorrhea was exhibited by two animals on day two of the test. All animals appeared normal subsequently and there were no mortalities during the study. With the exception of one animal (female) all animals had body weights that were considered unremarkable. There were no remarkable gross or microscopic findings.
- Reliability** : (1) valid without restriction  
26.10.2001 (2)

### 5.1.3 ACUTE DERMAL TOXICITY

- Type** : LD<sub>50</sub>  
**Species** : Rabbit  
**Strain** : New Zealand white  
**Sex** : Male/female  
**Number of animals** : 4  
**Vehicle** : Undiluted  
**Value** : > 3000 mg/kg bw  
**Year** : 1986  
**GLP** : Yes  
**Test substance** : LCCN (Sample API 83-20)  
**Method** : The skin of the patched area of four rabbits of each sex had been abraded whilst the other four had intact skin. A weighed quantity of undiluted test material was applied to the dorsal skin of each of 4 male and 4 female rabbits at a dose level of 2.0 g/kg. The applied dose was covered with an occlusive dressing (gauze and an impermeable covering). 24 hours after dosing, the patches were removed, the skin wiped and collars fitted to the rabbits to prevent oral intake of any residual test material. The collars were removed 24 hours later and the animals were observed for a total of 14 days post-dosing. At study termination, the animals were killed with carbon dioxide and a gross necropsy was performed. Any abnormalities were recorded. Since there were 2 mortalities recorded in this study, the experiment was repeated using a dose level of 3 g/kg.
- Result** : A pain response was elicited from all animals during application of the test material to the skin. Dermal irritation ranging from slight to severe was observed for erythema and slight to marked for atonia, desquamation, fissuring and coriaceousness. At the 2 g/kg dose level, clinical observations included: diarrhea, anorexia and hypoactivity and one female in this dose group died on day 7 following dosing. There were no clinical signs of toxicity in the 3 g/kg dose group and no animals in this group died. At necropsy, skin lesions were observed more frequently in the 2 g/kg group than the 3 g/kg group.
- Conclusion** : LD<sub>50</sub> was greater than 3 g/kg for both male and female rabbits.

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**Reliability** : (1) Valid without restriction  
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### 5.1.4 ACUTE TOXICITY, OTHER ROUTES

**Remark** : Not relevant  
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### 5.2.1 SKIN IRRITATION

**Species** : Rabbit  
**Concentration** : Undiluted  
**Exposure** : Occlusive  
**Exposure time** : 24 hour(s)  
**Number of animals** : 6  
**PDII** : 3.7  
**Result** : Moderately irritating  
**Method** : Draize Test  
**Year** : 1986  
**GLP** : Yes  
**Test substance** : LCCN (Sample API 83-20)  
**Method** : 0.5 ml of undiluted test material was applied to the shorn skin in two areas on each rabbit. One area was intact and the other abraded skin.  
The treated area was then covered with an occlusive dressing.  
After 24 hours, the dressing was removed and the treated skin was wiped to remove any residue of test material. The degree of erythema and edema was recorded according to the Draize scale. A second reading of skin responses was made at 72 hours and again at 96 hours, 7 and 14 days. Results of the 24 and 72-hour readings were used to determine the Primary Irritation Index.

**Result** : The scores for erythema and edema were marginally greater for abraded skin than intact skin, but the difference was not biologically significant.  
Scores for abraded skin at each of the observation intervals were:

Time	Erythema	Edema	Irritation score*
24 hours	1.8	2.0	3.5
72 hours	2.3	1.7	3.8
96 hours	1.5	1.5	2.6
7 days	1.2	0.2	1.2
14 days	0.0	0.0	0.0

\* Irritation score calculated as the sum of irritation scores for each test site divided by the number of animals at each observation period.  
PII is the sum of the 24- and 72- hour total irritation scores divided by 2

**Reliability** : (1) Valid without restriction  
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### 5.2.2 EYE IRRITATION

**Species** : Rabbit  
**Concentration** : Undiluted

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**Dose** : .1 ml  
**Comment** : Other: See method  
**Number of animals** : 9  
**Method** : Draize Test  
**Year** : 1986  
**GLP** : Yes  
**Test substance** : LCCN (Sample API 83-20)  
**Method** : 0.1 ml of undiluted test material was applied to the corneal surface of one eye of each of 9 rabbits; the other eye was untreated and served as control.  
After 20 to 30 seconds, the treated eyes of 3 rabbits were washed with lukewarm water for 1 minute. Eyes of the other 6 rabbits were not washed.  
Readings of ocular lesions for all animals were made at 1, 24, 48, 72 hours and 7 days after treatment. Sodium fluorescein was used to aid in revealing possible corneal injury.

**Result** : No pain response was elicited from any of the animals when the test material was applied to the corneal surface.  
The primary eye irritation score (=total eye irritation score for all animals divided by the number of animals) was 1.0 after 1 hour for those animals with unwashed eyes compared to 3.3 for those whose eyes had been washed. An irritation score of zero was recorded at all other times.  
Neither iridial nor corneal irritation resulted from application of the test material.

**Reliability** : (1) Valid without restriction  
26.10.2001 (1)

### 5.3 SENSITIZATION

**Type** : Buehler Test  
**Species** : Guinea pig  
**Concentration** : Induction undiluted occlusive epicutaneous  
Challenge 25 % occlusive epicutaneous  
**Number of animals** : 10  
**Vehicle** : Paraffin oil  
**Result** : Not sensitizing  
**Classification** : Not sensitizing  
**Year** : 1986  
**GLP** : Yes  
**Test substance** : LCCN (Sample API 83-20)  
**Method** : 0.4 ml of undiluted test material was applied under an occlusive dressing to the shorn skin of 10 male and 10 female animals. 6 hours after application, the dressings were removed and the skin wiped to remove residue of test material. The animals received one application each week for 3 weeks. The same application site was used each time. 2 weeks following the third application, a challenge dose (0.4 ml of a 25% mixture in paraffin oil) was applied in the same manner as the sensitizing doses. A previously untreated site was used for the challenge application.  
The application sites for sensitizing and challenge doses were read for erythema and edema 24 and 48 hours after patch removal. To assist in the reading of the response to the final challenge dose the test site was depilated 3 hours prior to reading by using a commercially available depilatory cream.

Positive control (2,4-dinitrochlorobenzene, as a 0.3% w/v

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### Result

solution in 80% aqueous ethanol), vehicle control (paraffin oil) and naive control groups were included in this study and the procedure for these was the same as for the test groups.

: No skin reactions were observed following the application of the challenge dose in either the naive controls or the group that had been exposed to test material. Scores of 0.2, 0.3 and 0.5 for erythema were recorded for the paraffin oil controls.

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In contrast, all positive control animals developed a skin response following the challenge procedure.

: (1) Valid without restriction

(1)

## 5.4 REPEATED DOSE TOXICITY

**Species** : Rat  
**Sex** : Male/female  
**Strain** : Sprague-Dawley  
**Route of admin.** : Inhalation  
**Exposure period** : 15 weeks  
**Frequency of treatment** : 6 hours/day, 5 days/week  
**Post obs. period** : 4 weeks  
**Doses** : Target: 750, 2500 & 7500 ppm. Actual: 756, 2507 & 7533 ppm  
**Control group** : Yes  
**Method** : EPA OTS 798.2450  
**Year** : 2001  
**GLP** : Yes  
**Test substance** : LCCN-D (Distillate of LCCN)  
**Method** : Groups of 16 male and 16 female rats underwent whole body exposures to 750, 2500 and 7500 ppm LCCN-D. Exposures were for 6 hours each day, 5 days per week, for at least 65 exposures, over a period of 15 weeks.  
Extra groups of 16 rats of each sex were exposed to the high dose level and for a recovery control group. These animals were maintained untreated for 28 days following cessation of the 15 weeks exposure.

Neurobehavioral evaluations of motor activity and functional activity were performed pretest and during weeks 5, 9, 14/15 and after the 4 week recovery period for the recovery animals. Animals were not exposed to LCCN-D during these tests.

Following 15 weeks of exposure, 16 animals/sex/group were necropsied and microscopic examination was performed on selected tissues. Nervous tissue from 6 rats/sex/group was also examined microscopically.

At the end of the 4-week recovery period, 16 animals of each sex from the high and control groups were necropsied and selected tissues were examined microscopically.

During the study, clinical observations were made twice daily. Ophthalmoscopic evaluations were performed pretest and just prior to the scheduled sacrifices at 15 weeks and 20 weeks (recovery groups). Body weights and food consumption were measured throughout the study. Blood samples were taken from 10 fasted rats/sex/group at 14

and 18 weeks for hematological and clinical chemical measurements.

At termination (after 15 weeks exposure for the main study and after 19 weeks for the recovery animals) all animals were killed and subjected to a complete macroscopic examination. 10 animals of each sex were designated for non-neuropathological examination and 6 of each sex for neuropathological examination.

For the non-neuropathology animals, the following organs were weighed: adrenals, brain, heart, kidneys, liver, lung, ovaries, prostate, spleen, testes (with epididymes), thymus and uterus. Brain lengths and widths were measured for each rat.

A wide range of tissues (39) was removed from the control and high dose animals and were fixed and examined histopathologically. Additionally, kidneys from selected animals were stained with Mallory-Heidenhain and examined. Tissues were also removed from the nervous system (central and peripheral) of all animals for subsequent special staining and histopathological examination. Animals designated for neuropathological examination were subjected to a detailed examination of central and peripheral nervous tissues.

Neurobehavioral studies were undertaken as follows:

#### Motor activity

Locomotor activity was monitored as the number of beam breaks in an activity box. Monitoring sessions were for 60 minutes, divided into twelve 5-minute intervals. Evaluation was made pretest and during weeks 5, 9, 15 and at the end of the 4 week recovery period. [A detailed description of the evaluation and analysis is provided in the publication but is not included here.]

#### Functional Operational Battery

An assessment of the following was made:

Home cage evaluations for Posture, vocalization, palpebral closure.

Handling evaluations for reactivity to general stimuli, signs of autonomic function.

Open field behavior: arousal level, gait, urination and defecation frequency, convulsions, tremor, abnormal behavior, piloerection and exophthalmos.

Reflex assessments for: response to visual and auditory stimuli, tail pinch, pupillary function.

Animals were also evaluated for fore limb and hind limb grip strength, landing foot splay and air righting ability.

#### Result

: No exposure-related clinical observations were noted either during exposure or during non-exposure periods and no ocular abnormalities were observed.

Although the males in the high dose group were slightly lighter than the controls, the difference was not significant. In the females however, the difference was statistically significant. At the end of the 4-week recovery period body weights of the high dose males and females were comparable to the corresponding controls. During the 4-week recovery period, the high dose males and females had food consumption that was greater

(statistically significant) than controls.  
At 15 weeks, the following hematological changes were recorded.

- 7500 ppm males
  - Decreased hemoglobin concentration (8%)
  - Decreased hematocrit (7%)
- 2500 ppm males
  - Decreased MCHC (3%)
- 7500 ppm females
  - Decreased MCHC (4%)

After the 4-week recovery period, all hematological values were considered normal.

At 15 weeks, there were no abnormal clinical chemistry values. After the 4-week recovery period however, glucose and albumin was raised in the 7500-ppm females by 21 and 15% respectively. Since the values were within the normal range, they were not considered toxicologically significant.

Neurobehavioral studies

There was no evidence of any effect on motor activity either after 15 weeks exposure or after the 4-week recovery period.  
There was no evidence of a treatment-related effect in the functional operational battery that was carried out.

Pathology

With the exception of those listed below, absolute and relative organ weights were not affected by treatment.

Parameter	2500 ppm	Group 7500 ppm	Recovery
<b>MALES</b>			
Abs Kidney		21% up	
Rel Kidney	15% up	32% up	
Rel Liver		23% up	
<b>FEMALES</b>			
Rel Kidney		18% up	
Abs Liver			24% up
Rel Liver		12% up	
Rel Brain			9% down

There were no microscopic findings in either the liver or brain of the groups in which organ weight changes had been recorded.

The only treatment-related microscopic changes were found in the kidneys and nasal turbinates as follows.

Nasal turbinates

Goblet cell hypertrophy/hyperplasia and hyperplasia of the respiratory epithelium were seen in a number of animals from all groups. Based on the incidence and/or severity both findings were slightly more pronounced in the 7500 ppm group and were seen primarily in the anterior region of the nasoturbinat tissues. At the end of the 4-week recovery, the incidence and severity of goblet cell hypertrophy/hyperplasia and hyperplasia of the respiratory epithelium were considered to be similar in high dose and control animals. In the 7500 ppm females severity of the findings in the nasoturbinat tissues was slightly less after recovery than at the end of the

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15 week exposure. In the males the severity and incidence of the effects was the same in the nasoturbinal tissues after the recovery period as it was after 15 weeks exposure. These findings are considered indicative of exposure to a mild irritant.

### Kidney

At the end of 15 weeks exposure intracytoplasmic eosinophilic/hyaline droplets were seen in the epithelium lining the proximal convoluted tubules of the kidneys. It was seen in several males of the control and 750 ppm group and all males in the 2500 and 7500 ppm groups. At the end of the 4-week recovery period the incidence and severity in controls and 7500 ppm males was similar.

Several males from the 7500 ppm group and one from the 2500 ppm group had cortico-medullary tubules which were dilated and contained granular casts. None of these findings were found in the females and furthermore, none were observed at the end of the 4 week recovery period.

Subacute/chronic interstitial inflammation was seen in several rats from all groups, except the control group males and the 7500 ppm females. In the males from the exposure groups, the incidence showed a dose-related increase. Basophilic convoluted tubules were seen in a small number of males from the 2500 and 7500 ppm groups.

At the end of the 4-week recovery period there was an indication of some reversibility of the kidney effects.

In the post exposure animals, brain length and width measurements showed no test-material-related effects.

**Reliability**  
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: (1) Valid without restriction

(9)

**Species**

: Rat and Mouse

**Sex**

: Male/female

**Strain**

: Rat - Sprague Dawley, Mouse - CD-1

**Route of admin.**

: Inhalation

**Exposure period**

: 13 weeks

**Frequency of treatment**

: 6 hours/ day, 5 days/week for 13 weeks

**Doses**

: Target: 500, 2000 & 8000. Actual: 530, 2060 & 7690 mg/m<sup>3</sup>

**Control group**

: Yes

**Year**

: 1996

**GLP**

: No data

**Method**

: Groups of 10 rats of each sex and 10 mice of each sex were individually housed in inhalation chambers. The rats and mice underwent whole body exposures to LCCN vapors. Exposures were for 6 hours/day, 5 days/week for approximately 13 weeks at nominal concentrations of 500, 2000 or 8000 mg/m<sup>3</sup>.

Extra groups of 10 rats and mice of each sex served as sham and untreated controls.

Food and water was available ad-lib, except during the exposure periods.

Clinical observations were made regularly and body weights were recorded weekly.

At the end of the 13 weeks exposure, the rats were fasted for 16 hours before blood samples were taken for hematological and clinical chemical measurements.

All animals were then sacrificed and necropsied. Organs were weighed and a wide range of tissues fixed for subsequent histology and microscopic examination. The wet and dry weights of the right apical and right middle lung lobes were also recorded. The cauda epididymis of the control and high dose male rats was used to determine the morphology and number of sperm and the left testis was used to

determine the number of testicular spermatids.

The following tissues from the high dose and sham treated animals were examined histologically: adrenals, kidney, bone and marrow (sternum), pancreas, brain, submaxillary salivary gland, eye, optic nerve, spleen, heart, stomach, colon, testes or ovaries, duodenum, kidneys, thymus, thyroid, liver, tracheobronchial lymph nodes, lung (left lobe), nasal turbinates, muscle, urinary bladder, sciatic nerve, and any gross lesions. Additional sections included lung from untreated controls and kidney from 0, 2060 and the 7690 mg/m<sup>3</sup> exposure groups.

**Result**

: No treatment-related changes were observed in either species in clinical signs, body weight, clinical chemistry or hematology except four male rats in the high dose group that had lesions on the skin in the scrotal area. This was attributed possibly to an interaction between abrasions of the skin against the floors of the cages and the exposure to the high concentrations of LCCN.

Organ weights were unaffected in either species, except for uterus weights. Uterine weights in the rats were less than untreated controls for all exposed groups, but not less than the sham controls. The actual weights (g) shown below, were not considered to be related to LCCN because they were not dose-related, and there was no difference between the sham and untreated controls. Additionally, no similar effect was observed in the mice.

Untreated controls	0.69 +/- 0.17
Sham controls	0.62 +/- 0.07
530 mg/m <sup>3</sup>	0.55 +/- 0.12
2060 mg/m <sup>3</sup>	0.52 +/- 0.05
7690 mg/m <sup>3</sup>	0.54 +/- 0.09

No treatment-related abnormalities were observed in any of the organs examined microscopically. The incidence of the occurrence of hyaline droplets in dilated tubules was similar in the controls and the high dose males and was not considered relevant.

The number of sperm per gram of cauda epididymis was significantly lower in the 7690 mg/m<sup>3</sup> group than in the sham controls but not the untreated controls. The number of epididymal sperm was not significantly affected by exposure. In addition, the number of testicular spermatids and the percentage of abnormal sperm in the cauda epididymis were not affected by exposure to 7690 mg/m<sup>3</sup> compared to either control group.

**Test substance**

: LCCN CAS # 64741-55-5

Vapors of LCCN were generated in a glass countercurrent generator (one for each concentration).

As liquid LCCN flowed down the coil, nitrogen passed upwards and carried off vapors of the more volatile components. Mainstream air was used to dilute the vapor to the required concentration.

Vapor concentration was monitored at approximately hourly intervals during each exposure period.

Concentrations (Target and actual) are shown below.

Target (mg/m <sup>3</sup> )	Actual (mg/m <sup>3</sup> )
500	530 +/- 90
2000	2060 +/- 250
8000	7690 +/- 730

In addition the composition of neat LCCN (liquid), its static headspace and the inhalation chambers was assessed. The results shown below confirm that the animals had been exposed to the lighter components of LCCN .

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Component	% in		Exposure Chamber
	Liquid LCCN	Static Headspace	
Total C <sub>4</sub> /C <sub>5</sub> nonaromatics	18.6	76.5	38.2
n-Butane	0.2	5.5	
Isobutane	-	1.0	0.2
Butenes	1.7	11.4	
n-Pentane	1.4	3.3	7.0
Pentenes	3.8	13.1	
Methyl butene	5.2	11.9	
Total C <sub>6</sub> nonaromatics	16.2	14	
n-Hexane	1.0	0.9	
2,3-Dimethylbutane	1.3	1.0	
2-Methylpentane	4.1	6.0	12.2
3-Methylpentane	2.5	2.7	7.5
Methylpentene	1.7	0.9	
Hexenes	2.1	1.1	
Methylcyclopentane	2.3	1.8	
4-Methylcyclopentane	0.6	0.5	
Total C <sub>7</sub> nonaromatics	10.2	1.9	
Total C <sub>8</sub> nonaromatics	8.0		
n-Octane	0.3		
2,2,4-Trimethylpentane	0.8	0.4	2.7
Total Aromatics	24.0		
Benzene	0.1	0.6	2.3
Toluene	4.6	0.8	4.7
Ethylbenzene	1.5	0.1	
Xylenes	7.6	0.5	2.3

### Reliability

: (2) Valid with restrictions  
The data presented in the publication are more or less limited to those showing effects. Where no effects are reported, actual data are not shown. Nevertheless, the study is sound and helpful in assessing the effects of LCCN light ends on this biological endpoint.

26.10.2001

(5)

### 5.5 GENETIC TOXICITY 'IN VITRO'

**Remark** : No data  
26.10.2001

### 5.6 GENETIC TOXICITY 'IN VIVO'

**Remark** : No data  
26.10.2001

### 5.7 CARCINOGENITY

**Remark** : No data  
26.10.2001

## 5.8 TOXICITY TO REPRODUCTION

<b>Type</b>	:	Reproductive/developmental toxicity screening study, (OECD 421)
<b>Species</b>	:	Rat
<b>Sex</b>	:	Male/female
<b>Strain</b>	:	Sprague-Dawley
<b>Route of admin.</b>	:	Inhalation
<b>Frequency of treatment</b>	:	6 hours/day, 7 days/week
<b>Premating exposure period</b>		
<b>Male</b>	:	2 weeks
<b>Female</b>	:	2 weeks
<b>Duration of test</b>	:	
<b>Doses</b>	:	Target: 750, 2500 & 7500 ppm. Actual: 752, 2512 & 7518 ppm
<b>Control group</b>	:	Yes
<b>NOAEL Parental</b>	:	2500 ppm
<b>other: NOAEL for reproductive performance and developmental toxicity</b>	:	7500 ppm
<b>Method</b>	:	OECD combined repeated dose and reproductive/developmental toxicity screening test
<b>Year</b>	:	1999
<b>GLP</b>	:	Yes
<b>Test substance</b>	:	TS: LCCN-D
<b>Method</b>	:	Groups of 10 rats of each sex were exposed to 750, 2500 or 7500 ppm. LCRN-D for 6 hours /day, seven days/week. A group of 10 rats of each sex served as sham treated controls. Parental females were exposed for 14 consecutive days prior to mating, throughout mating and days 0-19 of gestation. Dams and their litters were sacrificed on post-partum day 4. Unmated females and parental males were exposed to the test material for 14 days prior to mating, throughout mating and for 23 additional days following completion of the mating period. These animals were sacrificed shortly after the last litters were delivered reached post-partum day 4.
		<b>Mating</b> Within each group one male was co-housed with the same female until evidence of mating was observed (presence of sperm in vaginal smear or copulatory plug). The day of mating was designated day 0 of gestation. Following mating, the females were housed individually and continued their exposures to test material until day 19 of gestation. Females not showing evidence of mating following a 14-day mating period continued their exposures. If such a female showed signs of being pregnant, it was removed from the exposure regimen and observed for parturition.
		<b>Observations</b> All parental animals were regularly observed for mortality and gross pharmacologic signs. A physical examination, including palpation for tissue masses was carried out daily 30 mins after removal from the exposure chambers. Body weights and food consumption were measured throughout the study. From day 20 of gestation, females (pregnant and non-pregnant) were observed for signs of parturition. As soon as possible after delivery, litters were observed for the number of live and dead pups and for any

abnormalities. Litters were also observed twice daily for unusual findings and dead pups. On days 0 and 4 of lactation, the pups were counted, weighed and their sex was determined by external observation.

Pathology

Males were killed as a group shortly after the last litters delivered had reached day 4 of lactation. Females with litters that reached day 4 of lactation were killed the next day or shortly thereafter. Unmated females and those that did not deliver were killed 23 days after completion of the mating period. At post mortem, a complete macroscopic examination was carried out on all adult animals. The following organs were weighed and organ/body weight ratios were calculated: adrenals, brain, heart, kidneys, liver, lung, spleen, epididymes, testes and thymus. Post mortem examination of females included a count of uterine implantation scars when present.

Pups were sacrificed on day 4 of lactation and underwent a complete macroscopic examination and a determination of sex by internal examination. All pups were preserved with viscera intact. Pups found dead at birth and that died prior to day 4 of lactation also underwent a gross external and internal examination. Dead pups were not eviscerated, but were preserved intact.

27 tissues were preserved from all adult animals in all dose groups. Ovaries, testes, epididymes, nose with nasal turbinates, and any grossly observed abnormalities were processed and sections examined histologically for all males and female parental animals in the control and highest dose group. Four sections were prepared and examined microscopically of the skull containing the nasal turbinates. These were

- area between upper incisor and incisive papilla
- area between incisive papilla and first palatal ridge
- area between second palatal ridge and first upper molar
- area between first upper molar and nasopharynx.

Result

: All animals survived to scheduled sacrifice. Red staining on the snout was seen with increasing frequency in the mid and high dose animals of both sexes throughout the study. Microscopic examination of the nasal turbinates of the sham-exposed and high dose animals did not reveal any significant changes.

Although all treated groups gained slightly less weight than the sham treated controls, the differences were not statistically significant. Food consumption was comparable in all groups. Apart from those listed below, absolute and relative organ weights were unaffected by treatment.

- High dose Males
- Absolute kidney weight increased (18%)
- Relative kidney weights increased (24%)
- Relative liver weights increased (15%)

- High dose females.
- Absolute spleen weights increased by (19%)
- Relative spleen weights increased by (19%)

At necropsy, no organs appeared abnormal.

Microscopic examination of kidneys from one high dose male with a dilated

## 5. Toxicity

Id O. Naphthas  
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renal pelvis at necropsy revealed hyaline droplet formation and tubular dilatation of tubules in the cortico-medullary junction. This finding is consistent with male-rat-specific light hydrocarbon nephropathy. No test-related microscopic changes were observed in the testes or epididymes of adult male rats or ovaries of adult female rats in the high dose group.

### Reproductive/fertility effects

All groups had a fertility index of >90% and all groups had a live birth index greater than or equal to 98%.  
Data are summarized below.

Parameter	Dose group (ppm)			
	0	750	2500	7500
Females on study	10	10	10	10
Litters with liveborn	9	8	9	10
Implantation sites	155	126	139	160
Mean	17.2	15.8	15.4	16
Pups delivered (total)	149	110	132	152
Liveborn	149	108	131	151
Live birth index (%)	100	98	99	99
Pups dying				
Day 0	0	2	1	1
Days 1-4	4	2	2	1
Pups surviving 4 days	145	106	129	150
Viability index (%)	97	98	99	99
pup sex distribution				
Day 0 M/F (ratio)	72/77	50/58	65/66	87/64
Day 4 M/F (ratio)	72/73	49/57	65/64	87/63
Pup weight/litter (g)				
Day 0	6.3	6.6	6.4	6.4
Day 4	9.9	10.8	10.1	10.3

External and internal examination of pups sacrificed on day 4 of lactation were unremarkable.

Reliability  
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: (1) Valid without restriction

(13)

### 5.9 DEVELOPMENTAL TOXICITY/TERATOGENICITY

**Species** : Rat  
**Sex** : Female  
**Strain** : Sprague-Dawley  
**Route of admin.** : Inhalation  
**Exposure period** : During days 0 to 19 of gestation inclusive.  
**Frequency of treatment** : 6 hours each day  
**Doses** : Target: 2000 & 8000 mg/m<sup>3</sup>. Actual: 2150 & 7660 mg/m<sup>3</sup>  
**Control group** : Yes  
**Year** : 1996  
**GLP** : No data  
**Test substance** : LCCN CAS # 64741-55-5  
**Method** : Four groups of 15 presumed-pregnant female rats were assigned to the following groups: Untreated controls, sham-treated controls, 2000 and 8000 mg/m<sup>3</sup> test material.  
Exposures were for 6 hours each day on days 0 to 19 of gestation.  
All animals were observed daily and body weights were recorded on days 0, 6, 13 and 20 of gestation.  
On day 20, each female was sacrificed and all organs were examined

**Result**

grossly. Serum samples were analyzed for a variety of parameters, including serum iron and lactic dehydrogenase. The number of corporea lutea per ovary and the gravid uterine weights were recorded. Uterine contents were examined and the numbers of implantation sites, early resorptions and live and dead fetuses recorded. Each fetus was identified for its sex, was weighed and the crown-rump distance was measured. Each fetus was examined for external anomalies. Half the fetuses were fixed in Bouin's solution and examined for visceral anomalies and the remaining fetuses were prepared for examination for skeletal anomalies.

: There were no treatment-related clinical abnormalities or differences in body weight. Results of the reproductive parameters are listed below.

Parameter	No treat	LCCN		
		Control Sham treat	2150	7660
Females mated	15	15	15	15
Females pregnant	14	13	14	15
Corpora lutea	18	18	16	18
Implantation sites	16	16	14	16
Primplantation loss (&)	10	12	14	8
Viable fetuses/litter	15	14	14	15
Resorptions	0.7	0.6	0.8	1.7 *a
Resorptions (%)	4.6	3.9	4.7	10.4 *a
Dams with resorptions	9	5	8	13 *b

\*a Significant difference from untreated and sham treated controls

\*b Significant difference from sham treated controls

It is clear that with the exception of resorptions, no other parameter was affected by exposure.

During the external examination of fetuses, a sham treated animal had gastroschisis and one fetus from the 2150 mg/m<sup>3</sup> group had a tail that was short and filamentous.

Fetal body weights and crown-rump lengths were unaffected by treatment.

No visceral abnormalities were observed.

There was an increased number of skeletal variations in animals housed in the exposure chambers (exposed and sham treated controls) when compared to the untreated controls.

The authors concluded that these alterations were not related to LCCN since they occurred at the same incidence in the sham treated controls as well.

The findings are tabulated below.

## 5. Toxicity

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Parameter	No treat	Control Sham treat	LCCN	
			2150	7660
Caudal vertebrae: transverse process incompletely ossified	18(16)	42(40)	41(40)	45(39)
Sacral vertebrae transverse process incompletely ossified	7(6)	23(22)	17(17)	28(24)
Incompletely ossified sternebrae	83(75)	80(76)	91(89)	101(88)

**Test substance** : A description of atmosphere generation is given in a publication by the same authors in section 5.4.

**Reliability** : Actual concentrations in this study were:  
26.10.2001 2150 +/- 260 and 7660 +/- 570 mg/m<sup>3</sup>.

: (1) valid without restriction

(5)

## 6. References

Id O. Naphthas  
Date 10.12.2001

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## 6. References

**Id** O. Naphthas  
**Date** 10.12.2001

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**ROBUST SUMMARY  
OF INFORMATION ON**

**Substance Group:** **AROMATIC NAPHTHAS**

**Summary prepared by:** American Petroleum Institute

**Creation date:** 1 MARCH 2001

**Printing date:** 19 NOVEMBER 2001

**Date of last Update:** 10 DECEMBER 2001

**Number of Pages:** 37

NB. Reliability of data included in this summary has been assessed using the approach described by Klimisch, et al.

Klimisch, H. J., Andreae, M. and Tillman, U, (1997)

A systematic approach for evaluating the quality of experimental toxicological and ecotoxicological data.  
Regulatory Toxicology and Pharmacology 25, 1-5.

# 1. General Information

Id A. Naphthas  
Date 10.12.2001

## 1.1 GENERAL SUBSTANCE INFORMATION

**Substance type** : petroleum product  
**Physical status** : liquid  
**Remark** : Aromatic naphtha streams are obtained from the catalytic reforming of mainly n-alkane and cycloparaffinic feedstocks into aromatic and branched chain hydrocarbons. They are mainly in the range C5 to C12 and boil in the range of approximately 35 to 230°C.

Several materials have been employed in tests for toxicological properties of this category of products.

Details of the various samples are as follows:

API 83-05 is a sample of a catalytically reformed naphtha.  
LCRN is a sample of Light Catalytically Reformed Naphtha  
LCRN-D is a 154 °F distillation fraction derived from LCRN.  
Characterization of these samples is shown in the following table.

Parameter	API 83-05	Sample LCRN	LCRN-D
Gravity (°API)44			
Sulfur (wt %)	<0.01		
Nitrogen (ppm)	<2		
RVP (psia)	3		
IBP (°F)	132		
FBP (°F)	389		
Olefins (% by MS)	1.5	0.96	1.37
Paraffins (% by MS)	31	63.76	88.3
Naphthenes (% by MS)	5	2.28	1.24
Aromatics (% by MS)	62.5	33	9.09
Benzene (vol %)	2.2	6.93	4.65
Carbon No. (vol%)			
4		1.02	3.6
5		23.34	59.11
6		25.76	25.18
7		45.27	11.65
8		4.56	0.46
9		0.05	0

19.11.2001

## 2. Physico-Chemical Data

Id A. Naphthas  
Date 10.12.2001

### 2.1 MELTING POINT

Not relevant

### 2.2 BOILING POINT

35 to 230 °C see General substance information

### 2.5 PARTITION COEFFICIENT

**Log pow** : 2.13 - 4.5 at 25° C  
**Method** : Calculated by LOGKOWWIN ver. 1.65.  
**Year** : 2000  
**GLP** : No  
**Test substance** : Light Catalytically Reformed Naphtha  
**Remark** : Log P values represent the spread of calculated and/or measured values for C5 to C9 hydrocarbon components found in LCRN, CAS No 64741-63-5. Detailed hydrocarbon analysis performed by Chevron Research was used to identify the components of this specific LCRN sample. Calculation based on an atom/fragment contribution method of W. Meylan and P. Howard. Calculated SAR result for surrogate structure contained in program database (smilecas.dat).

**Reliability** : (2) valid with restrictions  
26.06.2001

(19)

**Log pow** : 2.13 - 4.76 at 25° C  
**Method** : Calculated by LOGKOWWIN ver. 1.65.  
**Year** : 2000  
**GLP** : No  
**Test substance** : Full -Range Catalytically Reformed Naphtha (FRCRN)- CAS No. 68955-35-1; API sample 83-05  
**Remark** : Log P values represent the spread of calculated and/or measured values for C5 to C9 hydrocarbon components found in FRCRN, CAS No 68955-35-1. Detailed hydrocarbon analysis was used to identify the components of this FRCRN (63% aromatics) sample. Calculated SAR result for surrogate structures contained in program database (smilecas.dat). Calculation based on an atom/fragment contribution method of W. Meylan and P. Howard.

**Reliability** : (2) valid with restrictions  
11.11.2001

(19)

## 2.6.1 WATER SOLUBILITY

<b>Method</b>	:	Preparation of Water Soluble Fraction
<b>Year</b>	:	1995
<b>GLP</b>	:	Yes
<b>Test substance</b>	:	Light Catalytically Reformed Naphtha
<b>Method</b>	:	Water Accommodated Fractions (WAFs) of LCRN were prepared at 50 mg/l loading in freshwater and saltwater and equilibrated for 48 hours in tightly closed systems with minimal headspace.
<b>Result</b>	:	Gas chromatographic analysis of selected components freshwater and saltwater solubilities of 13.7 and 14.0 ppm respectively. Measured test concentrations of the light catalytically reformed naphtha were based on the total combined concentrations of pentane, 2-methyl-pentane, benzene, toluene, ethylbenzene, ortho, meta and para-xylene, which represent more than 50% composition of the test substance. Concentrations for these components reached equilibrium by 24 hours.
<b>Conclusion</b>	:	Individual components of complex petroleum substances have specific and differing solubilities. Calculated and measured water solubilities for LCRN components range from approximately 3 to 2000 mg/l. At any particular loading rate, the resulting aqueous concentration of each chemical constituent is a function of the relative volume of the two phases (aqueous and the petroleum mixture), the partition coefficient between the phases, the amount of component present and the maximum water solubility of each component. Initially as the petroleum mixture is added in amounts below the solubility limit of the least soluble component the aqueous concentration increases proportionally until the least soluble component reaches a saturation concentration, and only the more soluble components continue to dissolve, resulting in a two phase system. Further addition of the petroleum mixture results in an aqueous concentration that is a non-linear function of the amount added.
<b>Reliability</b> 19.11.2001	:	(2) valid with restrictions  (1) (11) (13) (15) (16)
<b>Method</b>	:	Preparation of Water Soluble Fraction
<b>Year</b>	:	1995
<b>GLP</b>	:	yes
<b>Test substance</b>	:	Full -Range Catalytically Reformed Naphtha (FRCRN)- CAS No. 68955-35-1; API sample 83-05
<b>Method</b>	:	Water Accommodated Fractions (WAFs) of CONCAWE Reformate light naphtha (LCRN), CAS no. 64741-63-5 (CONCAWE sample ID W94/812) were prepared at 100 mg/l loading in freshwater and equilibrated for 48 hours in tightly closed systems with minimal headspace.
<b>Remark</b>	:	Detailed hydrocarbon analysis was used to identify the components of this CONCAWE Light Cracked Naphtha (63% aromatics) sample. The analysis indicated that the composition of the CONCAWE LCRN sample was essentially identical to the composition of API 83-05 FRCRN sample. Therefore the water solubility information for the CONCAWE LCRN sample is applicable to the FRCRN sample.
<b>Result</b>	:	Gas chromatographic analysis of LCRN components benzene, toluene, ethylbenzene, ortho, meta and para-xylene in WAFs indicated freshwater solubility of 6.3 ppm. Concentrations for these components reached equilibrium by 48 hours.

## 2. Physico-Chemical Data

**Id** A. Naphthas  
**Date** 10.12.2001

### **Conclusion**

: Individual components of complex petroleum substances have specific and differing solubilities. Calculated and measured water solubilities for FRCRN components range from approximately 3 to 2000 mg/l. At any particular loading rate, the resulting aqueous concentration of each chemical constituent is a function of the relative volume of the two phases (aqueous and the petroleum mixture), the partition coefficient between the phases, the amount of component present and the maximum water solubility of each component. Initially as the petroleum mixture is added in amounts below the solubility limit of the least soluble component the aqueous concentration increases proportionally until the least soluble component reaches a saturation concentration, and only the more soluble components continue to dissolve, resulting in a two phase system. Further addition of the petroleum mixture results in an aqueous concentration that is a non-linear function of the amount added.

### **Reliability** 19.11.2001

: (2) valid with restrictions

(11) (12) (13) (15) (16)



#### 3.1.2 STABILITY IN WATER

**Test substance** : Light Catalytically Reformed Naphtha  
**Conclusion** : Hydrolysis unlikely  
**Reliability** : (1) valid without restriction  
 26.06.2001 (17)

**Test substance** : Full -Range Catalytically Reformed Naphtha (FRCRN)  
 CAS No. 68955-35-1; API sample 83-05  
**Conclusion** : Hydrolysis unlikely  
**Reliability** : (1) valid without restriction  
 11.11.2001 (17)

#### 3.3.1 TRANSPORT BETWEEN ENVIRONMENTAL COMPARTMENTS

**Type** : Calculated according to Mackay Level 1  
**Media** : Soil, air, water, suspended sediment and sediment  
**Year** : 2000  
**Remark** : Model based on chemical fugacity. Physical properties input are those calculated by the EPIWIN Estimation 3.04 program and included in this summary. Values represent the spread of calculated values for C5 to C8 hydrocarbon components found in LCRN, CAS No 64741-63-5. Detailed hydrocarbon analysis performed by Chevron Research was used to identify the components of this specific LCRN sample.

The majority of LCRN components will partition rapidly to air, where these hydrocarbons will be rapidly oxidized by OH radicals.

**Result** : 

Medium	% distribution
Air	97 to 99.98
Soil	0.01 to 0.8
Water	0.01 to 2.7
Sediment	0.00
Suspended sediment	0.00

**Conclusion** : The constituents of this complex petroleum mixture are expected to partition primarily to air.  
**Reliability** : (2) valid with restrictions  
 05.10.2001 (18)

**Type** : Calculated according to Mackay Level I  
**Media** : Soil, air, water, suspended sediment, and sediment  
**Year** : 2000  
**Remark** : Model based on chemical fugacity. Physical properties input are those calculated by the EPIWIN Estimation 3.04 program and included in this summary. Values represent the spread of calculated values for C5 to C9 hydrocarbon components found in FRCRN, CAS No 68955-35-1. Detailed hydrocarbon analysis was used to identify the components of this specific FRCRN (63% aromatics) sample.  
 The majority of FRCRN components will partition rapidly to air, where these hydrocarbons will be rapidly oxidized by OH radicals.

### 3. Environmental Fate and Pathways

Id A. Naphthas  
Date 10.12.2001

**Result** : Medium % distribution  
Air 96.5 to 99.98  
Soil 0.01 to 1.83  
Water 0.01 to 2.7  
Sediment <0.001 to 0.03  
Suspended Sediment <0.001

**Test substance** : Full -Range Catalytically Reformed Naphtha (FRCRN)  
CAS No. 68955-35-1; API sample 83-05

**Conclusion** : The constituents of this complex petroleum mixture are expected to partition primarily to air.

**Reliability** : (2) valid with restrictions  
11.11.2001

(18)

#### 3.5 BIODEGRADATION

**Type** : aerobic  
**Inoculum** : mixed, adapted inoculum of domestic activated sludge and soil  
**Contact time** : 56 day  
**Result** : inherently biodegradable  
**Year** : 1999  
**GLP** : yes  
**Test substance** : Light Catalytically Reformed Naphtha  
**Method** : CONCAWE. Test method for determining the inherent aerobic biodegradability of oil products. 1996/1997, and modification of ISO/DIS 14593

: Type (test type): Water quality-Evaluation of ultimate aerobic biodegradability of organic compounds in aqueous medium-Method by analysis of inorganic carbon in sealed vessels (CO<sub>2</sub> headspace test)

**Result** : Test material was inherently biodegradable since it achieved >20% biodegradability based on CO<sub>2</sub> production. By day 28 approximately 96% of the test material was degraded, then essentially reached a plateau in degradation rate until day 56. The test was considered valid according to CONCAWE criteria, as >60% biodegradation of positive control (63% actual) was observed by day 14, and total blank CO<sub>2</sub> production at termination was less than 15% of the organic carbon added as test substance. Temperature ranged from 18 to 21 °C, which deviated from the protocol value of 22 ±2°. This deviation was not expected to have affected the outcome of this study.

Test Day	% Degradation (sd)	
	Hexadecane	Test Material
3	13.93 (1.85)	30.85 (3.85)
7	34.40 (4.54)	53.71 (3.52)
14	63.17 (0.94)	77.25 (3.65)
21	77.26 (6.52)	87.17 (8.87)
28	90.35 (7.14)	96.17(5.26)
35	85.13 (n=1)	107.9 (n=1)
42	85.21 (n=1)	96.95 (6.37)
49	96.93 (8.94)	92.02 (n=1)
56	94.69 (4.10)	84.92 (0.51)

**Test condition** : Mixed inoculum prepared from soil and activated sludge was incubated with test substance or hexadecane (positive control) during a two-week adaptation period. Triplicate test systems were incubated for both the test substance and hexadecane fed inoculum. Two additional, similar test substances were concurrently incubated in separate 160 ml test systems using the same inoculum and acclimation

procedure. Duplicate blank control test systems were prepared which consisted of the mixed inocula in mineral medium but no test or positive control substance. Test medium consisted of glass-distilled water and mineral salts (phosphate buffer, ferric chloride, magnesium sulfate, calcium chloride) prepared as described in ISO method.

Acclimation procedure-Activated sludge from aeration basin of Wareham Wastewater Treatment Plant (Mass., U.S.A.) was sieved through 2 mm and centrifuged at 1000 rpm for 10 minutes. After removal of supernatant the concentrated solids were diluted to 5 mg/ml suspended solids with reagent grade water. Soil was collected from a site located in a mixed hardwood and pine forest (Mass., U.S.A.). Site of sampling was cleared of debris and approximately 500 g of soil was obtained at a depth between 5-10cm from the soil surface. Soil was air-dried, sieved through a 2 mm sieve, and analyzed for moisture content (38%).

Test vessels (160 ml serum bottles) were filled with 103 ml of mineral medium containing 50 mg/l of yeast extract and 50 mg/l (dry weight) washed activated sludge, then approximately 0.16g of sieved soil (0.1 g dry wt) was added to each bottle. Test or reference substances were added directly to test systems using a 10 microliter Hamilton gas tight syringe. The volume required to achieve the specified mg carbon/l concentrations were calculated based on % carbon and specific gravity of the respective substance. The test substance % carbon (0.8856) and specific gravity (0.7325 mg/ $\mu$ l) information was supplied by the Sponsor. Hexadecane % carbon (0.8496) was calculated from the empirical formula and specific gravity (0.7749 mg/ $\mu$ l) was obtained from Verschuere (1983). Addition of respective substance was performed on an incremental basis to the appropriate vessels as follows: 4, 8 and 8 mg C/l were added on days 0, 7 and 11, respectively. Test vessels were sealed with butyl rubber septa/aluminum crimp caps and incubated at 22 ° ( $\pm$  2°C) in the dark.

#### Biodegradation by CO<sub>2</sub> determination

##### Test initiation and procedure

On day 14 of the acclimation phase, all test system inoculum from blanks, positive control, and each of the three test substances was combined and filtered through glass wool, and aerated prior to use. The aerated mixed inoculum was then added to mineral medium to achieve 10% concentration based on total volume (100 ml inoculum/l). Test vessels (160 ml serum bottles) were filled with 103 ml of inoculated mineral medium. Respective test systems were dosed with either test substance or hexadecane as described for the acclimation procedure to achieve 20 mg carbon/l concentration. Duplicate test systems for each test substance, positive control and blank treatments were prepared for sacrifice at weekly sampling intervals for subsequent CO<sub>2</sub> analysis. After test system preparation, all vessels were placed in a walk-in chamber and incubated in the dark at 22 ° ( $\pm$  2 ° C).

On days 3, 7, 14, 21, 28, 35, 42, 49 and 56, 1ml of conc H<sub>3</sub>PO<sub>4</sub> was injected through the septum of each sacrificed test vessel. The acidified samples were shaken for 1 hr at 200 ppm, then analyzed for CO<sub>2</sub> using gas chromatography-thermal conductivity detection. Quantitation

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of inorganic mg C/l evolved was determined by linear regression analysis based on response factors for sodium carbonate standards spanning 1-30 mg carbon/l concentrations.  
: (1) valid without restriction

(24)

## 4.1 ACUTE/PROLONGED TOXICITY TO FISH

<b>Type</b>	: 96 Hr Static Acute Toxicity Test w/ Daily Renewal
<b>Species</b>	: Pimephales promelas (Fish, fresh water)
<b>Exposure period</b>	: 96 hour(s)
<b>Unit</b>	: Mg/l
<b>Analytical monitoring Method</b>	: yes : No specific guideline was described as being used to conduct the test, however report references 1991 ASTM method E729-88a and 1982 EPA Support Documents for Environmental Testing: EPA 560/6-82-002.
<b>Year</b>	: 1998
<b>GLP</b>	: yes
<b>Test substance</b>	: Light Catalytically Reformed Naphtha
<b>Method</b>	: Statistical Method: (FT - ME) LL <sub>50</sub> and LC <sub>50</sub> calculated using binomial probability analysis. ASTM Special Technical Publication 634. 1977, pp 65-84.
<b>Result</b>	: Mortality (no. of deaths/treatment) at 96 hrs: 1, 0, 1, 0, 1 and 20 in 0, 3.1, 6.3, 13, 25 and 50 mg/l treatments. Abnormal behavior (surfacing, erratic swimming) was observed at 96 hrs for 3 organisms in 13 mg/l and 7 fish in 25 mg/l treatments. 96-hr LL <sub>50</sub> = 34 mg/l, 25-50 mg/l w/ 95% C.I. (as nominal loading rate) 96-hr LC <sub>50</sub> = 11 mg/l, 8.2-17.2 mg/l mg/l w/ 95% C.I. (measured concentrations) 96-hr NOEL = 3.1 mg/l (nominal); 96-hr NOEC = 1.03 mg/l (measured) based on lack of mortality and abnormal effects for these treatments.
<b>Test condition</b>	: Test solutions were prepared as water accommodated fractions (WAF). Control and dilution water was prepared by blending naturally hard well water with water that had been demineralized by reverse osmosis. Nominal loading rates of 0, 3.1, 6.3, 13, 25 and 50 mg/l were used to prepare test solutions.  WAFS were prepared for each test concentration by mixing the appropriate mass of substance in 9.4 liters of water for 24 hr in 9 liter glass bottles. The bottles were filled to neck height with water to minimize volatility. A measured amount of test substance was added into each bottle, and the bottles were capped tightly with a positive pressure siphoning apparatus. The siphoning apparatus consisted of a teflon lined neoprene stopper housing two teflon tubes. One tube extended to the bottom of the bottle for removal of the WAF solution, the other tube ended above the WAF surface, and was used to control air pressure during siphoning. During WAF preparation, parafilm was used to seal the external joint between the neoprene stopper and glass bottle, and the bottles were covered with aluminum foil. After stirring for 24 hrs using 25% or less vortex, the contents of the WAF solution bottles were allowed to settle for approximately 45 minutes to two hours, then siphoned by the positive pressure apparatus port and used for testing. Samples were also analyzed by Purge & trap/GC-FID for concentrations of the following: pentane, 2-methyl-pentane, benzene, toluene, ethylbenzene, ortho, meta and para-xylene, which represent more than 50% composition of the test substance. Measured test concentrations of the light catalytically reformed naphtha were based on the total combined concentrations of all analytes.

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Fish were hatched and raised from ABC Laboratories' in-house culture, and were acclimated prior to experimentation for a minimum of 14 days on a 16/8hr light/dark cycle. Test vessels were 3.8-liter glass containers with teflon-lined caps. Fish were acclimated to the test water and temperature approximately 72 hr before the test, and were not fed during this 72 hr period. Two replicates per treatment and 10 organisms per replicate were tested for each treatment and the control. Exposure containers were filled with no headspace and tightly sealed to prevent volatilization. Test solution renewal was performed daily by removal of most of the test solution (leaving approximately one liter of solution to avoid stressing test organisms) and replacing it with fresh WAF solution prepared at least 24 hrs prior to use.

Water temperature was 21-22 °C. Test photoperiod was 16 hrs. light and 8 hr dark. Dissolved oxygen measurements were between 6.0 and 8.5, pH values between 7.7 and 8.5. Hardness values ranged from 138 to 148 mg/l; alkalinity values ranged from 150 to 158 mg/l and conductivity values ranged from 299 to 313 microsiemens.

<b>Reliability</b>	:	(2) valid with restrictions Measured concentrations may not represent 100% of components, remaining hydrocarbon components in WAFs may be equally toxic and should have been quantitated to determine total measured concentrations.	
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<b>Type</b>	:	Predicted value for fish toxicity	
<b>Exposure period</b>	:	96 hour(s)	
<b>Unit</b>	:	mg/l	
<b>LL50</b>	:	2.09	
<b>Method</b>	:	Predicted	
<b>Method</b>	:	Calculated based on hydrocarbon block principle. In this procedure, the dissolved concentrations of individual hydrocarbons from a petroleum substance are estimated for a given loading rate and then normalized by their acute toxicity to yield Toxic Units (TU) which can be summed to predict the toxicity of the parent material (see below).	
<b>Result</b>	:	Estimated 96 hour(s) fish acute toxicity LL <sub>50</sub> : 2.09 mg/l	
<b>Reliability</b>	:	(2) valid with restrictions	
11.11.2001			(21)

## 4.2 ACUTE TOXICITY TO AQUATIC INVERTEBRATES

<b>Type</b>	:	48 Hr Static Acute Toxicity Test with 24Hr Renewal in Closed Systems
<b>Species</b>	:	Daphnia magna (Crustacea)
<b>Exposure period</b>	:	48 hour(s)
<b>Unit</b>	:	Mg/l
<b>Analytical monitoring Method</b>	:	Yes Procedure patterned after:1991 ASTM method E729-88a and 1985 USEPA TSCA Test Guidelines: Daphnid Acute Toxicity Test. Fed. Reg., vol. 50 (No. 188) Sept 27, 1985, 797.1300. Statistical Method: (FT - ME) EL 50 and EC50 calculated using binomial probability analysis. ASTM Special Technical Publication 634. 1977, pp 65-84.
<b>Year</b>	:	1998
<b>GLP</b>	:	Yes
<b>Test substance</b>	:	Light Catalytically Reformed Naphtha
<b>Result</b>	:	Immobility (no. of organisms) at 48 hrs: 0, 0, 0, 15, 20 and 20 for 0, 3.0, 6.0, 12, 24 and 48 mg/l treatments. At the 6 and 12mg/l nominal treatments, 20 and 5 organisms were observed at the bottom of the test chambers, respectively.  48-hr EL <sub>50</sub> = 10 mg/l based upon nominal loading rate (95% C.I. 6 to 12 mg/l);  48 hr EC <sub>50</sub> was 2.6 mg/l (95% C.I. 1.06 to 3.6 mg/l); based on total measured concentrations.  48-hr NOEL = 3 mg/l based upon nominal loading rate.  48 hr NOEC was 0.465 ppm based on total measured concentrations.
<b>Test condition</b>	:	Test solutions were prepared as water accommodated fractions (WAF). Control and dilution water were a blend of aged well water and reverse osmosis well water.  WAFS were prepared for each test concentration by mixing the appropriate mass of substance in 2.4 liters of water for 24 hr in aluminum foil covered 2.5 liter aspirator bottles fitted w/ a port near the bottle bottom. The bottles were filled to neck height with water to minimize volatility. A measured amount of test substance was pipetted below the water surface. After stirring for 24 hrs using 25% or less vortex, the contents of the aspirator bottles were allowed to settle for approximately one hour, then drained from the port and used for testing. Samples were also analyzed by purge & trap/GC-FID for concentrations of the following: pentane, 2-methyl-pentane, benzene, toluene, ethylbenzene, ortho, meta and para-xylene, which represent more than 50% composition of the test substance. Measured test concentrations of the light catalytically reformed naphtha were based on the total combined concentrations of all analytes.  Range finding toxicity studies were conducted at 0.5, 1.0, 10 and 100 mg/l loading, using WAFS which were divided into duplicate aliquots and tested. Definitive toxicity studies were conducted at 3.0, 6.0, 12, 24 and 48 mg/l loading, using WAFS which were divided into duplicate aliquots and tested.  Test vessels were teflon cap-sealed 273 ml glass jars with 10 daphnids per jar and were completely filled with test

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solution.

During the study test system solutions: dissolved oxygen concentration range: 8.0 to 8.5; pH ranged from 8.2 to 8.4; temperature was 20 to 21 °C; hardness (mg/l) ranged from 146 - 152; alkalinity (mg/l) was 158-168 and conductivity (umhos) values were 312 - 317.

Daphnia magna, were supplied by testing laboratory; age < 24 hours old; obtained from culture maintained in-house since January 1998.

**Reliability** : (2) valid with restrictions  
Measured concentrations may not represent 100% of components; remaining hydrocarbon components in WAFs may be equally toxic and should have been quantitated to determine total measured concentrations.

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(3)

**Type** : Predicted value for invertebrate 48 hour acute toxicity  
**Exposure period** : 48 hour(s)  
**Unit** : mg/l  
**EL50** : 0.9  
**Method** : Calculated based on hydrocarbon block principle. In this procedure, the dissolved concentrations of individual hydrocarbons from a petroleum substance are estimated for a given loading rate and then normalized by their acute toxicity to yield Toxic Units (TU) which can be summed to predict the toxicity of the parent material (see below).

**Result** : Estimated 48 hour(s) Daphnid acute toxicity EL<sub>50</sub>: 0.9 mg/l.  
**Reliability** : (2) valid with restrictions

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### 4.3 TOXICITY TO AQUATIC PLANTS E.G. ALGAE

**Species** : Selenastrum capricornutum (Algae)  
**Exposure period** : 96 hour(s)  
**Analytical monitoring** : yes  
**Method** : No specific guideline was described as being used to conduct the test, however report references 1991 ASTM method E729-88a and 1982 EPA Support Documents for Environmental Testing: EPA 560/6-82-002.

**Year** : 1998  
**GLP** : yes  
**Test substance** : Light Catalytically Reformed Naphtha  
**Method** : Statistical Method: EL 50 and EC50 calculated using nonlinear logistics sigmoid model (SAS). All NOEL/NOEC values based on visual review and Dunnett's test for significance.

**Result** : Percent inhibition on growth determined by cell density (cells/ml):  
96 hour EL<sub>10</sub>=6.0 mg/l (3.1-8.8 mg/l CI @95% )  
96 hour EL<sub>50</sub>=8.5mg/l (7.3-9.8 mg/l CI @95% )  
96 hour EL<sub>90</sub>=12 mg/l (9.9-14 mg/l CI @95% )  
96 hour NOEL=5.0 mg/l

96 hour EC<sub>10</sub>=1.1 mg/l (0.41-1.8 mg/l CI @95% )  
96 hour EC<sub>50</sub>= 1.7mg/l (1.4-2.1 mg/l CI @95% )  
96 hour EC<sub>90</sub>=2.7 mg/l (2.1-3.4 mg/l CI @95% )  
96 hour NOEC=0.866 mg/l

Subcultures of the 10, 20 and 40 mg/l treatment cultures were placed in fresh media (no test substance) after acute

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testing for ten days indicated that growth inhibition was algistatic in all treatments. No excursions from the protocol were noted which would have affected the integrity of the study.

Concentration (mg/l)		96hr cell density (cells/ml)
Nominal	Measured	
Control	0.0147	40.5 x10 <sup>4</sup>
1.3	0.126	40.92 x10 <sup>4</sup>
2.5	0.211	42.33 x10 <sup>4</sup>
5.0	0.866	41.17 x10 <sup>4</sup>
10	2.12	11.11 x10 <sup>4</sup>
20	5.26	0.70 x10 <sup>4</sup>
40	13.3	0.04 x10 <sup>4</sup>

**Test condition** : Individual test treatment solutions were prepared as Water Accommodated Fractions (WAFs). Test material was added to 2.4 liters of sterilized AAP test media (enriched with 515 mg/l of sodium bicarbonate, 300 µg/l EDTA chelator, pH adjusted to 7.5 + 0.1 with 0.1NHCl and sterilized by 0.45 micron filtration) in 2.5 liter aspirator bottles. The mixing vessels were sealed with foil covered stoppers, covered with aluminum foil and the contents mixed on magnetic stir plates with teflon coated stir bars for approximately 24 hours at room temperature. After mixing the solutions were allowed to settle for one hour and the WAF was removed from the spout at the base of each bottle and used for testing. Test vessels were 125ml glass Erlenmeyer flasks that were completely filled (148 ml) with treatment solution and inoculated with 6 day old algae. Algal cells obtained from testing laboratory cultures grown in sterile, nutrient enriched AAP media. Original algal cultures (stock UTEX-1648) obtained from Dept of Botany, Culture Collection of Algae, University of Texas at Austin, 1997. Cell density of the algal stock culture inoculate was determined prior to study initiation with a hemacytometer cell and compound microscope. Twelve replicates were prepared for each treatment level. Nominal treatment levels were 0, 1.3, 2.5, 5.0, 10, 20 and 40 mg/l. The initial algal concentration was 1.0 x 10<sup>3</sup> cells/ml. All test replicates were placed on a shaker table at 100 oscillations per minute during the study and exposed to continuous fluorescent light, illumination range 371 to 442 ft candles. Triplicate samples were taken daily for cell counts and analytical testing. Cell densities were determined by direct microscopic examination. Samples at 0,24, 48, 72 and 96 hrs were also analyzed by Purge & trap/GC-FID for concentrations of the following: pentane, 2-methyl-pentane, benzene, toluene, ethylbenzene, ortho, meta and para-xylene, which represent more than 50% composition of the test substance. Measured test concentrations of the light catalytically reformed naphtha were based on the total combined concentrations of all analytes.

**Reliability** : Test temperature was 24-25 °C. Test solution pH ranged from 7.5 to 8.9.  
(2) valid with restrictions  
Measured concentrations may not represent 100% of components; remaining hydrocarbon components in WAFs may be equally toxic and should have been quantitated to determine total measured concentrations.

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### 5.1.1 ACUTE ORAL TOXICITY

**Type** : LD<sub>50</sub>  
**Species** : rat  
**Strain** : Sprague-Dawley  
**Sex** : male/female  
**Number of animals** : 10  
**Vehicle** : undiluted  
**Year** : 1985  
**GLP** : yes  
**Test substance** : API 83-05  
**Method** : The test material was administered undiluted, as a single oral dose to groups of 5 male 5 female rats at dose levels ranging from 3.57 to 9.8 g/kg. The dose volume varied per dosage level based on an average bulk density of 0.8 g/ml. Food had been withheld from the rats overnight prior to dosing, but they had free access to water. Following dosing, food and water were available ad-lib for a period of 14 days. The animals were observed for clinical signs of toxicity and mortality every hour for the first 6 hours after dosing and twice daily thereafter for 14 days. The rats were weighed the day before dosing and then at 7 and 14 days after dosing. All animals, whether dying during the study or surviving to termination were subjected to a gross necropsy and any abnormalities were recorded.

**Result** : Clinical signs seen during the study included: hypoactivity, diarrhea, brown-stained anal area, ataxia, prostration, red stained nose and mouth, lacrimation, dyspnea, yellow-stained abdomen, hair loss on abdomen, decreased limb tone, piloerection, hair loss on front legs, excess salivation, yellow- or reddish-brown-stained urogenital region, tremors and death. All mortality occurred within the first three days after dosing. All surviving animal had returned to normal by day 11 except for those with hair loss.

At necropsy, there were few findings in the animals that survived to termination.

Mortalities and body weights were as follows:

Dose level (g/kg)	Body weights (g)		Mortality	No with lesions at necropsy
	Initial (fasted)	Terminal		
<u>Male</u>				
5	285	336	0/5	0/5
6	315	370	1/5	1/5
6.5	324	386	1/5	1/5
7	306	-	5/5	4/5
9.8	310	-	5/5	5/5
<u>Female</u>				
3.57	212	227	0/5	1/5
4.29	199	233	0/5	0/5
5	202	235	3/5	3/5
7	210	213	3/5	2/5
9.8	208	-	5/5	5/5

Typically at non-lethal dose levels the lesions frequently observed included: presence of dark colored material in the stomach, glandular mucosa of stomach with dark red to black

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area. Additionally at the highest dose levels the urinary bladder contained a red fluid in 4 of the five males examined.

The estimated LD<sub>50</sub> values and 95% confidence limits were:

Males: 6.62 g/kg (6.2 - 7.08)

Females: 5.39 g/kg (3.23 - 6.86)

**Reliability** : (1) valid without restriction  
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(6)

### 5.1.2 ACUTE INHALATION TOXICITY

**Type** : LC<sub>50</sub>  
**Species** : rat  
**Strain** : Sprague-Dawley  
**Sex** : male/female  
**Number of animals** : 5  
**Vehicle** : air  
**Exposure time** : 4 hour(s)  
**Value** : > 5.22 mg/l  
**Year** : 1984  
**GLP** : yes  
**Test substance** : API 83-05  
**Method** : A group of 5 male and 5 female rats were exposed by whole body inhalation to API 83-05 at a nominal concentration of 5mg/l for 4 hours.  
After the 4-hour exposure the rats were observed twice daily for mortality. The animals were weighed prior to exposure and again on days 7 and 14 post exposure. On day 14 all surviving animals were killed by exsanguination following methoxyflurane anesthesia and were subjected to a full necropsy. For all animals, including those found dead during the study the lungs were removed, fixed and examined histologically.

**Result** : The exposure chamber TWA concentration was determined to be 5.22±0.14 mg/l.  
No animal died during the study and no clinical signs of systemic toxicity were observed.  
There were no significant gross observations at necropsy and no histological changes were observed in the lungs.  
The 4 hour LC<sub>50</sub> was therefore greater than 5.22 mg/l.

**Reliability** : (1) valid without restriction  
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(5)

### 5.1.3 ACUTE DERMAL TOXICITY

**Type** : LD<sub>50</sub>  
**Species** : Rat  
**Strain** : New Zealand white  
**Sex** : Male/female  
**Number of animals** : 4  
**Vehicle** : undiluted  
**Value** : > 2000 mg/kg bw  
**Year** : 1985  
**GLP** : yes  
**Test substance** : API 83-05  
**Method** : The skin of the patched area of two rabbits of each sex had been abraded whilst the other two had intact skin.  
A weighed quantity of undiluted test material (equivalent to

**Result**

a dose of 2 g/Kg) was applied to the dorsal skin of each of 4 male and 4 female rabbits. The applied dose was covered with an occlusive dressing (gauze and an impermeable covering). 24 hours after dosing, the patches were removed, the skin wiped and collars fitted to the rabbits to prevent oral intake of any residual test material. Collars were used to restrain the animals during the application period. The animals were observed for a total of 14 days post-dosing.

Body weights were recorded just prior to dosing and again seven and 14 days after dosing.

At study termination the animals were killed with carbon dioxide and a gross necropsy was performed. Any abnormalities were recorded.

: There were no mortalities during the study and there were no clinical signs of toxicity with the exception of one rabbit. This animal had soft stools, diarrhea, hypoactivity and an inflamed urogenital area during the last three days of the study.

All animals had gained weight by the end of the study.

Dermal irritation occurred during the study and this ranged from slight to severe for erythema, slight to marked for edema and slight to marked for atonia, desquamation, coriaceousness and fissuring.

At necropsy, the only findings in the males were on the treated area of the skin and were consistent with the gross observations of irritation. In the females similar skin lesions were observed and in addition, the vagina was reddened in 3 of the four animals and in one of these the trachea contained a red liquid on the inside walls and the lungs had multiple red pinpoint foci on all lobes.

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: (1) valid without restriction

(6)

**5.1.4 ACUTE TOXICITY, OTHER ROUTES**

Not relevant

**5.2.1 SKIN IRRITATION**

**Species** : rabbit  
**Concentration** : undiluted  
**Exposure** : Occlusive  
**Exposure time** : 24 hour(s)  
**Number of animals** : 6  
**PDII** : 3.1  
**Method** : Draize Test  
**Year** : 1985  
**GLP** : yes  
**Test substance** : API 83-05  
**Method** : 0.5 ml of undiluted test material was applied to the shorn skin in two areas on each rabbit. One area was intact and the other abraded skin. The treated area was then covered with an occlusive dressing.  
 After 24 hours the dressing was removed and the treated skin was wiped to remove any residue of test material. The degree of erythema and edema was recorded according to the Draize scale. A second reading of skin responses was made at 72 hours and again at 5, 7 and 14 days. Results of the 24 and 72 hour readings were used to determine the Primary Irritation Index.

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**Result** : The scores for erythema and edema at each of the observation times were as follows:

	Erythema		Edema	
	Intact	Abraded	Intact	Abraded
24 h	1.2	1.5	1.5	1.8
72 h	1.5	1.5	1.7	1.8
5 days	1.0	1.3	1.5	1.7
7 days	0.8	1.0	1.0	1.0
14 days	0	0	0	0

The Primary dermal Irritation index was 3.1

Growth rates were normal throughout the study and there were no clinical signs of systemic toxicity.

**Reliability** : (1) valid without restriction  
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(6)

### 5.2.2 EYE IRRITATION

**Species** : Rat  
**Concentration** : Undiluted  
**Dose** : 0.1 ml  
**Comment** : rinsed eye after 30 sec exposure in 3 rabbits only  
**Number of animals** : 9  
**Method** : Draize Test  
**Year** : 1985  
**GLP** : yes  
**Test substance** : TS: API 83-05  
**Method** : 0.1 ml of undiluted test material was dripped onto the corneal surface of one eye of each of 9 rabbits; the other eye was untreated and served as control. After 30 seconds the treated eyes of 3 rabbits were washed with lukewarm water for 1 minute. Eyes of the other 6 rabbits were not washed. Readings of ocular lesions for all animals were made at 1, 24, 48, 72 hours and 7 days after treatment. At the 72 hour and seven day readings, sodium fluorescein was used to aid in revealing possible corneal injury. Body weights were recorded just prior to treatment and one week afterwards. At termination of the study, the rabbits were euthanized by an overexposure of carbon dioxide and were subjected to a gross necropsy. Any abnormalities found were recorded.

**Result** : No signs of systemic toxicity were observed during the study. The primary eye irritation scores were as follows:

Observation period	Primary eye irritation score	
	Unwashed*	Washed**
1 hour	7.2	7.3
24 hour	5.5	2.7
48 hour	4.3	2.0
72 hour	3.0	2.0
7 day	1.0	1.3
14 day	0	0

\* Mean of six rabbits

\*\* Mean of three rabbits

In rabbits whose eyes had not been washed, irridial irritation that had occurred had subsided by 24 hours and

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all corneal involvement had subsided by 48 hours.  
No corneal or irridial irritation was observed in the group whose eyes had been washed followed by application of test material.

**Reliability** : (1) valid without restriction (6)  
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### 5.3 SENSITIZATION

**Type** : Buehler Test  
**Species** : guinea pig  
**Concentration** : Induction 50 % occlusive epicutaneous  
Challenge 25 % occlusive epicutaneous  
**Number of animals** : 20  
**Vehicle** : Paraffin oil  
**Result** : not sensitizing  
**Year** : 1986  
**GLP** : yes  
**Test substance** : API 83-05  
**Method** : 0.4 ml of a 50% mixture of test material and paraffin oil was applied under an occlusive dressing to the shorn skin of 10 male and 10 female animals. 6 hours after application, the dressings were removed and the skin wiped to remove residues of test material. The animals received one application each week for 3 weeks. The same application site was used each time. 2 weeks following the third application a challenge dose (0.4 ml of a 25% mixture in paraffin oil) was applied in the same manner as the sensitizing doses. A previously untreated site was used for the challenge application. The application sites for sensitizing and challenge doses were read for erythema and edema 24 and 48 hours after patch removal. To assist in the reading of the response to the final challenge dose, the test site was depilated 3 hours prior to reading by using a commercially available depilatory cream.

**Result** : Positive control (2,4-dinitrochlorobenzene), vehicle control and naive control groups were included in this study and the procedure for these was the same as for the test groups. The positive control was used at a concentration of 0.3% in 80% aqueous ethanol for the induction doses and at 0.1% w/v in acetone for the challenge dose.

: There was no abnormal appearance in any of the animals exposed to the test material during the study. The skin reactions to the challenge dose are summarized as follows:

Test material:	No dermal reactions by any animal
Naive control:	Very slight erythema in 2/20 animals
Vehicle control:	No dermal reactions by any animal
Positive control:	Very slight to moderate irritation by all 20 animals. The reactions of 16 of the animals exceeded the highest reaction observed in the naive positive control animals.

Naive positive control: 10/20 animals exhibited very slight erythema.

**Reliability** : (1) valid without restriction (10)  
19.11.2001

## 5.4 REPEATED DOSE TOXICITY

<b>Species</b>	: rat
<b>Sex</b>	: male/female
<b>Strain</b>	: Sprague-Dawley
<b>Route of admin.</b>	: inhalation
<b>Exposure period</b>	: 13 weeks
<b>Frequency of treatment</b>	: 6 hours/day, 5 days/week
<b>Post obs. period</b>	: Recovery group 4 weeks
<b>Doses</b>	: 750, 2500 and 7500 ppm
<b>Control group</b>	: yes
<b>NOAEL</b>	: 2500 ppm
<b>Method</b>	: OECD Guide-line 413 "Subchronic Inhalation Toxicity: 90-day Study"
<b>Year</b>	: 2000
<b>GLP</b>	: yes
<b>Test substance</b>	: LCRN-D (distillate of LCRN)
<b>Method</b>	: The method used was described in OECD guideline 413.

Groups of 16 male and 16 female rats underwent whole body exposures to 750, 2500 and 7500 ppm LCRN-D. Exposures were for 6 hours each day, 5 days per week for 13 weeks. Extra groups of 16 rats of each sex were exposed to the high dose level and also for a recovery control group. These animals were maintained untreated for 28 days following cessation of the 13 weeks exposure.

Neurobehavioral evaluations of motor activity and functional activity were performed pretest and during weeks 5, 9, 14 and 18 recovery groups. Animals were not exposed to LCRN-D during these tests.

Following 13 weeks of exposure, 16 animals/sex/group were necropsied and microscopic examination was performed on selected tissues. Nervous tissue from 6 rats/sex/group was also examined microscopically.

At the end of the 4 week recovery period, 16 animals of each sex from the high and control groups were necropsied and selected tissues were examined microscopically.

During the study, clinical observations were made twice daily. Ophthalmoscopic evaluations were performed pretest and just prior to the scheduled sacrifices at 14 weeks and 19 weeks (recovery groups). Body weights and food consumption was measured throughout the study. Blood samples were taken from 10 fasted rats/sex/group at 14 and 18 weeks for hematological and clinical chemical measurements.

At termination (after 13 weeks exposure for the main study and after 19 weeks for the recovery animals) all animals were killed and subjected to a complete macroscopic examination. 10 animals of each sex were designated for non-neuropathological examination and 6 of each sex for neuropathological examination.

For the non-neuropathology animals, the following organs were weighed: adrenals, brain, heart, kidneys, liver, lung, ovaries, prostate, spleen, testes (with epididymes), thymus and uterus. Brain lengths and widths were measured for each rat.

A wide range of tissues (39) was removed from the control and high dose animals, were fixed and examined histopathologically. Additionally, kidneys from selected animals were stained with Mallory-Heidenhain and examined. Tissues were also removed from the nervous system (central and peripheral) of all animals for subsequent special staining and histopathological examination. Animals designated for neuropathological examination were subjected to a detailed examination of central and peripheral nervous tissues. N

Neurobehavioral studies were undertaken as follows:

#### Motor activity

Locomotor activity was monitored as the number of beam breaks in an activity box. Monitoring sessions were for 60 minutes, divided into twelve 5-minute intervals. Evaluation was made pretest and during weeks 5, 9, 14 and at the end of the 4 week recovery period. [A detailed description of the evaluation and analysis is provided in the publication but is not included here.]

#### Functional Operational Battery

An assessment of the following was made:

Home cage evaluations for Posture, vocalization, palpebral closure.

Handling evaluations for reactivity to general stimuli, signs of autonomic function.

open field behavior: arousal level, gait, urination and defecation frequency, convulsions, tremor, abnormal behavior, piloerection and exophthalmos.

Reflex assessments for: response to visual and auditory stimuli, tail pinch, pupillary function.

Animals were also evaluated for fore limb and hind limb grip strength, landing foot splay and air righting ability.

#### Result

: There were no mortalities during the study and there were no treatment-related signs of toxicity. The ophthalmic examinations did not reveal any treatment-related effects. Mean body weights, body weight gains and food consumption were unaffected by treatment.

No treatment-related effects were recorded in the Functional Operational Battery. In the examinations of motor activity, there were no treatment-related effects recorded during the 13 week exposure period but a slight increased activity was found in the highest treatment group after the 4 week recovery period.

After 13 weeks exposure there was a significant decrease in total WBC count (36%) and lymphocyte counts in the high dose males and a slight decrease in neutrophil counts for the mid dose males. A trend towards decreased WBC (2.1%) and lymphocyte counts was also seen in the mid dose males and high dose females. After the 4 week recovery period, leukocyte values were comparable to control values. However, MCV was slightly decreased (2.8%) in the high dose males. It was concluded that these changes were suggestive of a reversible slight effect of the LCRN-D.

## 5. Toxicity

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Clinical chemistry parameters were unaffected by treatment.

After 13 weeks exposure relative kidney weights in the high dose males were increased (15.9%) and this correlated with the occurrence of hyaline droplets in the proximal convoluted tubules. This finding has been described as a "light hydrocarbon nephropathy" and is sex and species specific and is not relevant for human health risk assessment.

In the high dose males decreased absolute (25.7%) and relative (22%) spleen weights were also recorded. It was concluded that this was associated with the minor hematological changes that had been observed. These differences were not apparent after the recovery period and no abnormal microscopic findings were found in either the spleen or bone marrow.

Brain length and width measurements were unaffected by treatment and there were no abnormal microscopic findings in the brain, spinal cord or peripheral nerves.

**Reliability** : (1) valid without restriction (22)  
19.11.2001

**Species** : Rat  
**Sex** : male/female  
**Strain** : Sprague-Dawley  
**Route of admin.** : Inhalation  
**Exposure period** : 13 weeks  
**Frequency of treatment** : 6 hours/day, 5 days/week  
**Doses** : 410, 1970 and 8050 ppm  
**Control group** : Yes  
**NOAEL** : 1970 ppm  
**Year** : 1996  
**GLP** : no data  
**Test substance** : Partially vaporized full range catalytic reformed naphtha  
**Method** : Groups of 15 rats of each sex were individually housed in 1m<sup>3</sup> inhalation chambers. The rats underwent whole body exposures to partially vaporized full range catalytic reformed naphtha (FRCRN). Exposures were [6 hours/day, 5 days/week] for 13 weeks at nominal concentrations of 500, 2000 and 8000 mg/m<sup>3</sup>.  
Two extra groups of 15 rats of each sex served as sham and untreated controls. (NB. This is not stated in the publication but from other comments in the paper, it is clear that exposure was not continual during the study).  
Water was available ad lib, but food was withheld during the exposure periods.  
Clinical observations were made regularly and body weights were recorded weekly.

At the end of the 13 weeks exposure, blood samples were taken for hematological and clinical chemical measurements. The rats were then sacrificed and necropsied. Organs were weighed and a wide range of tissues fixed for subsequent histology and microscopic examination. The wet weight of the right middle lung lobe was also weighed. The lobes were then dried and their dry weights determined. The cauda epididymis of the control and high dose male rats was used to determine the morphology and number of sperm and the left testis was used to determine the number of testicular spermatids.

**Result**

The following tissues from the high dose animals were examined histologically: adrenals, bone and marrow (sternum), pancreas (head), brain (three sections), submaxillary salivary gland, eye, optic nerve, spleen, heart, stomach (squamous and glandular), colon, testes or ovaries, duodenum, kidneys, thymus, thyroid, liver, tracheobronchial lymph nodes, lung (left lobe), nasal turbinates (four sections), thigh muscle, urinary bladder, sciatic nerve, and any gross lesions. In addition, tracheobronchial lymph nodes and any gross lesions from untreated control animals were also evaluated.

: There were no treatment-related clinical signs during the study, no effects on serum chemistry values or parameters of the male reproductive system a terminal sacrifice. Body weights of males were exposed t the mid and high dose groups were higher than the controls throughout the study and the differences were statistically significant in the high dose group from week 10 onwards.

WBC count was significantly lower in sham treated controls and all three treated groups in both sexes compared to untreated controls. Additionally the WBC count was decreased by approximately 24% in the high dose females when compared to the sham controls. No other parameters were affected.

The only organ weights affected were the liver and kidney. In the male high dose group, mean kidney weight was approximately 13% greater than the sham treated animals (but not the untreated controls), and the liver weight was approximately 14% greater.

**Test substance**

No treatment-related gross lesions were observed at necropsy and no treatment-related abnormalities were noted during microscopic examination. Because of the lack of effects in the histology, no tissues were examined from the lower dose groups.

: Test atmospheres were generated by partially vaporizing FRCRN. The concentrations in the chamber were adjusted by dilution with air. Concentrations were monitored throughout the study The actual concentrations for each of the dose levels are shown below.

Parameter	Exposure group		
	Low	Medium	High
Target conc. (mg/m3)	500	2000	8000
Actual conc. (mg/m3)	410	1970	8050
Butane	4.33	3.91	4.05
Methylbutane	20.56	17.26	17.55
Pentane	13.24	11.44	11.86
Hexane	6.53	5.71	6.36
Heptane	2.32	2.35	2.33
Benzene	2.19	4.93	5.79
Toluene	10.02	12.23	10.93
m- and p- Xylenes	3.57	4.05	3.4
2- Ethyltoluene	0.43	0.35	0.17
Trimethylbenzene	0.01	0.01	0.04

**Reliability**

: (2) valid with restrictions  
The publication is not clear in its description of the frequency and duration of exposures. However, it is assumed that the exposures are 6 hours/day, 5 days/week since this would be consistent with other studies reported from the same laboratory.

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(14)

## 5. Toxicity

**Id** A. Naphthas  
**Date** 10.12.2001

**Species** : rabbit  
**Sex** : male/female  
**Strain** : New Zealand white  
**Route of admin.** : dermal  
**Exposure period** : 6 hours  
**Frequency of treatment** : 3 times per week for 28 days (12 applications total)  
**Doses** : 200, 1000 & 2000 mg/kg  
**Control group** : yes, concurrent no treatment  
**Year** : 1986  
**GLP** : yes  
**Test substance** : API 83-05  
**Method** : Prior to the study a 5-day range finding study was conducted. The method and results of the range-finding study are not included in this summary.

Undiluted API 83-05 was applied at doses of 200, 1000 and 2000 mg/kg/day to the shorn dorsal skin of groups of five male and five female rabbits. The test material was applied to the skin 3 times each week for 4 weeks (12 applications total). The applied material was covered with an occlusive dressing for 6 hours, which was then removed and the skin was wiped with a dry gauze to remove any residual material. A group of five rabbits of each sex served as sham controls. The test skin site of each animal was examined and scored for irritation prior to each application of test material. Mortality and moribundity checks were performed twice daily and body weights were recorded weekly. At termination, blood samples were taken for a range of hematological and clinical chemical measurements. Urine samples were also collected and frozen for possible future examination.

A complete gross necropsy was performed on all animals. Major organs were weighed and tissues were processed for subsequent histopathological examination.

**Result** : Two males in the 2000 mg/kg group and one male in the 1000 mg/kg died during the study. The deaths occurred on days 12 and 17 for the highest dose group and day 19 at the mid dose group. There were no clinical signs of intoxication in any other animal on the study. At 200 and 1000 mg/kg there were no treatment related effects on body weight gains over the study period although there were isolated differences between treated and control animals during the study. At the highest dose level, the females showed no weight gain and the males had an overall weight loss. A mean irritation score was calculated for each day and overall means were also calculated. (The mean irritation score, MIS, was the sum of irritation scores for both erythema and edema for all animals of a given dose group and sex.)

The overall MIS for each dose group was:

Group (mg/kg)/sex	MIS	Classification
2000 M	5.1	Severe irritant
2000 F	4.9	Severe irritant*
1000 M	4.3	Moderate irritant
1000 F	4.1	Moderate irritant
200 M	2.9	Moderate irritant
200 F	2.5	Moderate irritant
Control M	0	Non irritant
Control F	0	Non irritant

\*Severe irritation was observed in the high dose females during the study and the authors concluded that even though the overall MIS for this group led to a moderate classification, a severe classification would be more appropriate. Some differences were observed between the control and treated groups for a few hematological and clinical chemical parameters. The differences from control values are shown below. However, the authors concluded that since the values fell within the normal range for the laboratory, they should not be regarded as treatment related.

<u>Dose group</u>	<u>Parameter</u>	<u>Difference</u>
1000 mg (M)	WBC	25% higher
	Alkaline phosphatase	37% lower
2000 mg (M)	Hemoglobin	5% lower
	Blood urea nitrogen	24% lower
	Alkaline phosphatase	35% lower
2000 mg (F)	SGPT	26% lower

There were also a few differences between control and treated animals for absolute and relative organ weight for a small number of organs. Since there was no associated histopathological findings and since the differences were not dose-related, they were not considered to be significant. At necropsy, few gross findings were recorded other than effects on the treated skin. The findings in the liver of males and females of treated and control groups consisted of areas of discoloration and were considered to incidental to treatment.

Histological changes were mainly confined to the skin except for two males that died in the highest dose group. The kidneys of these two animals contained slight to moderate tubular degeneration.

The changes in the skin consisted of slight to moderate proliferative and inflammatory changes at the highest dose group. Concurrent with these changes in the skin there was an increased granulopoiesis of the bone marrow, probably related to the stress or other factors associated with the skin irritation. Such changes were not observed in the control animals.

**Reliability**  
 19.11.2001

: (1) valid without restriction

(9)

**5.5 GENETIC TOXICITY 'IN VITRO'**

<b>Type</b>	: Mouse lymphoma assay
<b>System of testing</b>	: Forward mutation assay using cell line L5178Y TK+/-
<b>Cytotoxic conc.</b>	: 250µl/ml, lethal at 500µl/ml
<b>Metabolic activation</b>	: with and without
<b>Year</b>	: 1985
<b>GLP</b>	: yes
<b>Test substance</b>	: API 83-05
<b>Method</b>	: The test material was dissolved in acetone for this assay. Three positive control substances were used viz Ethyl methane sulphonate (EMS) at concentrations of 0.25 & 0.4 µl/ml for the non activation assay, Dimethylnitrosamine (DMN) at a concentration of 0.3 and Methylcholanthrene (MCA) at a concentration of 2.5 µg/ml for the activation assay.

A cytotoxicity study was carried out prior to the mutagenicity assay. The test material was lethal at a concentration of 500 µl/ml and highly toxic at 250 µl/ml without S-9. These results were used to select a dose range of 6.25 to 500 µl/ml for the non-activation assay and 3.13 to 400 µl/ml for the activation assay.

For the mutation assay, the lymphoma cells were exposed for 4 hours to test material. After exposure to the test material, the cells were allowed to recover for 2 days and then cultures were selected for cloning and mutant selection; TFT was used as the restrictive agent.

six non-activated and six activated cultures were selected for cloning based on their degree of toxicity. The non-activated cultures that were cloned were treated with 6.25, 25, 37.5, 50, 75 and 100µl/ml of test material and resulted in a range of growth of 30 to 97% compared to the solvent control. The activated cultures that were cloned were treated with 18.8, 37.5, 75, 100, 150 and 200 µl/ml of test material. This resulted in growth ranging from 4.6 to 67.9% compared to solvent control.

Plates were prepared from TFT and from the VC cultures and after 10 to 12 days incubation these plates were scored for total number of colonies per plate. A mutation frequency was then determined.

The following criteria were used in judging the significance of the activity of the test article.

Positive - if there is a positive dose response and one or more of the 3 highest doses exhibit a mutant frequency which is two-fold greater than background level.

Equivocal - if there is no dose response but any one or more doses exhibit a two-fold increase in mutant frequency over background.

Negative - if there is no dose response and none of the test cultures exhibit mutant frequencies which are two-fold greater than background.

<b>Result</b>	: The mutant frequencies and the percentage total growth at each of the test concentrations is summarized in the following table.
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## 5. Toxicity

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Concentration ( $\mu\text{l/ml}$ )	Mutant frequency	% Relative growth
<u>Non-Activated</u>		
6.25	24.2	97.3
25	22.5	64.3
37.5	18.2	32.6
50	23	47.8
75	39.6	59.4
100	22.3	29.6
Solvent 1	100	22.7
Solvent 2	100	30.6
Untreated control	20.7	110.6
EMS 0.25 $\mu\text{l/ml}$	364.5	53.8
EMS 0.4 $\mu\text{l/ml}$	504.5	23.2

<u>S-9 Activated</u>		
18.8	54.2	67.9
37.5	57.3	56.1
75	72.1	60.3
100	85.2	32.8
150	73	27.4
200	146.2	4.6
Solvent 1	31.3	100
Solvent 2	30.8	100
Untreated control	42.1	123.9
DMN 0.3 $\mu\text{l/ml}$	258.8	12.7
MCA 2.5 $\mu\text{l/ml}$	243.6	78.5

The authors concluded that the test material was not mutagenic in the non-activated assay because there was no dose response relationship and furthermore the mutant frequency was not significantly different from the solvent and untreated controls.

The minimum criteria for indicating mutagenesis would have been  $47 \times 10^{-6}$ . Since the 100  $\mu\text{l/ml}$  treatment represented a close approach to the excessively toxic treatment at 150  $\mu\text{l/ml}$ , this assay was considered sufficient to evaluate the test material as non-mutagenic under non-activation conditions.

In the presence of the S-9 mix, the test material was converted into one or more mutagenic products.

The minimum criterion for a significant response was a mutant frequency exceeding  $62.1 \times 10^{-6}$ . This value was exceeded for 4 of the 6 analyzed cultures. The response was dose related.

The results were judged sufficient to evaluate the test material as mutagenic in the presence the metabolic activation system.

Reliability  
19.11.2001

: (1) valid without restriction

(8)

## 5.6 GENETIC TOXICITY 'IN VIVO'

Type	: Cytogenetic assay
Species	: rat
Sex	: male/female
Strain	: Sprague-Dawley
Route of admin.	: i.p.
Exposure period	: one single dose given
Doses	: 0.26, 0.82 & 2.42 g/kg
Result	: negative
Year	: 1985
GLP	: yes
Test substance	: API 83-05
Method	: Two studies were carried out. In the first study, the test material did not induce a significant increase in the percentage of aberrant cells above the controls in either sex. Furthermore, the positive control (TEM at a dose of 0.8 mg/kg) did not induce a significant elevation in the percentage of cells with structural aberrations. The assay was, therefore, repeated using a higher dose of TEM. In this summary, only the results of the repeat study are described.

The study design was as follows:

Treatment	Animals/sex/sacrifice time		
	6 hrs.	24 hrs.	48 hrs.
Corn oil (vehicle)	5	5	5
API 83-05, 2.42 g/kg	5	5	5
API 83-05, 0.82 g/kg	5	5	5
API 83-05, 0.26 g/kg	5	5	5
Triethylenemelamine (Positive control)		5	

Test material in vehicle was given intraperitoneally at a dose of 5 ml/kg to groups of rats as shown above. Corn oil was used as vehicle control and TEM (1.5 mg/kg) as the positive control.

Four hours prior to sacrifice the rats were given a single intraperitoneal dose of colchicine (4 mg/kg).

One male in the 2.42 g/kg group and one male in the 0.82 g/kg dose group died immediately after dosing, these were replaced by substitute animals.

Immediately after sacrifice, bone marrow was obtained from the tibiae of the animals. The marrow was washed and the cells were fixed before being spread on slides (at least 3 from each animal) for examination.

Slides were scored for chromosomal aberrations.

Where possible, a minimum of 50 metaphase cells from each animal were examined and scored for chromatid and chromosome gaps and breaks, fragments, structural rearrangements and ploidy (1-3).

A mitotic index (= No. of cells in mitosis/500 counted X 100) was calculated and recorded.

The type of aberration, its frequency, the statistical significance of any increases and its correlation to dose in a given time period will all be considered in evaluating a test article as being mutagenically positive or negative. Criteria for a positive response are generally a

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### Result

statistically significant dose-related increase in the number of structural aberrations at three dose levels. The final decision is based on scientific judgment.

: The dose levels used in the assay were selected on the basis of a preliminary screen.

In the cytogenetics assay, one male died at each of the dose levels 2.42 and 0.82 g/kg, the mortality occurred immediately after dosing. Toxic signs included lethargy and a moribund appearance at the high dose and slow uncoordinated movement in the mid dose group. The results of the cytogenetics evaluations are summarized in the following table.

#### MALES

	Dose level (g/kg)			Positive control	Vehicle control
	0.26	0.82	2.42		
Cells with 1 or more aberrations					
6 hrs	0.5	0.4	1.0		0.5
24 hrs	0.4	0.8	1.0	32.4	0
48 hrs	0	1.6	0.5		0.8
Cells with 2 or more aberrations					
6 hrs	0	0	0		0.5
24 hrs	0	0	0	10.8	0
48 hrs	0	0	0		0
Frequency of structural aberrations					
6 hrs	.005	.004	.01		.05
24 hrs	.004	.008	.01	.708	0
48 hrs	0	.016	.005		.008
Frequency of numerical aberrations					
6 hrs	.005	0	.016		.015
24 hrs	.008	.008	.01	.008	.005
48 hrs	.01	.008	0		.004
Mitotic Index					
6 hrs	4.1	3.6	2.4		5.4
24 hrs	4.4	5.7	5.5	6.3	5.0
48 hrs	6.5	5.1	5.2		5.7

#### FEMALES

Cells with 1 or more aberrations					
6 hrs	0	0.5	1.6	0.8	
24 hrs	0	0.4	1.5	0	33.2
48 hrs	0	1.2	0.8	1.2	
Cells with 2 or more aberrations					
6 hrs	0	0	0	0	
24 hrs	0	0	0.5	0	13.2
48 hrs	0	0	0	0	
Frequency of structural aberrations					
6 hrs	0	.005	.016	.008	
24 hrs	0	.004	.02	0	.804
48 hrs	0	.012	.008	.012	
Frequency of numerical aberrations					
6 hrs	.005	.005	.020	0	
24 hrs	.01	.016	.005	.020	.020

## 5. Toxicity

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48 hrs	.008	0	.012	.005
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### Mitotic Index

6 hrs	5.4	6.3	3.1	6.1	
24 hrs	4.9	5.4	4.1	4.8	4.7
48 hrs	5.5	4.9	7.0	5.2	

NB.1. For simplicity only, mean values without standard errors are shown in the above table although they are given in the laboratory report.

On the basis of the criteria defined for assessing the results, the authors concluded that API 83-05 was not mutagenic in this assay.

**Reliability** : (1) valid without restriction  
19.11.2001

(7)

### 5.7 CARCINOGENITY

No data available

### 5.8 TOXICITY TO REPRODUCTION

**Type** : Reproductive/developmental toxicity screening study, (OECD 421)  
**Species** : Rat  
**Sex** : male/female  
**Strain** : Sprague-Dawley  
**Route of admin.** : inhalation  
**Frequency of treatment** : 6 hours/day, 7 days/week  
**Premating exposure period**  
**Male** : 2 weeks  
**Female** : 2 weeks  
**Duration of test** :  
**Doses** : Target conc.: 750, 2500 & 7500 ppm. Actual conc.: 750, 2490 & 2490 ppm  
**Control group** : yes  
**Method** : OECD combined repeated dose and reproductive/developmental toxicity screening test  
**Year** : 2000  
**GLP** : yes  
**Test substance** : LCRN distillate (LCRN-D)  
**Method** : Groups of 10 rats of each sex were exposed to 750, 2500 or 7500 ppm. LCRN-D for 6 hours /day, seven days/week. A group of 10 rats of each sex served as sham treated controls. Parental females were exposed for 14 consecutive days prior to mating, throughout mating and days 0-10 of gestation. Dams and their litters were sacrificed on post partum day 4. Unmated females and parental males were exposed to the test material for 14 days prior to mating, throughout mating and 18 additional days following completion of the mating period. These animals were sacrificed shortly after the last litters were delivered reached post partum day 4.

### Mating

Within each group one male was co-housed with the same female until evidence of mating was observed (presence of sperm in vaginal smear or copulatory plug). The day of mating was designated day 0 of gestation. Following mating, the females were housed individually and continued their

exposures to test material until day 19 of gestation. Females not showing evidence of mating following a 14 day mating period continued their exposures. If such a female showed signs of being pregnant, it was removed from the exposure regimen and observed for parturition.

Observations

All parental animals were regularly observed for mortality and gross pharmacologic signs. A physical examination, including palpation for tissue masses was carried out daily 30 mins. after removal from the exposure chambers. Body weights and food consumption were measured throughout the study.

From day 20 of gestation, females (pregnant and non-pregnant) were observed for signs of parturition.

As soon as possible after delivery, litters were observed for the number of live and dead pups and for any abnormalities. Litters were also observed twice daily for unusual findings and dead pups.

On days 0 and 4 of lactation, the pups were counted, weighed and sex was determined by external observation.

Pathology

Males were killed as a group shortly after the last litters delivered had reached day 4 of lactation.

Females with litters that reached day 4 of lactation were killed the next day or shortly thereafter.

Unmated females and those that did not deliver were killed 23 days after completion of the mating period.

At post mortem, a complete macroscopic examination was carried out on all adult animals. The following organs were weighed and organ/body weight ratios were calculated: adrenals, brain, heart, kidneys, liver, lung, spleen, epididymes, testes and thymus. Post mortem examination of females included a count of uterine implantation scars when present.

Pups were sacrificed on day 4 of lactation and underwent a complete macroscopic examination and a determination of sex by internal examination. All pups were preserved with viscera intact. Pups found dead at birth and that died prior to day 4 of lactation also underwent a gross external and internal examination. Dead pups were not eviscerated, the intact pups were preserved.

27 tissues were preserved from all adult animals in all dose groups. Ovaries, testes, epididymes, nose with nasal turbinates, and any grossly observed abnormalities were processed and sections examined histologically for all males and female parental animals in the control and highest dose group. Four sections were prepared and examined microscopically of the skull containing the nasal turbinates. These were

- area between upper incisor and incisive papilla
- area between incisive papilla and first palatal ridge
- area between second palatal ridge and first upper molar
- area between first upper molar and nasopharynx.

Result

: All parental animals survived to scheduled sacrifice and no treatment related clinical signs were observed. Except for a slight reduction in body weights in the high dose males there were no other effects on either body weight or food consumption. When compared to the controls, at week

3 the decrease in weight of the high dose males was 3.8% and at week 7 was 7.8.

The only treatment related organ weight changes were an increase in relative kidney (15%) and relative liver (5%) weights in the high dose males. No other organ weight changes were recorded.

There were no treatment-related microscopic changes in the testes, epididymes, ovaries or nasal turbinates in the animals in the high dose group.

**Reproductive/fertility effects**

All groups had a mating index and a fertility index of 100% and all animals in all groups had mated within 4 days of cohabitation.

Delivery and litter data did not demonstrate any effects of treatment see data summarized below.

Parameter	Dose group (ppm)			
	0	750	2500	7500
Females on study	10	10	10	10
Litters with liveborn	10	10	10	10
Implantation sites	147	154	155	154
Mean	14.7	15.1	15.5	15.4
Pups delivered (total)	145	151	146	145
Liveborn	142	151	143	144
Live birth index (%)	98	100	98	99
Pups dying				
Day 0	0	1	1	1
Days 1-4	2	4	0	0
Pups surviving 4 days	140	146	142	143
Viability index (%)	99	97	99	99
pup sex distribution				
Day 0 M/F (ratio)	63/79	67/84	69/74	68/76
Day 4 M/F (ratio)	63/77	64/82	68/74	68/75
Pup weight/litter (g)				
Day 0	6.0	6.6	6.2	6.1
Day 4	9.3	8.9	9.2	9.6

External and internal examination of pups sacrificed on day 4 of lactation resulted in only one pup in a single litter of the control group with abnormalities.

**Reliability**  
19.11.2001

: (1) valid without restriction

(23)

## 5.9 DEVELOPMENTAL TOXICITY/TERATOGENICITY

<b>Species</b>	:	rat
<b>Sex</b>	:	male/female
<b>Strain</b>	:	Sprague-Dawley
<b>Route of admin.</b>	:	inhalation
<b>Exposure period</b>	:	Gestation days 6-19 inc.
<b>Frequency of treatment</b>	:	6 hours/ day
<b>Doses</b>	:	Target concentrations: 2000 and 8000 ppm. Actual: 2160 and 7800 ppm
<b>Control group</b>	:	yes
<b>NOAEL Maternalt.</b>	:	7800 ppm
<b>NOAEL Teratogen</b>	:	7800 ppm
<b>Year</b>	:	1996
<b>GLP</b>	:	no data
<b>Test substance</b>	:	Partially vaporized full range catalytic reformed naphtha
<b>Method</b>	:	Groups of 11 or 12 presumed pregnant female rats were exposed 6 hours each day from days 6-19 of gestation to whole body exposures of 2000 or 8000 ppm partially vaporized FRCRN. Two extra groups served as untreated and sham treated controls. All animals were observed daily and body weights were recorded on days 0, 6, 13 and 20 of gestation. On day 20 each female was sacrificed and blood samples removed for serum chemistry evaluations. Parameters measured were the same as those in the subchronic study by the same authors, and in addition included iron and lactic dehydrogenase.  All organs were examined grossly and liver and thymus weights were recorded. In addition, the number of corpora lutea per ovary and the gravid uterine weights were recorded. Uterine contents were examined and the numbers of implantation sites, early and late resorptions and live and dead fetuses were recorded.  Each fetus was gendered, weighed and grossly examined for external abnormalities. Half the fetuses were fixed in Bouin's fluid and examined subsequently for soft tissue abnormalities. Remaining fetuses were stained with Alizarin red and examined for skeletal anomalies.
<b>Result</b>	:	There were no adverse effects on maternal body weight gain, liver weight or thymus weight. In the high dose group, maternal serum glucose levels were significantly decreased (1.5%) and potassium levels increased (1%) relative to the untreated controls.  Reproductive performance during gestation and in-utero survival and development of concepti were unaffected by treatment. Furthermore, there were no treatment-related increases in gross abnormalities or anomalies of soft or skeletal tissues.
<b>Reliability</b>	:	(1) valid without restriction
19.11.2001		

(14)

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Date 10.12.2001

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## Additional mutagenicity information

# Olefinic Naphthas

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## 5.5 GENETIC TOXICITY 'IN VITRO'

<b>Type</b>	: Mouse lymphoma assay
<b>Metabolic activation</b>	: With and without
<b>Result</b>	: Negative
<b>Year</b>	: 1987
<b>GLP</b>	: Yes
<b>Test substance</b>	: API 83-20
<b>Method</b>	: The test material was dissolved in Ethanol for this assay. Two positive control substances were used viz Ethyl methane sulfonate (EMS) at concentrations of 0.25 & 0.5 µl/ml for non activation assays and 3-methylcholanthrene (MCA) at concentrations of 2.5 & 4.0 µg/ml for activation assays.

A cytotoxicity study carried out prior to the mutagenicity assay established that the sample was highly toxic at 500 nl/ml without activation and lethal at the same concentration in the presence of metabolic activation.

Therefore, for the mutation assay the lymphoma cells were exposed for 4 hours to test material at treatments from 50 to 800 nl/ml without activation and with treatments from 25 to 500 nl/ml with S-9 activation. After exposure to the test material, the cells were allowed to recover for 2 days and then cultures were selected for cloning and mutant selection. Plates containing colonies of selected cells were incubated for 10 to 14 days after which they were scored for total number of colonies per plate. A mutation frequency was then determined.

Assay evaluation criteria were:

The minimum criterion considered necessary to demonstrate mutagenesis for any given treatment is a mutant frequency that is at least 150% of the concurrent background frequency plus  $10 \times 10^{-6}$ . The background frequency is defined as the average mutant frequency of the solvent negative controls. The minimum increase is based on extensive experience which indicates that assay variability increases with higher backgrounds and the calculated minimum increase as defined above is often a repeatable result; statistical analysis for the confidence limits is not yet available.

The observation of a mutant frequency that meets the minimum criterion for a single treated culture within a range of assayed concentrations is not sufficient evidence to evaluate a test material as a mutagen. The following test results must be obtained to reach this conclusion for either a activation or non-activation conditions.

A dose-related or toxicity-related increase in mutant frequency should be observed. It is desirable to obtain this relation for at least three doses, but this depends on the concentration steps chosen for the assay and the toxicity at which mutagenic activity appears.

If an increase of about two times the minimum criterion or greater is observed for a single dose near the highest testable toxicity, as defined in the Assay acceptance criteria, the test material will be considered mutagenic. Smaller increases at a single dose near the highest testable toxicity will require confirmation by a repeat assay.

Treatments that induce less than 10% relative growth are included

in the assay, but are not used as primary evidence for mutagenicity as it relates to risk assessment.

In the assay reported in this particular study, under non-activation conditions, the test material was excessively toxic at 300 nl/ml. Five treatments from 50 to 250 nl/ml were therefore chosen for the analysis of mutant induction and non-detectable to moderate toxicities were induced (relative growths 205.3% to 25.5%). None of the assayed treatments induced a mutant frequency that exceeded the minimum criterion of  $89.7 \times 10^{-6}$ . However, since it is desirable to include highly toxic treatments (10 to 20% relative growth) in an analysis, another non-activation assay was performed in an attempt to obtain a wider range of toxicities. In the second assay the test material was analyzed for mutant induction from 50 to 150 nl/ml.

In the presence of metabolic activation six treatments from 75 to 400 nl/ml were analyzed for mutant induction and a wide range of toxicities was induced (86.3 to 6.9% relative growths). The minimum criterion for mutagenesis in this assay was a mutant frequency exceeding  $81.4 \times 10^{-6}$ . None of the acceptable treatments induced a mutant frequency that exceeded the minimum criterion. One treatment with less than 10% relative growth (400 nl/ml) induced a mutant frequency that exceeded the minimum criterion, but the treatment was not acceptable for analysis because it did not fulfill the requirements of the assay evaluation criteria.

A second assay was therefore performed at treatments ranging from 200 to 300 nl/ml.

**Result** : Only the results of the second assays are summarized since the first assay was not considered acceptable (for the reasons given in the method section above).

Test condition	Cloning efficiency	Relative growth (%)	Mutant frequency ( $10E^6$ units)
<b>Non activation</b>			
Solvent control	100.5	100	49.1
Solvent control	109.7	100	42.5
Solvent control	107.5	100	45.6
EMS 0.25 µl/ml	86.8	82.5	286.7
EMS 0.4 µl/ml	72.7	56.2	469.7
Sample 83-20			
50 nl/ml	128.4*	144.7	38.5
100 nl/ml	100.1*	88.8	58.2
150 nl/ml	78.5*	81.6	60.5
<b>S9 activation</b>			
Solvent control	114.7	100	46.5
Solvent control	121.0	100	44.3
Solvent control	100.2	100	57.2
MCA 2.5 µg/ml	87.3	55.5	235.1
MCA 4 µg/ml	73.2	53.5	210.0
<b>Sample 83-20</b>			
200 nl/ml	88.7*	62.4	48.3
250 nl/ml	88.1*	68.8	65.5
250 nl/ml	64.3*	8.6	66.2
300 nl/ml	63.7*	11.1	74.3
300 nl/ml	59.7*	7.3	96.7

\* Cloning efficiency relative to solvent control

In the non activation assay, at most, low toxicities were induced without inducing significant increases above the background mutant frequency

(average of solvent controls). Higher toxicities could not be assayed because of a very sharp toxicity curve; a small increase in concentration from 150 to 175 nl/ml was excessively toxic. The test material was, therefore, considered non mutagenic without activation at concentrations that approached excessive toxicity.

In the activation assay, the 250 and 300 nl/ml treatments were duplicated to determine reproducibility. Low and high toxicities were induced by the assayed treatments (68.8 to 7.3% relative growths).

For a treatment to be considered mutagenic in this trial, a mutant frequency exceeding  $84.0 \times 10^{-6}$  was required. One treatment at 300 nl/ml induced a mutant frequency that exceeded this criterion but the increase was observed at less than 10% relative growth and a duplicate treatment at the same concentration was inactive.

The test material was, therefore, considered non-mutagenic with activation in this assay.

In the assays used in this evaluation, the average cloning efficiencies for the solvent controls varied from 70.5% and 105.9% without activation to 96.2% and 112.0% with activation, which demonstrated acceptable cloning conditions for the assays.

The negative control mutant frequencies were all within the expected range and the positive control compounds yielded mutant frequencies that were greatly in excess of the background.

Sample 83-20 is considered inactive in the mouse lymphoma assay, with and without metabolic activation.

**Remark**

- : Two other olefinic naphtha streams have been tested in a mouse lymphoma assay. The results are summarized below.

<b>Sample</b>	<b>Result</b>	<b>Reference</b>
API 81-03	Negative	32-31300
API 81-04	Negative without S -9 Equivocal with S-9	32-31710

**Reliability**  
13.08.2002

- : (1) Valid without restriction (2) (3) (4)

**Type****System of testing****Metabolic activation****Year****GLP****Test substance****Method**

- : Sister chromatid exchange assay  
: Chinese Hamster Ovary (CHO) cells  
: With and without

: 1988

: Yes

: API 81-03

: A cytotoxicity study was performed in order to select dose levels for the SCE assay.

For the SCE assay CHO cells were seeded in duplicate for each treatment condition and were incubated at 37°C in a humidified atmosphere for 16 to 24 hours.

Treatment was carried out by re-feeding two complete sets of flasks with complete medium for the non activation study or with S-9 reaction mixture for the activated study to which was added 50 µl of dosing solution of test control or article in solvent or solvent alone. An untreated control of cells in complete medium was also included.

In the non-activation study the cells were exposed for 28 hours. Two hours after exposure 0.01 mM BrdUrd was added to the treatment medium. At the end of the treatment period, the treatment medium was removed, the cells were rinsed and were then

exposed to colcemid (0.1 µg/ml) for a further 2 hours.

In the activation study exposure was for 2 hours. After the exposure period, the treatment medium was removed, the cells were washed with PBS, re-fed with medium containing BrdUrd and then incubated for a further 28 hours. Colcemid was added at a final concentration of 0.1 µg/ml for the last 2 hours of incubation.

For activated and non-activated assays, metaphase cells were harvested 2 hours after addition of colcemid. Cells were collected and fixed and stored until slides were prepared.

Slides were coded and scored without regard to treatment group. Only cells with  $20 \pm 2$  centromeres were selected for evaluation of SCEs. A total of 4 doses were scored including the highest test article dose where sufficient second-division metaphase cells were available. SCEs were scored in 25 cells from each duplicate culture to make up a total of 50 cells per treatment. The percentage of cells in first (M1), second (M2) or third division (M3) metaphase was also recorded for a total of 100 metaphase cells scored.

TEM was used as positive control at a concentration of 0.025 µg/ml in the non-activated assay.

In the activated assay CP was used at a concentration of 2.5 µg/ml. The solvent vehicle for the test article was used as the solvent control.

A test was deemed valid if the mean SCE/cell in the untreated control did not exceed 13 and the mean SCE/cell for the positive control must be at least double that of the negative control.

A test material is considered positive if it induces a doubling in SCE frequency over the solvent control at a minimum of three consecutive dose levels or if a dose responsive and statistically significant increase is observed over a minimum of 3 dose levels affected.

A statistically significant increase at one or more dose levels with no evidence of a dose response is assessed as equivocal or as negative according to the magnitude of the response and the number of dose levels affected.

**Result**

- : Dose levels for the SCE assay were selected following a preliminary toxicity test based upon cell proliferation after treatment relative to the solvent control.

CHO cells were exposed to solvent alone and to nine concentrations of test material ranging from 1 to 0.0001 µl/ml in the absence and presence of an S-9 reaction mixture.

The test material was partially insoluble at 1 µl/l. Further dilutions were completely soluble.

Based on the growth inhibition and cell cycle delay, dose levels of 0.3, 0.2, 0.1 and 0.05 µl/ml were selected for use in the assay without metabolic activation. For the assay with metabolic activation concentrations of 0.2, 0.1, 0.05 and 0.03 µl/ml were selected.

A harvest time of 30 hours after treatment initiation was selected to assure collection of enough analyzable second division metaphases at the high dose.

Summarized results for the assays are as follows:

## 5. Toxicity

Id Onaphtha  
Date 13.08.2002

Treatment/ replicate	Cell cycle kinetics			SCEs/ chromosome	Group mean SCEs/ cell ( $\pm$ SD)	
	M1	M2	M3			
<b><u>Without metabolic activation</u></b>						
Untreated	A	2	98	0	0.5	9.92 (3.16)
	B	5	95	0	0.48	
Acetone	A	4	96	0	0.52	9.94 (3.03)
	B	5	95	0	0.50	
<b>API 81-03 (<math>\mu</math>l/ml)</b>						
0.05	A	3	97	0	0.51	9.88 (3.12)
	B	2	98	0	0.51	
0.1	A	12	88	0	0.54	10.32 (3.01)
	B	9	91	0	0.52	
0.2	A	6	94	0	0.51	9.94 (3.11)
	B	5	95	0	0.52	
0.3	A	1	99	0	0.53	10.34 (2.58)
	B	4	96	0	0.53	
<b>TEM (<math>\mu</math>g/ml)</b>						
0.025	A	0	65	35	2.79	59.02 (12.31)
	B	0	81	19	3.21	
<b><u>With metabolic activation</u></b>						
Untreated	A	2	97	1	0.52	10.10 (3.12)
	B	2	95	3	0.51	
Acetone	A	4	96	0	0.45	9.38 (3.36)
	B	2	98	0	0.51	
<b>API 81-03 (<math>\mu</math>l/ml)</b>						
0.03	A	3	97	0	0.48	8.82 (2.46)
	B	5	95	0	0.44	
0.05	A	8	92	0	0.54	10.54 (2.7)
	B	5	95	0	0.56	
0.1	A	6	94	0	0.59	11.20 (3.45)
	B	3	97	0	0.55	
0.2	A	2	96	2	0.48	10.10 (2.59)
	B	1	94	5	0.57	
<b>CP (<math>\mu</math>g/ml)</b>						
2.5	A	2	98	0	1.63	32.72 (6.51)
	B	4	96	0	1.72	

In the assay with metabolic activation, the group mean SCEs/cell were significantly increased compared to controls,  $P=0.05$  for 0.05  $\mu$ l/ml concentration and  $P\leq 0.01$  for the 0.02  $\mu$ l/ml concentration. Positive controls in both assays were significant  $P=0.01$

The positive and negative controls fulfilled the requirements for a valid test. API 81-03 did not induce an increase in sister chromatid exchanges in

CHO cells when tested in the absence of metabolic activation. However the test material did induce a small but statistically significant increase in SCEs at two intermediate dose levels in the presence of metabolic activation which was concluded to be equivocal.

**Reliability** : (1) Valid without restriction (6)  
09.08.2002

06.08.2002

**5.6 GENETIC TOXICITY 'IN VIVO'**

**Type** : Cytogenetic assay  
**Species** : Rat  
**Sex** : Male/female  
**Strain** : Sprague-Dawley  
**Route of admin.** : i.p.  
**Doses** : 0.3, 1.0 and 3.0 g/kg  
**Result** : Negative  
**Year** : 1985  
**GLP** : Yes  
**Test substance** : API 81-04  
**Method** : Test material was given intraperitoneally as a solution in corn oil at a rate of 5 ml/kg to groups of 15 rats of each sex at three different dose levels (0.3, 1.0 & 3.0 g/kg). A group of 15 rats of each sex were given corn oil and these animals served as vehicle controls. A group of 5 animals of each sex to be used as positive controls was dosed with triethylenemelamine (TEM) at a level of 0.5 mg/kg and these animals were killed 24 hours afterwards. Two to four hours prior to being killed the rats were given a single ip dose of colchicine (1 mg/kg).  
 For each dose level of test material and the negative controls 5 rats of each sex were killed 6, 24 and 48 hours after dosing.

Immediately following sacrifice bone marrow was aspirated from the femur. The marrow was washed and the cells were fixed before being spread on slides (at least 3 slides were prepared from each animal).  
 Slides were stained and scored without regard to treatment group. Where possible, a minimum of 50 metaphase cells from each animal were examined and scored for chromatid and chromosome gaps and breaks, fragments, structural rearrangements and ploidy.  
 The ratio of the number of cells in mitosis per 500 cells counted x 100 is defined as the mitotic index.

The data on chromosomal aberrations for the treated animals was compared to that for the negative controls.

**Result** : There was a 9% weight loss in males 48 hours after receiving 3 g/kg API 81-04 and a 2% weight loss in females at the same time and dose level. Clinical signs of toxicity in the 3 g/kg group included lethargy in both sexes and increased tearing as indicated by a crusty appearance of fur around the eyes of the male animals. Animals in the vehicle control group appeared normal.  
 The report includes for each animal the following information: number of cells scored, mitotic index, modal chromosome number, number of gaps, breaks (chromatid and chromosome) and fragment, number of rearrangements (exchange figures, dicentric and ring) and number of severely damaged cells.  
 Treatment with API 81-04 did not affect any of these parameters. There were no apparent sex differences and consequently the data for both

sexes are combined in the following summary table.

500 cells were examined for every treatment at each time period shown below.

No rearrangements were recorded for either the vehicle control or any of the groups treated with API 81-04, whereas 13 rearrangements were recorded for the positive control group.

No aberrations from severely damaged cells were recorded for either the vehicle control or API 81-04 groups but 80 were recorded for the positive control group.

Time (hrs)	Incidence of aberrations (%)	Total No. of		Aberrations per cell
		Gaps	Breaks	
<b>Cor n oil control (5 ml/kg)</b>				
6	0.8	5	4	0.008
24	0.4	3	2	0.004
48	0.4	2	2	0.004
<b>API 81-04 (3 g/kg)</b>				
6	1	3	5	0.010
24	0	5	0	0
48	0.4	1	2	0.004
<b>API 81-04 (1 g/kg)</b>				
6	0.6	3	3	0.006
24	0.4	2	2	0.004
48	0.4	0	2	0.004
<b>API 81-04 (0.3 g/kg)</b>				
6	1.0	1	5	0.010
24	0	2	0	0
48	0.2	3	1	0.002
<b>TEM 0.5 mg/kg)</b>				
24	16.6	22	126	0.438

The data above demonstrate that API 81-04 did not cause chromosomal aberrations in either male or female rats at the dose levels tested.

<b>Remark</b>	:	In a separate study in which exposure was by inhalation at 63, 297 and 2046 ppm, 6hr/day for 5 days, there was no evidence of a light catalytic cracked naphtha causing chromosomal aberrations in rats. (API Ref 32-31300)
<b>Reliability</b> 09.08.2002	:	(1) Valid without restriction (1) (3)
<b>Type</b>	:	Sister chromatid exchange assay
<b>Species</b>	:	Mouse
<b>Sex</b>	:	Male/female
<b>Strain</b>	:	B6C3F1
<b>Route of admin.</b>	:	i.p.
<b>Doses</b>	:	200, 1200 and 2400 mg/kg
<b>Result</b>	:	Positive
<b>Year</b>	:	1988
<b>GLP</b>	:	Yes
<b>Test substance</b>	:	API 81-03
<b>Method</b>	:	Dose levels were selected on the basis of a dose range finding study that had been conducted previously.

Six experimental groups of five male and five female mice were used for the SCE assay.

Four hours prior to administration of test material, the mice were

anesthetized with Metofane and an agar-coated 50 mg BrdUrd pellet was implanted subcutaneously in the lower abdominal region.

The test material in corn oil or the corn oil alone were administered by ip injection at a rate of 10 ml/kg body weight. The positive control (CP) was injected ip at a dose level of 10 mg/kg. The positive control (API 81-15) was administered at a dose of 4 g/kg, which was administered by ip injection at a rate of 10 ml/kg.

All mice were weighed immediately prior to administration of test dose. Colchicine, used to arrest dividing cells in metaphase, was administered ip at 1 mg/kg to all mice two to four hours prior to sacrifice.

24 to 26 hours after BrdUrd pellet implantation the mice were sacrificed. Marrow was collected from both femurs. After washing and fixing bone marrow cells slides were prepared for subsequent staining and examination. Two to five slides were prepared from each animal.

A minimum of 50 second-division metaphase spreads from each animal were examined and scored for SCEs and chromosome number. The mitotic index was recorded as the percentage number of cells in mitosis base upon 500 cells counted. The percentage of first, second and third-division metaphase cells was also recorded as the number per 100 cells counted.

A test article is considered to induce a positive response if a dose-related increase in SCEs/metaphase is observed relative to the vehicle control. The test is considered valid if the mean number of SCE per second division metaphase cell must not exceed 8 SCEs/cell/animal in the negative (vehicle) control. The mean SCE/cell/animal for the positive control animals must be statistically increased relative to the vehicle control using the Mann Whitney test ( $P = 0.05$ )

**Result** : There was little or no apparent weight loss between the pretreatment body weights and those at the time of colchicine dosing. No clinical signs of toxicity were observed. No mitotic delay or adverse effect on mitotic indices were observed at any test article dose level.

The results are summarized in the following table.

Sex	Range of mean SCEs/cell for individual animals		Average SCEs/cell per mouse <sup>(1,2)</sup>
<b>Corn oil</b>			
M	4.68	5.84	5.44±-0.47 (5.64)
F	5.28	7.36	6.25±-0.86 (6.06)
<b>API 81-03 (2.4 mg/kg)</b>			
M	7.06	10.46	8.88±-1.24 (9.14)**
F	8.16	11.58	9.61±-1.4 (9.12)**
<b>API 81-03 (1.2 mg/kg)</b>			
M	8.58	10.2	9.15±-0.65 (8.96)**
F	8.92	12.28	10.5±-1.49 (10.2)**
<b>API 81-03 (0.2 mg/kg)</b>			
M	7.54	9.28	8.52±-0.71 (8.86)**
F	9.2	11.44	10.0±-0.92 (10.14)**
<b>API 81-15 positive control (4 g/kg)</b>			
M	6.68	9.28	7.94±-0.93 (7.94)**
F	7.28	8.54	7.86±-0.58 (7.56)*

**CP (10 mg/kg)**

M	36.6	44.18	40.3±-3.53 (38.5)**
F	18.34	31.44	25.5±-5.38 (25.06)**

1 Mean± standard deviation (median SCEs/cell)  
2 \* P = 0.05 \*\* P = 0.01

There was a significant increase in SCEs/cell when analyzed by sex. Pairwise comparisons by sex of each treatment group with its vehicle control were significantly different. CP and 81-15 also caused an increase in SCEs/cell/mouse in both males and females.

The negative and positive controls fulfilled the requirements for determination of a valid test. Therefore, API 81-03 was shown to be positive in the SCE assay.

**Reliability**  
09.08.2002

: (1) Valid without restriction

(5)

- (1) American Petroleum Institute (1985)  
Activity of API 81-04 in the acute in vivo cytogenetics assay in male and female rats  
API Med. Res. Pub. 32-32288
- (2) American Petroleum Institute (1985)  
L5178Y TK +/- Mouse lymphoma mutagenesis assay of API 81-04  
API Med. Res. Pub. 32-31710
- (3) American Petroleum Institute (1985)  
Mutagenicity evaluation studies  
in the rat bone marrow cytogenetic assay  
in the mouse lymphoma forward mutation assay  
Light catalytic cracked naphtha API sample 81-03  
API Med. Res. Pub. 32-31300
- (4) American Petroleum Institute (1987)  
Mutagenicity of API 83-20, Light catalytic cracked naphtha (CAS 64741-55-5) in a mouse  
lymphoma mutation assay  
HESD Pub. No. 34 -30633
- (5) American Petroleum Institute (1988)  
In vivo sister chromatid exchange assay with API 81-03 (light catalytic cracked naphtha)  
HESD Publ. No. 36-30044
- (6) American Petroleum Institute (1988)  
Sister chromatid exchange assay in Chinese hamster ovary (CHO) cells with API 81 -03  
(Light catalytic cracked naphtha)  
HESD Pub. No. 36 -30045

7.1 END POINT SUMMARY

7.2 HAZARD SUMMARY

7.3 RISK ASSESSMENT